Synthesis of N-Acetylglucosaminyl- and N-Acetylgalactosaminylceramides as Cerebroside Analogs and Their Anti-human Immunodeficiency Virus Type 1 Activities

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Monoglycosylceramide derivatives containing mimicks of ceramide were synthesized as cerebroside analogs from D-glucosamine or D-galactosamine derivatives and N-benzyloxycarbonyl-L-serine myristylamide by using trimethylsilyl trifluoromethanesulfonate (TMSOTf) as a promoter. The synthesized sulfated glycolipids show moderate anti-HIV-1 activities.

Key words sulfated glycosylceramide; cerebroside analog; anti-HIV-1 activity

Galactosylceramide is an essential component of neural receptor for type 1 human immunodeficiency virus (HIV) surface glycoprotein gp120.¹⁾ Various galactosylceramide analogs have been synthesized and their biological activities were examined.²⁾ In preceding papers, we reported the synthesis of sulfated gangliosides³⁾ and sulfated cerebroside analogs⁴⁾ containing L-serine diamide derivatives as mimicks of the ceramide moieties of gangliosides showing anti-influenza virus activities and anti-HIV type 1 activities. N-Acetyl D-glucosamine and N-acetyl D-galactosamine are often found as constituents of glycoconjugates,⁵⁾ and they have various biological activities and functions.

As a part of our synthetic studies on biologically active new compounds designed by modifying natural glycoconjugates, we describe here the synthesis of sulfated monoglycosylceramides, as indicated in Chart 1, together with some results of biological testing.

Chart 2 shows the synthetic route to 5a and 5b. First, the neighboring-group-assisted coupling of 2-chloroacetamido-2-deoxy-1,3,4,6-tetra-O-acetyl- β -D-glucopyranose and 2-chloroacetamido-2-deoxy-1,3,4,6-tetra-O-acetyl- β -D-galactopyranose with an N-benzyloxycarbonyl-L-serine myristylamide derivative in the presence of Me₃SiOSO₂-CF₃ (TMSOTf) and molecular sieves 4 Å in ClCH₂CH₂Cl gave the desired glycosides (1a and 1b) in yields of 69 and 74%, respectively. The ¹H-NMR data for the anomeric proton H-1 [δ 4.71 ($J_{1,2}$ =6.8 Hz) in **1a**, δ 4.69 ($J_{1,2}$ = 7.0 Hz) in 1b] indicated the stereochemistry of the newly formed glycosidic bond to be β . Reduction of the benzyloxycarbonyl and chloroacetyl groups in 1a and 1b by hydrogenolysis over Pd-C in MeOH gave the alcohols 2a and 2b (quantitative and 93% yield, respectively). De-O-acetylation of 2a and 2b had to be done under mild conditions, due to base-sensitivity (retro-Michael reaction) of the O-serinyl glycosyl portion in particular. 6) The best results were achieved by treatment of 2a and 2b with triethylamine (TEA)–MeOH $(1:10)^{7}$) at room temperature to give the triols 3a and 3b (quantitative and 88% yield, respectively). During the de-O-acetylation, no β -elimination product was detected. Acylation of the free amino groups of 3a and 3b with stearoyl chloride and aqueous NaHCO₃ gave diacylated compounds 4a and 4b (87%

yield and quantitative, respectively). Finally, O-sulfation of 4a and 4b was achieved with sulfur trioxide-tripyridine complex in N,N-dimethylformamide (DMF). Removal of pyridine was readily accomplished by brief treatment with trifluoromethanesulfonic acid (TFA) in dichloromethane and the products were purified by chromatography on Sephadex LH-20 (CHCl₃: MeOH: H₂O=6:6:1) and lyophilized to afford the 3,4,6-tri-O-sulfated glycosides 5a and 5b (42 and 46% yields, respectively). Absorptions at 1215—1268 cm⁻¹ (due to S=O stretching) and 756—834 cm⁻¹ (due to C-O-S vibration) were observed in the infrared (IR) spectra of 5a and 5b, indicating the presence of sulfate esters. Furthermore, 5a and 5b gave a positive test with a specific spray-reagent (azure A reagent) for sulfated glycolipids.⁸⁾

The structures of all compounds were characterized by ¹H-NMR spectroscopy, as well as IR spectroscopy, elemental analyses, and positive FAB-mass spectrometry.

The anti-HIV-1 activities of the two nonsulfated glycosylceramides (4a and 4b) and the two sulfated glycosylceramides (5a and 5b) are shown in Table 1. The anti-HIV-1 activity was tested by the syncytium-formation

Chart 1

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Reagents: a) TMSOTf, ClCH₂CH₂Cl; b) H₂, Pd-C, MeOH; c) TEA-MeOH (1:10); d) CH₃(CH₂)₁₆COCl, NaHCO₃, ether-H₂O; e) i) SO₃-pyridine, DMF-pyridine (1:1); ii) TFA, CH₂Cl₂, then LH-20 (CHCl₃-MeOH-H₂O= 6:6:1)

Chart 2

Table 1. Results of Anti-HIV Assay by IFA Using MT-4 Cells

Compd. No.	HIV-1 infection (IC ₅₀) $(\mu g/ml)^{a}$	$CT^{b)}$
4 a	>100	(++)
4b	>100	(++)
5a	30—100	(-)
5b	30—100	(-)

a) Concentrations (μ g/ml) of compounds at which 50% of MT-4 cells expressed HIV-1 antigens. b) CT: cytotoxic (- to ++).

assay method using MT-4 cells according to our previously reported method. Among the synthesized compounds, the sulfated compounds ($\bf 5a$ and $\bf 5b$) showed moderate activities with 50%-inhibitory concentration (IC₅₀) values of 30—100 μ M, and they were noncytotoxic. The nonsulfated compounds ($\bf 4a$ and $\bf 4b$) were practically inactive (IC₅₀>100 μ M) against HIV-1 and were cytotoxic.

Experimental

All melting points are uncorrected. Optical rotations were measured with a JASCO DIP-140 digital polarimeter. IR spectra were recorded on a JASCO A-202 infrared spectrophotometer $^1\text{H-NMR}$ spectra were taken on a JEOL JNM-GX 270 (270 MHz) spectrometer with tetramethylsilane (in CDCl₃) as an internal standard, and the chemical shifts are given in δ values. The abbreviations of signal patterns are as follows: s, singlet; br s, broad singlet; d, doublet; t, triplet; q, quartet; m, multiplet. Mass spectra (MS) were recorded on a JEOL JMS-SX102

spectrometer. Column chromatography was carried out on Silica gel 60 (70—230 mesh, Merck). Thin-layer chromatography (TLC) on silica gel 60- F_{254} (Merck) was used to monitor the reaction and to ascertain the purity of the reaction products. The spots were visualized by spraying the plates with 5% aqueous sulfuric acid and then heating. Sulfated glycolipids were visualized with azure A reagent. The bands of lipids containing sulfate esters were stained blue.

N-Benzyloxycarbonyl-*O*-(3,4,6-tri-*O*-acetyl 2-chloroacetylamino-2-deoxy- β -D-glucopyranosyl)-L-serine Myristylamide (1a) A solution of 2-chloroacetamido-2-deoxy-1,3,4,6-tetra-*O*-acetyl- β -D-glucopyranose (0.85 g, 2 mmol) and *N*-benzyloxycarbonyl-L-serine myristylamide (0.90 g, 2 mmol) in anhydrous ClCH₂CH₂Cl (20 ml) was stirred for 1 h at room temperature under argon in the presence of powdered molecular sieves 4 Å. The mixture was cooled to 0 °C, then TMSOTf (0.45 g, 2 mmol) was added. The mixture was stirred for 15 h at room temperature, and the reaction mixture was filtered through Celite 545. The filtrate was washed with aqueous NaHCO₃ and H₂O, dried (MgSO₄), and concentrated *in vacuo*. The residual product was chromatographed on SiO₂ with CH₂Cl₂-CH₃COCH₃ (20:1) to give **1a** (1.10 g, 69%) as an amorphous powder. [α]_D +1.1° (c=0.90, CHCl₃). IR (Nujol): 1744 (ester), 1657, 1535 (amide) cm⁻¹.

¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J=7.0 Hz, -CH₃), 1.26 (22H, br s, -CH₂-), 1.42—1.49 (2H, m, NCH₂CH₂), 2.03, 2.04, 2.06 (each 3H, s, OAc), 3.23 (2H, t, J=6.5 Hz, NCH₂CH₂), 3.77 (1H, dd, J=10.5, 7.3 Hz, OCH₄H_bCHN), 3.86—3.98 (2H, m, H-2,5), 3.94 (2H, br s, NHCOCH₂Cl), 4.08 (1H, dd, J=10.5, 4.3 Hz, OCH₄H_bCHN), 4.15 (1H, dd, J=11.7, 2.2 Hz, H-6b), 4.24 (1H, dd, J=11.7, 4.9 Hz, H-6a), 4.34—4.36 (1H, m, OCH₂CHN), 4.71 (1H, d, J=6.8 Hz, H-1), 5.05 (1H, dd, J=9.5 Hz, H-4), 5.11 (2H, br s, CH₂Ph), 5.25 (1H, t, J=9.5 Hz, H-3), 5.68 (1H, br s, OCH₂CHNH), 6.37 (1H, br s, CONH), 6.61 (1H, d, J=8.9 Hz, NHCOCH₂Cl), 7.36 (5H, br s, Ph). *Anal.* Calcd for C₃₉H₆₀ClN₃O₁₂: C, 58.67;H, 7.58; N, 5.26. Found: C, 58.32; H, 7.61;

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O-(3,4,6-Tri-O-acetyl-2-acetylamino-2-deoxy-β-D-glucopyranosyl)-Lserine Myristylamide (2a) A mixture of 1a (0.98 g, 0.84 mmol) and 10% Pd-C (0.24 g) in MeOH (40 ml) was stirred under H₂ overnight at room temperature. The catalyst was removed by filtration and the filtrate was concentrated to dryness. The residue was chromatographed on SiO_2 with CH₂Cl₂-MeOH (10:1) to give **2a** (0.529 g, quant.), mp 125—128 °C. $[\alpha]_D - 14.3^\circ$ (c = 0.46, MeOH). IR (KBr): 3300 (NH), 1744 (ester), 1657, 1535 (amide) cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 7.3 Hz, $-\text{CH}_3$), 1.26 (22H, br s, $-CH_2$ -), 1.50 (2H, br s, $NCH_2C\underline{H}_2$), 1.95, 2.03, 2.09 (12H, s, OAc, NAc), 3.18—3.26 (2H, m, NCH₂CH₂), 3.54 (1H, dd, J=7.6, 4.9 Hz, OCH₂CHN), 3.64—3.70 (1H, m, H-5), 3.82 (1H, dd, J=10.3, 7.6 Hz, OCH_aH_bCHN), 3.90 (1H, dd, J=10.3, 4.9 Hz, OCH_aH_bCHN), 3.98 (1H, dd, J=8.4, 8.9 Hz, H-2), 4.13 (1H, dd, J=12.2, 2.2 Hz, H-6a), 4.25 (1H, dd, J=12.2, 4.6 Hz, H-6b), 4.59 (1H, d, $J=8.4 \,\mathrm{Hz}$, H-1), 5.07 (1H, t, $J=9.2 \,\mathrm{Hz}$, H-4), 5.17 (1H, t, $J=9.2 \,\mathrm{Hz}$, H-3), 5.71 (1H, d, J=8.9 Hz, AcNH). Anal. Calcd for $C_{31}H_{55}N_3O_{10}$: C, 59.12; H, 8.80; N, 6.67. Found: C, 59.10; H, 8.90; N, 6.70.

O-(2-Acetylamino-2-deoxy-β-D-glucopyranosyl)-L-serine Myristylamide (3a) A solution of 1b (0.55 g, 0.87 mmol) in NEt₃-MeOH (1:10) (30 ml) was stirred at room temperature for 5 h, then concentrated to dryness under reduced pressure. The residue was purified by column chromatography with CH₂Cl₂-MeOH (3:1) to give 3a (0.44 g, quant.), mp 188—191 °C. [α]_D – 14.5° (c=0.36, CHCl₃-MeOH (1:1)). IR (KBr): 3300 (NH), 1646, 1557 (amide) cm⁻¹. ¹H-NMR (CDCl₃: CD₃OD=10:1) δ: 0.88 (3H, t, J=7.0 Hz, -CH₃), 1.26 (22H, br s, -CH₂-), 1.44—1.51 (2H, m, NCH₂CH₂), 2.01 (3H, s, AcN), 3.20 (2H, t, J=6.8 Hz, NCH₂CH₂), 3.29—3.41 (4H, m, NCH₂CHN, H-5, 6), 3.44 (1H, t, J=8.4 Hz, H-3), 3.61 (1H, t, J=8.4 Hz, H-2), 3.73 (1H, dd, J=10.3, 4.9 Hz, OCH₃H_bCHN), 3.87 (1H, dd, J=10.3, 2.7 Hz, OCH₃H_bCHN), 4.40 (1H, d, J=8.4 Hz, H-1). *Anal*. Calcd for C₂₅H₄₉N₃O₇·H₂O: C, 57.56; H, 9.85; N, 8.05. Found: C, 58.04; H, 9.58; N, 7.55. Positive FAB-MS m/z: 503 (M+1)+.

N-Stearoyl-*O*-(2-acetylamino-2-deoxy-β-D-glucopyranosyl)-L-serine Myristylamide (4a) A solution of stearoyl chloride (0.112 g, 0.36 mmol) in ether (10 ml) was added to a solution of **3a** (0.171 g, 0.36 mmol) and saturated aqueous NaHCO₃ (50 ml) at 0 °C. The mixture was stirred for 4 h. The resulting precipitates was washed with ether and dried *in vacuo* to give **4a** (0.24 g, 87%), mp 229—231 °C, as an amorphous powder. IR (KBr): 3280 (OH, NH), 1654, 1542 (amide) cm⁻¹. ¹H-NMR (DMSO- d_6) δ: 0.86 (6H, t, J=6.5 Hz, -CH₃), 1.24 (52H, br s, -CH₂-), 1.40 (2H, br s, COCH₂CH₂), 1.49 (2H, br s, NCH₂CH₂), 1.83 (3H, s, AcNH), 4.34 (1H, d, J=7.3 Hz, H-1), 4.48 (1H, br s, NH), 4.89 (1H, br s, NH). *Anal.* Calcd for C₄₃H₈₃N₃O₈·5H₂O: C, 60.04; H, 10.90; N, 4.88. Found: C, 60.20; H, 10.96; N, 4.11. Positive FAB-MS m/z: 771 (M+1)⁺, 793 (M+Na)⁺.

N-Stearoyl-O-(2-acetylamino-2-deoxy-3,4,6-tri-O-sulfo-β-D-glucopy-ranosyl)-L-serine Myristylamide (5a) A solution of 4a (0.093 g, 0.12 mmol) in DMF (4 ml) was stirred for 20 h at 40—50 °C in the presence of sulfur trioxide–pyridine complex (0.086 g, 0.54 mmol). The mixture was cooled and chromatographed on a column of Sephadex LH-20 equilibrated in 1:1 (v/v) CHCl₃-MeOH. Elution with the same solvent gave a residue that was dissolved in CH₂Cl₂ (4 ml). The solution was treated with CF₃SO₃H (0.062 g, 0.54 mmol) for 3 h under ice-cooling, then the solvent was evaporated in vacuo. The residue was dissolved in H₂O (1 ml) and chromatographed on a column of Sephadex LH-20. Elution with 6:6:1 (v/v/) CHCl₃-MeOH-H₂O afforded 5a (0.052 g, 42%) as an amorphous powder, after lyophilization from H₂O, mp 165—167 °C. IR (KBr): 1652, 1542 (amide), 1251 (S=O), 816 cm⁻¹ (C-O-S). Anal. Calcd for C₄₈H₈₃N₃O₁₇S₃·2H₂O: C, 52.11; H, 7.93; N, 3.80. Found: C, 52.33; H, 7.71; N, 3.75.

N-Benzyloxycarbonyl-*O*-(3,4,6-tri-*O*-acetyl-2-chloroacetylamino-2-deoxy-β-D-galactopyranosyl)-L-serine Myristylamide (1b) The same procedure as described for the preparation of 1a provided a crude product from 2-chloroacetamido-2-deoxy-1,3,4,6-tetra-*O*-acetyl-β-D-galactopyranose (0.425 g, 1.0 mmol), *N*-benzyloxycarbonyl-L-serine myristylamide (0.455 g, 1.0 mmol) and TMSOTf (0.222 g, 1.0 mmol) and this was purified by column chromatography (elution with 20:1 CHCl₃–CH₃COCH₃) to give 1b (0.59 g, 74%) as an amorphous powder. [α]_D +2.3° (c=1.30, CHCl₃). IR (KBr): 1744 (ester), 1657, 1535 (amide) cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J=6.8 Hz, -CH₃), 1.25 (22H, br s, -CH₂-), 1.42—1.49 (2H, m, NCH₂CH₂), 2.03, 2.04, 2.14 (each 3H, s, AcO), 3.23 (2H, t, J=5.9 Hz, NCH₂CH₂), 3.78 (1H, dd, J=10.5, 7.0 Hz, OCH_aH_bCHN), 3.95 (2H, br s, NHCOCH₂Cl), 4.01—4.18 (3H, m, H-2, 5, OCH_aH_bCHN), 4.34 (1H, dd, J=4.9, 7.0 Hz, OCH₂CH₂N), 4.69 (1H, d, J=7.0 Hz, H-1), 5.11 (2H, s, OCH₂Ph), 5.20 (1H, dd, J=3.0, 8.1 Hz,

H-3), 5.36 (1H, br d, J=3.0 Hz, H-4), 5.71 (1H, br s, OCH₂CHN $\underline{\text{H}}$), 6.38 (1H, br s, CONH), 6.54 (1H, d, J=8.6 Hz, N $\underline{\text{H}}$ COCH₂Cl), 7.35 (5H, br s, Ph). *Anal*. Calcd for C₃₉H₆₀ClN₃O₁₂: C, 58.67; H, 7.58;N, 5.26. Found: C, 58.75; H, 7.77; N, 5.06.

O-(3,4,6-Tri-O-acetyl-2-acetylamino-2-deoxy-β-D-glucopyranosyl)-Lserine Myristylamide (2b) The same procedure as described for the preparation of 2a provided a crude product from 1b (0.59 g, 0.74 mmol), Pd-on-charcoal (0.25 g) and MeOH (15 ml), and this was purified by column chromatography (elution with 10:1 CH₂Cl₂-MeOH) to give **2b** (0.471 g, 93%) as an amorphous powder, mp 110—113 °C. $[\alpha]_D$ -12.0° (c=0.99, MeOH). IR (KBr): 3354 (NH, OH), 1745 (ester), 1656, 1527 (amide) cm⁻¹. 1 H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 7.0 Hz, -CH₃), 1.26 (22H, br s, -CH₂-), 1.51 (2H, br s, NCH₂CH₂), 1.96, 2.01, 2.05, 2.15 (each 3H, s, OAc, NAc), 3.18—3.27 (2H, m, NCH₂CH₂), 3.56 (1H, dd, J = 7.6, 4.6 Hz, OCH₂CHNH), 3.83 (1 H, dd, J = 10.0, 7.6 Hz, H-6a), 3.90 (1H, dd, J = 10.0, 4.9 Hz, H-6b), 4.04-4.08 (1H, m, H-5), 4.12-4.22(1H, m, H-2), 4.62 (1H, d, J=8.4 Hz, H-1), 5.15 (1H, dd, J=11.3, 3.5 Hz, H-3), 5.35 (1H, br d, J = 3.5 Hz, H-4), 5.70 (1H, br d, J = 8.6 Hz, NHAc). Anal. Calcd for C₃₁H₅₅N₃O₁₀: C, 59.12; H, 8.80; N, 6.67. Found: C, 58.67; H, 8.78; N, 6.65.

O-(2-Acetylamino-2-deoxy-β-D-glucopyranosyl)-L-serine Myristylamide (3b) The same procedure as described for the preparation of 3a provided a crude product from 2b (0.25 g, 0.50 mmol) and NEt₃–MeOH (1:10) (20 ml), and this was purified by column chromatography (elution with 3:1 CH₂Cl₂–MeOH) to give 3b (0.15 g, 88%) as an amorphous powder, mp 160–162 °C. [α]_D – 7.6° (c=1:1 CHCl₃–MeOH). IR (KBr): 3282 (NH, OH), 1648, 1560 (amide) cm^{-1.1}H-NMR (CDCl₃: CD₃OD=10:1) δ: 0.88 (3H, t, J=7.0 Hz, -CH₃), 1.27 (22H, br s, -CH₂–), 1.51 (2H, br s, NCH₂CH₂), 2.02 (3H, s, AcN), 3.20 (2H, t, J=7.0 Hz, NCH₂CH₂), 3.49 (1H, dd, J=11.3, 4.9 Hz, H-6a), 3.56 (1H, dd, J=10.5, 3.5 Hz, H-3), 3.71—3.75 (1H, m, H-5), 3.77 (1H, dd, J=4.6, 3.5 Hz, H-4), 3.85 (1H, t, J=10.5 Hz, H-2), 3.84 (1H, dd, J=11.3, 3.2 Hz, H-6b), 4.37 (1H, d, J=8.1 Hz, H-1). *Anal.* Calcd for C₂₅H₄₉N₃O₇: C, 59.62; H, 9.81; N, 8.34. Found: C, 59.32; H, 9.81; N, 8.13.

N-Stearoyl-*O*-(2-acetylamino-2-deoxy-β-D-glucopyranosyl)-L-serine Myristylamide (4b) The same procedure as described for the preparation of 4a provided a crude product from 3b (0.12 g, 0.25 mmol), stearoyl chloride (0.078 g, 0.25 mmol) and saturated aqueous NaHCO₃ (25 ml), and this was washed with ether to give 3b (0.19 g, quant.) as an amorphous powder, mp 219—222 °C. IR (KBr): 3276 (OH), 1639, 1559 (amide) cm⁻¹. ¹H-NMR (DMSO- d_6) δ: 0.86 (6H, t, J=5.9 Hz, -CH₃), 1.24 (52H, br s, -CH₂-, 1.40 (2H, br s, COCH₂CH₂), 1.49 (2H, br s, NCH₂CH₂), 1.84 (3H, s, AcNH). *Anal*. Calcd for C₄₃H₈₃N₃O₈·5H₂O: C, 60.04; H, 10.90; N, 4.88. Found: C, 60.07; H, 10.39; N, 4.23.

N-Stearoyl-*O*-(2-acetylamino-2-deoxy-3,4,6-tri-*O*-sulfo-β-D-glucopyranosyl)-L-serine Myristylamide (5b) The same procedure as described for the preparation of **5a** provided a crude product from **4b** (0.070 g, 0.09 mmol) and sulfur trioxide-pyridine complex (0.065 g, 0.41 mmol), followed by TFA (0.047 g, 0.41 mmol), and this was purified on a column of Sephadex LH-20 equilibrated in CHCl₃–MeOH–H₂O (6:6:1, v/v) to give **9** (0.042 g, 46%) as an amorphous powder. mp 154—157 °C. IR (KBr): 1653, 1543 (amide), 1250 (S=O), 812 (C–O–C) cm⁻¹. *Anal.* Calcd for $C_{48}H_{83}N_3O_{17}S_3 \cdot H_2O$: C, 52.97; H, 7.87; N, 3.86. Found: C, 53.17; H, 8.26; N, 3.81.

References

- a) Harouse J. M., Bhat S., Spitalnik. , Laughlin M., Stefano K., Silberberg D. H., Gonzalez-Scarano F., Science, 253, 320—323 (1991); b) Bhat S., Spitalnik S. L., Gonzalez-Scarano F., Silberberg D. H., Proc. Natl. Acad. Sci. U.S.A., 88, 7131—7134 (1991).
- a) Morita M., Natori T., Akimoto K., Osawa T., Fukushima H., Koezuka Y., Bioorg. Med. Chem. Lett., 5, 699—704 (1995); b) Motoki K., Morita M., Kobayashi E., Uchida T., Akimoto K., Fukushima H., Koezuka Y., Biol. Pharm. Bull., 18, 1487—1491 (1995); c) Kobayashi E., Motoki K., Natori T., Uchida T., Fukushima H., Koezuka Y., ibid., 19, 350—353 (1996); d) Motoki K., Morita M., Kobayashi E., Uchida T., Fukushima H., Koezuka Y., ibid., 19, 952—955 (1996); e) Clary L., Greiner J., Santaella C., Vierling P., Tetrahedron Lett., 36, 539—542 (1995).
- a) Yoshida H., Ikeda K., Achiwa K., Hoshino H., Chem. Pharm. Bull., 43, 594—602 (1995);
 b) Handa A., Hoshino H., Nakajima K., Adachi M., Ikeda K., Achiwa K., Ito T., Suzuki Y., Biochem. Biophys. Res. Commun., 175, 1—9 (1991).
- Nagao Y., Nekado T., Ikeda K., Achiwa K., Chem. Pharm. Bull.,

- **43**, 1536—1542 (1995).
- a) Schmidt R. R., Kinzy W., Adv. Carbohydr. Chem. Biochem., 50, 21—123 (1994); b) Garg K., von dem Bruch K., Kunz H., ibid.,
 50, 277—310 (1994).
 6) Polt R., Szabo L., Treiberg J., Li Y., Hruby V. J., J. Am. Chem.
- Soc., 114, 10249—10258 (1992).
- 7) Derevitskaya V. A., Vafina M. G., Kochetkov N. K., Carbohydr. Res., 3, 377-388 (1967).
- Iida N., Toida T., Kushi Y., Handa S., Fredman P., Svennerholm L., Ishizuka I., J. Biol. Chem., 264, 5974—5980 (1989).