

Synthetic Studies on Glycopeptides Concerned with Defense Response of Plants. I. Syntheses of Suppressins A and B

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Two glycopeptides, suppressins A and B, that suppress the production of pisatin, a phytoalexin of pea, were synthesized. In the synthesis of suppressin A, condensation of 3,4,6-tri-*O*-acetyl-2-azido-2-deoxy- α -D-galactopyranosyl trichloroacetimidate or its glycosidic β isomer with *N*-(carbobenzoxy)-L-seryl-*O*-benzyl-L-seryl-glycine methyl ester was carried out in the presence of trimethylsilyl trifluoromethanesulfonate (TMSOTf) to give the monoglycosyl tripeptide derivatives. For the synthesis of suppressin B, glycosylation of 2,3,4,6-tetra-*O*-acetyl- α -D-galactopyranosyl bromide and 1,2,3,6-tetra-*O*-benzoyl- α -D-galactopyranose was promoted by silver trifluoromethanesulfonate (AgOTf) to provide a disaccharide derivative. The coupling of diglycosyl imidate, 2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-3,6-di-*O*-benzoyl-2-azido-2-deoxy-D-galactopyranosyl trichloroacetimidate, and *N*-(carbobenzoxy)-L-seryl-*O*-benzyl-L-seryl-glycyl-4-benzyl-L-aspartyl-5-benzyl-L-glutamyl-*O*-benzyl-L-threonine methyl ester in the presence of TMSOTf afforded the diglycosyl hexapeptide derivatives. Reduction, followed by *N*-acetylation, and then removal of the remaining protecting groups afforded the desired suppressin B.

Key words defense response; glycopeptide; *Mycosphaerella pinodes*; suppressor; suppressin A, B; synthesis

A pea pathogen, *Mycosphaerella pinodes*, secretes an elicitor and a suppressor of the defense responses of pea in its pycnosporangium germination fluid.¹⁾ Various plants synthesize antimicrobial substances, called phytoalexins, as a defense mechanism against invasive microorganisms.²⁾ The elicitor induces active defense reactions, such as the production of a major phytoalexin, pisatin. On the other hand, factors that can suppress the defense responses of plants that follow an attack by microorganisms, tentatively called suppressors, have been found in the culture filtrates and spore germination fluids of fungal pathogens. Shiraishi *et al.* characterized the chemical structures and some aspects of the biological activities of two suppressors,³⁾ suppressins A and B, purified from the spore germination fluid of a pea pathogen, *Mycosphaerella pinodes*. As shown in Fig. 1, both suppressors were found to be mucin-type glycopeptides composed of *N*-acetylgalactosamine and galactose as the carbohydrate moiety, and aspartic acid,

glutamic acid, glycine, serine and threonine as the peptide moiety. These suppressors inhibit both the ATPase activity⁴⁾ and polyphosphoinositide metabolism⁵⁾ in pea plasma membranes, causing the temporary suppression of the signal-transduction pathway that leads to the expression of defense genes, which encode key enzymes in the biosynthetic pathway to phytoalexin.

In order to investigate the structural requirements for bioactive glycopeptides in detail, we have carried out synthetic studies and we present here syntheses of suppressin A, suppressin B and their glycosidic β isomers.

Results and Discussion

The dipeptides, *N*-(*tert*-butoxycarbonyl)-*O*-benzyl-L-seryl-glycine methyl ester (**7**) and *N*-(*tert*-butoxycarbonyl)-5-benzyl-L-glutamyl-*O*-benzyl-L-threonine methyl ester (**8**) were synthesized by coupling of *N*-(*tert*-butoxycarbonyl)-*O*-benzyl-L-serine (**2**) with glycine methyl ester hydro-

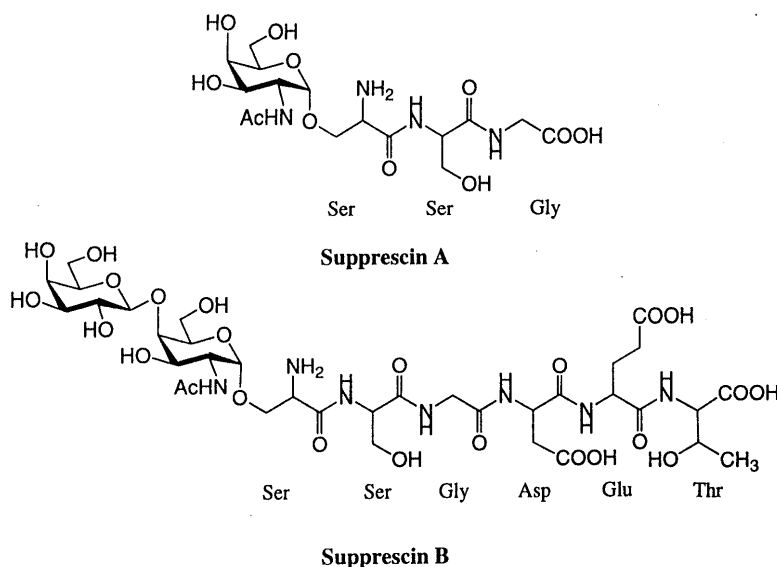


Fig. 1

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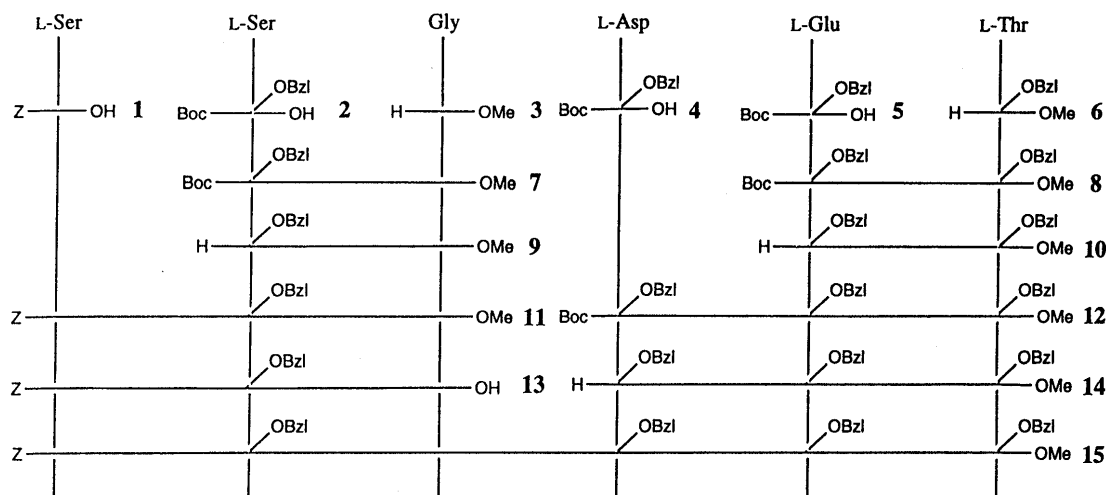


Chart 1

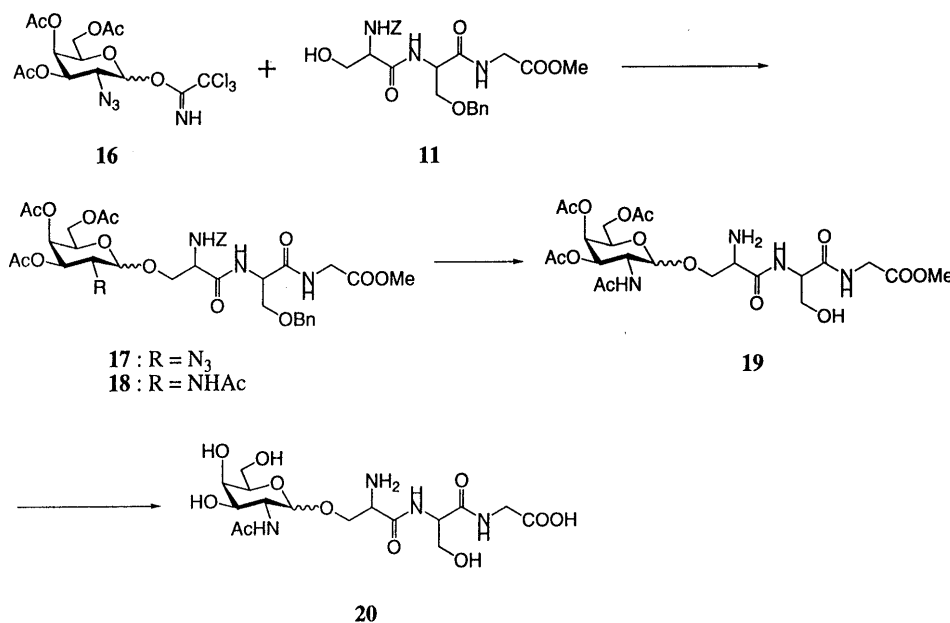


Chart 2

chloride (3) and *N*-(*tert*-butoxycarbonyl)-L-glutamic acid 5-benzyl ester (5) with *O*-benzyl-L-threonine methyl ester hydrochloride (6), respectively. Removal of the *tert*-butoxycarbonyl groups of 7 and 8 with trifluoroacetic acid gave compounds 9 and 10. Combination of the dipeptide 9 with *N*-(carbobenzoxy)-L-serine (1), and also the dipeptide 10 with *N*-(*tert*-butoxycarbonyl)-*O*-benzyl-L-aspartic acid 4-benzyl ester (4) in the presence of diethylphosphorocyanidate (DEPC)⁶ afforded *N*-(carbobenzoxy)-L-seryl-*O*-benzyl-L-seryl-glycine methyl ester (11) and *N*-(*tert*-butoxycarbonyl)-4-benzyl-L-aspartyl-5-benzyl-L-glutamyl-*O*-benzyl-L-threonine methyl ester (12) in 34% and 69% yields, respectively. The ¹H- and ¹³C-NMR spectra of these tripeptide derivatives were in accordance with the proposed structures. Removal of the methyl ester group of 11 with NaOMe in aqueous MeOH gave the C-terminus-free derivative 13 (94%). A brief treatment of 12 with trifluoroacetic acid provided the N-terminus-free derivative 14 in 84% yield. Combination of 13 with 14 in the presence of *N*-ethoxycarbonyl-2-ethoxy-1,2-dihydro-

quinoline (EEDQ)⁷ in dichloromethane gave the hexapeptide 15 in 62% yield (Chart 1). The ¹H-NMR spectrum of this hexapeptide showed aromatic proton signals at δ 7.31—7.23 ppm (m, 25 H), and the signal of the methyl ester appeared at δ 3.62 ppm.

In the synthesis of suppressin A, condensation of 3,4,6-tri-*O*-acetyl-2-azido-2-deoxy- α -D-galactopyranosyl trichloroacetimidate⁸ (16 α) and/or its β -isomer (16 β) with the tripeptide 11 was carried out in the presence of TMSOTf in dichloromethane at -30°C to give a 63% yield of a mixture of α and β glycopeptides 17. Sodium borohydride-nickel chloride reduction of the azido group in 17, followed by *N*-acetylation,⁹ gave compound 18. The α (18 α) and β (18 β) anomers of 18 could be separated by silica gel chromatography. The benzyl ester, benzyl ether and carbobenzoxy (Z) groups were cleaved from 18 α and 18 β by hydrogenolysis (Pd-C) to give 19 α and 19 β in 79% and 82% yields, respectively. Removal of the ester groups (acetyl and methyl ester) of 19 α and 19 β with NaOMe in aqueous MeOH afforded the desired 2-acet-

amido-2-deoxy-D-galactopyranosyl tripeptide **20 α** (suppressin A) and **20 β** in 60% and 80% yields, respectively (Chart 2). Deblocking of **19 α** and **19 β** with NaOMe in MeOH did not promote β -elimination of the *O*-glycosidic linkage bearing an L-seryl residue. Compounds **20 α** and **20 β** showed an $[M+H]^+$ ion peak at m/z

Table 1. ^{13}C -NMR Data (δ) for Compounds **18 α** , **18 β** , **19 α** , **19 β** , **20 α** , **20 β**

Carbon atom	Compound					
	18α	18β	19α	19β	20α	20β
C-1	98.5	101.7	100.2	101.9	101.7	104.0
2	47.5	51.2	48.5	50.7	52.2	54.9
3	67.9	69.8	68.6	70.2	71.2	71.3
4	67.2	69.1	68.3	66.7	70.3	70.4
5	67.1	66.7	68.3	70.8	69.6	73.5
6	61.9	61.6	62.8	61.5	64.0	63.7
Ser- α	53.9	54.3	56.9	55.2	58.3	58.4
	52.4	53.0	54.4	54.8	55.8	56.4
β	69.3	71.2	69.8	72.1	74.2	78.0
	67.9	69.6	63.1	62.6	64.2	64.0
Gly	41.4	41.4	41.3	41.3	46.1	46.1
OMe	52.4	52.3	52.4	52.5		
Z-CH ₂	67.3	67.2				
Bn-CH ₂	73.6	73.5				

453 in the FAB-MS. The structures and purity of **20 α** and **20 β** were established by ^1H -, ^{13}C -NMR (Table 1) spectroscopy and FAB-MS spectrometry.

Synthesis of suppressin B (**32 α**) and its glycosidic isomer (**32 β**) was carried out by the coupling of diglycosyl imidate **28** and the hexapeptide acceptor **15**. Compound **28** was prepared by employing readily available acetobromogalactose¹⁰⁾ **21** as a glycosyl donor and 1,2,3,6-tetra-*O*-benzoyl- α -D-galactopyranose¹¹⁾ **22** as a glycosyl acceptor. Compound **21** was coupled with **22** in the presence of silver trifluoromethanesulfonate (AgOTf) as a promoter, and 2,4,6-trimethylpyridine (collidine) as a neutralizing agent¹²⁾ of the liberated acid in toluene to afford **23** in 56% yield. The anomeric configuration of compound **23** was confirmed by ^1H -NMR spectroscopy, the signals for H-1 and H-1' being observed at δ 6.76 ($J=3.7\text{ Hz}$) and 4.75 ($J=7.9\text{ Hz}$), respectively. A heteronuclear multiple-bond correlation (HMBC) experiment¹³⁾ showed a correlation between H-1' (4.75 ppm) and the C-4-carbon (70.9 ppm). The ^{13}C -NMR data were in accordance with the proposed structure (see Table 2). Treatment of **23** with hydrogen bromide in acetic acid gave an α -bromide **24**. Elimination of the bromine and the 2-benzoyloxy group was achieved with zinc-copper reagent in acetate buffer to

Table 2. ^{13}C -NMR Data (δ) for Selected Compounds

Carbon atom	Compound									
	23	24	25	26α	26β	27α	27β	28α	28β	
C-1	90.9	89.0	145.8	97.4	98.1	92.7	96.5	94.9	96.7	
2	67.3	68.0	97.9	57.0	58.6	58.9	63.2	58.0	61.0	
3	70.8	73.2	64.5	71.3	71.1	70.7	73.4	71.2	73.3	
4	70.9	71.2	70.2	71.0	70.8	69.3	69.3	71.0	72.9	
5	73.9	73.5	74.1	73.7	74.4	74.8	73.4	74.1	73.6	
6	63.5	63.3	62.7	63.4	63.4	64.0	63.9	63.9	63.2	
C-1'	100.2	101.2	100.3	101.4	101.3	101.4	101.2	101.4	101.2	
2'	68.9	68.9	68.7	69.3	68.9	68.3	68.3	69.2	69.2	
3'	70.7	70.7	70.7	70.5	70.7	70.7	70.7	70.6	70.7	
4'	66.9	66.9	67.0	66.7	66.9	66.9	67.1	66.8	66.8	
5'	70.9	70.9	71.0	70.9	70.8	70.7	72.4	70.9	70.9	
6'	61.1	61.1	61.2	61.1	61.1	61.1	61.1	61.9	61.4	
OC(NH) CCl ₃								160.5	160.5	
								90.7	90.7	

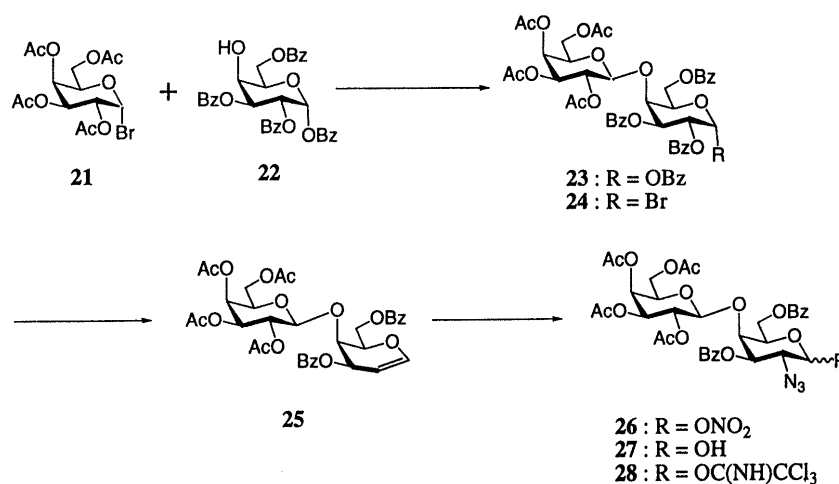


Chart 3

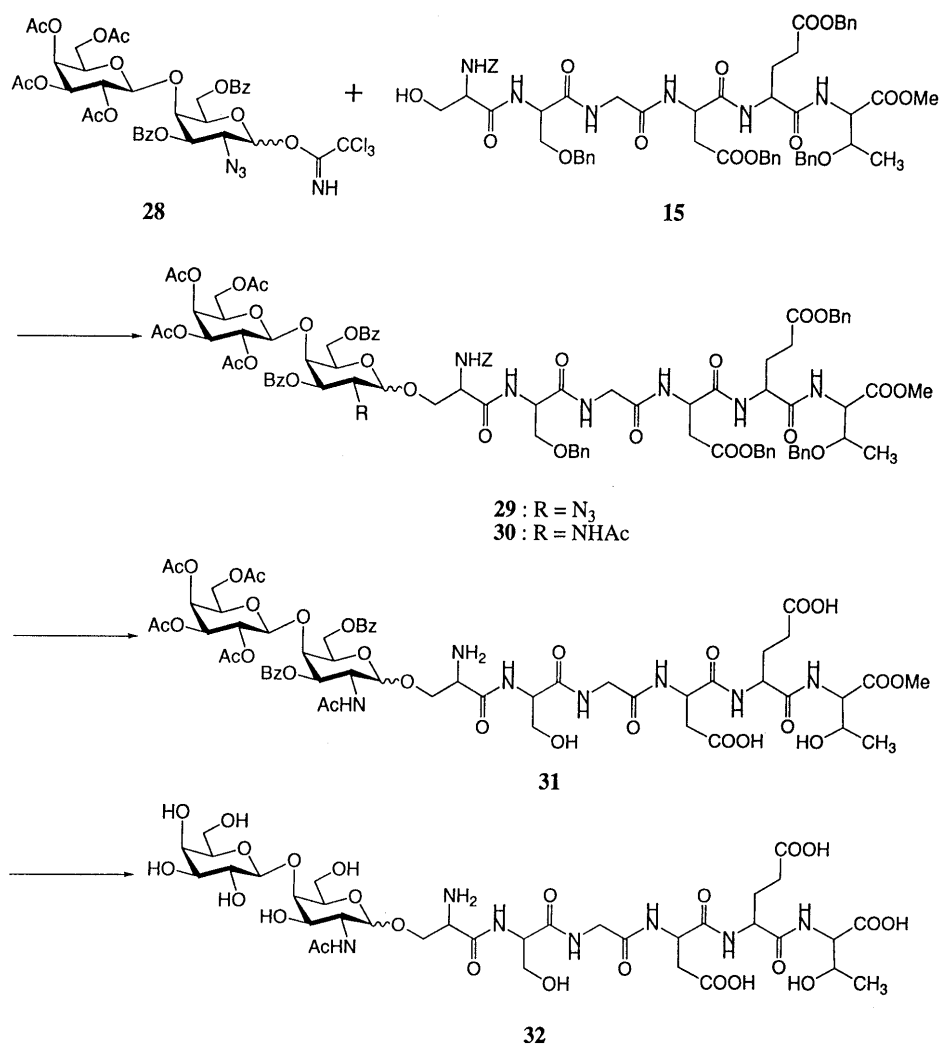


Chart 4

provide a galactal derivative **25**. The azidonitration¹⁴⁾ of compound **25** with NaN₃ and cerium(IV) ammonium nitrate (CAN) in CH₃CN gave a mixture of α and β nitrates **26**. Removal of the nitrate group was achieved with NaNO₂ in 1,4-dioxane at 80 °C, and the resultant alcohol **27** was transformed into the trichloroacetimidate **28** (Chart 3). Condensation of the diglycosyl donor **28** with the hexapeptide acceptor **15** in the presence of TMSOTf in dichloromethane at -30 °C gave the diglycosyl hexapeptide derivatives **29 α** (31%) and **29 β** (22%). Reduction of **29 α** and **29 β** , followed by *N*-acetylation afforded compounds **30 α** and **30 β** in 86% and 93% yields, respectively. The benzyl ester, benzyl ether and Z group were cleaved from **30 α** and **30 β** by hydrogenolysis (Pd-C) to give **31 α** and **31 β** in 66% and 68% yields, respectively. Removal of the acetyl, benzoyl and methyl ester groups of **31 α** or **31 β** with NaOMe in aqueous MeOH gave **32 α** (suppressin B) or **32 β** , respectively, each in 71% yield (Chart 4).

Experimental

General Methods Melting points were measured with a Yanagimoto micro melting-point apparatus and are uncorrected. Optical rotations were determined with a Jasco DIP-140 digital polarimeter. ¹H-NMR and ¹³C-NMR spectra were recorded with JEOL JNM EX-270 and JNM A 500 FT NMR spectrometers, Me₄Si was the internal standard for solutions in CDCl₃ and/or CD₃OD, and sodium 4,4-dimethyl-4-

silapentane-1-sulfonate for solutions in D₂O. FAB-MS was recorded on a JEOL JMS SX 102 mass spectrometer. TLC was performed on Silica gel-60F₂₅₄ (E. Merck) with detection by quenching of UV fluorescence and by spraying with either 10% H₂SO₄ or 5% methanolic ninhydrin solution. Column chromatography was carried out on Silica Gel-60 (E. Merck).

***N*-(*tert*-Butoxycarbonyl)-*O*-benzyl-L-seryl-glycine Methyl Ester (**7**)** A solution of *N*-(*tert*-butoxycarbonyl)-*O*-benzyl-L-serine (**2**) (45 g, 150 mmol) and glycine methyl ester hydrochloride (**3**) (26 g, 210 mmol) was prepared in 6:1:1 CH₂Cl₂-DMF-THF (300 ml), then Et₃N (26 ml, 190 mmol) and diethylphosphorocyanidate (DEPC) (30 ml, 200 mmol) were added and the mixture was held at 0 °C for 1 h. It was allowed to warm to room temperature for 12 h, then diluted with CHCl₃, and the CHCl₃ solution was washed with water, dried over Na₂SO₄, filtered, and concentrated. The residue was purified by column chromatography with CHCl₃-MeOH (100:1) to give **7** (34 g, 44%). *R*_f 0.68 (20:1 CHCl₃-MeOH), 0.58 (2:1 benzene-acetone); [α]_D²⁵ +21.0° (*c*=0.5, CHCl₃). ¹H-NMR (CDCl₃) δ : 7.36-7.27 (m, 5 H, Ar), 7.04 (s, 1H, NH), 5.44 (s, 1H, NH), 4.56 (dd, 2H, *J*=11.6, 25.0 Hz, Bn-CH₂), 4.35 (brs, 1H, Ser α), 4.05 (d, 2H, *J*=5.5 Hz, Gly), 3.91 (d, 1H, *J*=6.4 Hz, Ser β -Ha), 3.74 (s, 3H, OMe), 3.61 (dd, 1H, *J*=6.4, 9.4 Hz, Ser β -Hb), 1.45 (s, 9H, 3 \times *tert*-Bu-CH₃). ¹³C-NMR (CDCl₃) δ : 53.9 (Ser α), 69.8 (Ser β), 41.3 (Gly), 52.3 (OMe), 28.3 (*tert*-Bu-CH₃), 80.3 (*tert*-Bu-C), 73.5 (Bn-CH₂). *Anal.* Calcd for C₁₈H₂₆N₂O₆: C, 59.00; H, 7.15; N, 7.65. Found: C, 58.63; H, 7.08; N, 7.57.

***N*-(*tert*-Butoxycarbonyl)-5-benzyl-L-glutamyl-*O*-benzyl-L-threonine Methyl Ester (**8**)** A solution of *N*-(*tert*-butoxycarbonyl)-*O*-benzyl-L-glutamic acid 5-benzyl ester (**5**) (2.7 g, 8.0 mmol) and *O*-benzyl-L-threonine methyl ester hydrochloride (**6**) (1.0 g, 3.9 mmol) was prepared in CH₂Cl₂ (15 ml), then Et₃N (0.6 ml, 4.2 mmol) and DEPC (1.4 ml, 9.3 mmol) were added and the mixture was held at 0 °C for 12 h. It was

diluted with CHCl_3 , and the CHCl_3 solution was washed with water, dried over Na_2SO_4 , filtered, and concentrated. The residue was purified by column chromatography with CHCl_3 -MeOH (50:1) to give **8** (1.8 g, 88%). *Rf* 0.65 (4:1 benzene-acetone); $[\alpha]_{\text{D}}^{25} + 2.0^\circ$ ($c=2.0$, CHCl_3). $^1\text{H-NMR}$ (CDCl_3) δ : 7.32–7.22 (m, 10H, Ar), 5.08 (s, 2H, Bn- CH_2), 4.64 (dd, 1H, $J=2.4$, 9.2 Hz, Thr α), 4.54, 4.35 (each d, 2H, $J=11.8$ Hz, Bn- CH_2), 4.33 (br s, 1H, Glu α), 4.13 (m, 1H, Thr β), 3.59 (s, 3H, OMe), 2.50 (br s, 2H, Glu γ), 2.17, 1.98 (each m, 2H, Glu β), 1.41 (s, 9H, 3 \times *tert*-Bu- CH_3), 1.18 (d, 3H, $J=6.1$ Hz, Thr γ). $^{13}\text{C-NMR}$ (CDCl_3) δ : 53.6 (Glu α), 27.9 (Glu β), 30.4 (Glu γ), 56.7 (Thr α), 73.9 (Thr β), 16.0 (Thr γ), 52.2 (OMe), 28.3 (*tert*-Bu- CH_3), 79.7 (*tert*-Bu-C), 70.6, 66.3 (Bn- CH_2). *Anal.* Calcd for $\text{C}_{29}\text{H}_{38}\text{N}_2\text{O}_8$: C, 64.19; H, 7.06; N, 5.16. Found: C, 64.00; H, 7.23; N, 5.46.

O-Benzyl-L-seryl-glycine Methyl Ester (9) A solution of compound **7** (600 mg, 1.6 mmol) in trifluoroacetic acid (2.0 ml) was stirred for 1 h at room temperature, then diluted with CHCl_3 , and the CHCl_3 solution was washed with water, dried over Na_2SO_4 , filtered, and concentrated. The residue was chromatographed on silica gel with CHCl_3 -MeOH (5:1) to give **9** (470 mg, quant.). *Rf* 0.32 (20:1 CHCl_3 -MeOH); $[\alpha]_{\text{D}}^{25} + 11.1^\circ$ ($c=1.0$, CHCl_3). $^1\text{H-NMR}$ (CDCl_3) δ : 7.28 (m, 5H, Ar), 4.61 (d, 2H, $J=5.5$ Hz, Bn- CH_2), 4.20 (dd, 1H, $J=3.7$, 6.1 Hz, Ser α), 4.00 (d, 2H, $J=6.7$ Hz, Gly), 3.90 (dd, 1H, $J=3.7$, 10.4 Hz, Ser β -Ha), 3.80 (dd, 1H, $J=6.1$, 10.4 Hz, Ser β -Hb), 3.70 (s, 3H, OMe). $^{13}\text{C-NMR}$ (CDCl_3) δ : 53.2 (Ser α), 67.9 (Ser β), 41.2 (Gly), 52.3 (OMe), 73.6 (Bn- CH_2). *Anal.* Calcd for $\text{C}_{13}\text{H}_{18}\text{N}_2\text{O}_4$: C, 58.64; H, 6.81; N, 10.52. Found: C, 58.33; H, 6.55; N, 10.45.

5-Benzyl-L-glutamyl-O-benzyl-L-threonine Methyl Ester (10) A solution of compound **8** (940 mg, 1.7 mmol) in CH_2Cl_2 (4.0 ml) was treated with trifluoroacetic acid (2.0 ml) and the solution was stirred for 20 min at room temperature, then concentrated *in vacuo*. The residue was chromatographed on silica gel with CHCl_3 -MeOH (30:1) to give **10** (570 mg, 74%). *Rf* 0.65 (10:1 CHCl_3 -MeOH); $[\alpha]_{\text{D}}^{24} + 4.8^\circ$ ($c=2.0$, CHCl_3). $^1\text{H-NMR}$ (CDCl_3) δ : 7.30–7.16 (m, 10H, Ar), 5.00 (s, 2H, Bn- CH_2), 4.64 (dd, 1H, $J=2.4$, 6.1 Hz, Thr α), 4.50, 4.28 (each d, 2H, $J=11.9$ Hz, Bn- CH_2), 4.35 (br s, 1H, Glu α), 4.11 (dd, 1H, $J=1.8$, 6.1 Hz, Thr β), 3.54 (s, 3H, OMe), 2.67 (m, 2H, Glu γ), 2.22, 2.16 (each m, 2H, Glu β), 1.17 (d, 3H, $J=6.1$ Hz, Thr γ). $^{13}\text{C-NMR}$ (CDCl_3) δ : 52.4 (Glu α), 26.6 (Glu β), 29.2 (Glu γ), 57.2 (Thr α), 73.5 (Thr β), 15.6 (Thr γ), 52.3 (OMe), 70.5, 66.9 (Bn- CH_2). *Anal.* Calcd for $\text{C}_{24}\text{H}_{30}\text{N}_2\text{O}_6$: C, 65.14; H, 6.83; N, 6.33. Found: C, 65.30; H, 6.78; N, 6.01.

N-(Carbobenzoxy)-L-seryl-O-benzyl-L-seryl-glycine Methyl Ester (11) A solution of *N*-(carbobenzoxy)-L-serine (**1**) (350 mg, 1.5 mmol) and compound **9** (390 mg, 1.5 mmol) was prepared in 3:1 CH_2Cl_2 -THF (4.0 ml), then Et_3N (0.3 ml, 2.1 mmol) and DEPC (0.5 ml, 3.3 mmol) were added and the mixture was held at 0°C for 4 h. It was diluted with CHCl_3 , and the CHCl_3 solution was washed with water, dried over Na_2SO_4 , filtered, and concentrated. The residue was purified by column chromatography with CHCl_3 -MeOH (40:1) to give **11** (240 mg, 34%), which was recrystallized from MeOH. mp 123–125 $^\circ\text{C}$. *Rf* 0.54 (10:1 CHCl_3 -MeOH); $[\alpha]_{\text{D}}^{25} + 7.2^\circ$ ($c=0.9$, CHCl_3). $^1\text{H-NMR}$ (CDCl_3) δ : 7.32–7.24 (m, 10H, Ar), 5.08 (br d, 2H, Z- CH_2), 4.71 (s, 1H, Ser α), 4.51 (br d, 2H, Bn- CH_2), 4.34 (t, 1H, $J=3.7$ Hz, Ser α), 3.64 (s, 3H, OMe). $^{13}\text{C-NMR}$ (CDCl_3) δ : 53.1, 55.9 (Ser α), 63.1, 69.1 (Ser β), 41.3 (Gly), 52.4 (OMe), 73.5 (Bn- CH_2), 67.2 (Z- CH_2). *Anal.* Calcd for $\text{C}_{24}\text{H}_{29}\text{N}_3\text{O}_8$: C, 59.13; H, 6.00; N, 8.62. Found: C, 59.00; H, 6.15; N, 8.39.

N-(tert-Butoxycarbonyl)-4-benzyl-L-aspartyl-5-benzyl-L-glutamyl-O-benzyl-L-threonine Methyl Ester (12) A solution of *N*-(tert-butoxycarbonyl)-O-benzyl-L-aspartic acid 4-benzyl ester (**4**) (560 mg, 1.7 mmol) and compound **10** (390 mg, 0.87 mmol) was prepared in CH_2Cl_2 (5.0 ml), then Et_3N (0.13 ml, 0.91 mmol) and DEPC (0.26 ml, 1.7 mmol) were added and the mixture was held at 0°C for 14 h. It was diluted with CHCl_3 , and the CHCl_3 solution was washed with water, dried over Na_2SO_4 , filtered, and concentrated. The residue was purified by column chromatography with CHCl_3 -MeOH (50:1) to give **12** (450 mg, 69%). *Rf* 0.48 (4:1 benzene-acetone); $[\alpha]_{\text{D}}^{25} + 2.8^\circ$ ($c=2.0$, CHCl_3). $^1\text{H-NMR}$ (CDCl_3) δ : 7.33–7.23 (m, 15H, Ar), 5.10 (d, 2H, $J=2.4$ Hz, Bn- CH_2), 5.05 (dd, 2H, $J=12.2$, 25.0 Hz, Bn- CH_2), 4.64–4.59 (m, 2H, Glu α , Thr α), 4.55, 4.35 (each d, 2H, $J=11.9$ Hz, Bn- CH_2), 4.52 (br s, 1H, Asp α), 4.13 (m, 1H, Thr β), 3.61 (s, 3H, OMe), 3.02 (dd, 1H, $J=3.7$, 17.1 Hz, Asp α -Ha), 2.71 (dd, 1H, $J=5.5$, 17.1 Hz, Asp α -Hb), 2.53 (m, 2H, Glu γ), 2.21, 2.00 (each m, 2H, Glu β), 1.43 (s, 9H, 3 \times *tert*-Bu- CH_3), 1.17 (d, 3H, $J=6.7$ Hz, Thr γ). $^{13}\text{C-NMR}$ (CDCl_3) δ : 50.8 (Asp α), 36.1 (Asp β), 52.4 (Glu α), 28.0 (Glu β), 30.0 (Glu γ), 56.8 (Thr α), 73.7 (Thr β),

16.2 (Thr γ), 52.3 (OMe), 28.2 (*tert*-Bu- CH_3), 80.5 (*tert*-Bu-C), 70.7, 66.8, 66.4 (Bn- CH_2). *Anal.* Calcd for $\text{C}_{46}\text{H}_{49}\text{N}_3\text{O}_{11}$: C, 64.24; H, 6.60; N, 5.62. Found: C, 65.25; H, 6.23; N, 5.32.

N-(Carbobenzoxy)-L-seryl-O-benzyl-L-seryl-glycine (13) A solution of compound **11** (60 mg, 0.12 mmol) in 5:1 MeOH- H_2O (3.0 ml) was treated with NaOMe (30 mg) at room temperature for 30 min. The mixture was neutralized with Amberlite IR-120 (H^+), filtered, and concentrated. The residue was purified by column chromatography with CHCl_3 -MeOH (10:1) to give **13** (55 mg, 94%), which was recrystallized from MeOH. mp 105–107 $^\circ\text{C}$. *Rf* 0.37 (3:2 CHCl_3 -MeOH); $[\alpha]_{\text{D}}^{25} + 4.9^\circ$ ($c=1.0$, CHCl_3). $^1\text{H-NMR}$ (CD_3OD) δ : 7.34–7.22 (m, 10H, Ar), 5.09 (d, $J=4.9$ Hz, 2H, Z- CH_2), 4.67 (br s, 1H, Ser α), 4.53 (d, $J=3.7$ Hz, 2H, Bn- CH_2), 4.29 (t, $J=5.8$ Hz, 1H, Ser α). $^{13}\text{C-NMR}$ (CD_3OD) δ : 58.3, 54.8 (Ser α), 70.4, 63.2 (Ser β), 41.9 (Gly), 74.2 (Bn- CH_2), 67.9 (Z- CH_2). *Anal.* Calcd for $\text{C}_{23}\text{H}_{27}\text{N}_3\text{O}_8$: C, 58.35; H, 5.75; N, 8.87. Found: C, 58.56; H, 5.77; N, 9.12.

4-Benzyl-L-aspartyl-5-benzyl-L-glutamyl-O-benzyl-L-threonine Methyl Ester (14) A solution of compound **12** (340 mg, 0.46 mmol) in CH_2Cl_2 (2.0 ml) was treated with trifluoroacetic acid (1.0 ml) and the solution was stirred for 20 min at room temperature, then concentrated *in vacuo*. The residue was chromatographed on silica gel with CHCl_3 -MeOH (30:1) to give **14** (250 mg, 84%). *Rf* 0.69 (10:1 CHCl_3 -MeOH); $[\alpha]_{\text{D}}^{25} + 7.5^\circ$ ($c=1.0$, CHCl_3). $^1\text{H-NMR}$ (CDCl_3) δ : 7.29–7.16 (m, 15H, Ar), 5.08–4.99 (m, 4H, 2 \times Bn- CH_2), 4.62–4.57 (m, 2H, Glu α , Thr α), 4.51 (t, 1H, $J=6.7$ Hz, Asp α), 4.48, 4.29 (each d, 2H, $J=11.9$ Hz, Bn- CH_2), 4.04 (ddd, 1H, $J=2.4$, 6.1, 12.8 Hz, Thr β), 3.53 (s, 3H, OMe), 3.00 (d, 2H, $J=6.1$ Hz, Asp β), 2.48 (t, 2H, $J=7.3$ Hz, Glu γ), 2.14, 2.00 (each m, 2H, Glu β), 1.09 (d, 3H, $J=6.1$ Hz, Thr γ). $^{13}\text{C-NMR}$ (CDCl_3) δ : 49.7 (Asp α), 34.8 (Asp β), 53.3 (Glu α), 27.2 (Glu β), 30.1 (Glu γ), 56.8 (Thr α), 73.6 (Thr β), 15.9 (Thr γ), 52.2 (OMe), 70.5, 67.5, 66.5 (Bn- CH_2). *Anal.* Calcd for $\text{C}_{35}\text{H}_{41}\text{N}_3\text{O}_9$: C, 64.90; H, 6.38; N, 6.49. Found: C, 64.85; H, 6.12; N, 6.65.

N-(Carbobenzoxy)-L-seryl-O-benzyl-L-seryl-glycyl-4-benzyl-L-aspartyl-5-benzyl-L-glutamyl-O-benzyl-L-threonine Methyl Ester (15) A solution of compound **13** (230 mg, 0.47 mmol) and compound **14** (230 mg, 0.36 mmol) in CH_2Cl_2 (3.0 ml) were treated with EEDQ (230 mg, 0.93 mmol) at 0°C and the mixture was allowed to stand at room temperature for 24 h. It was diluted with CHCl_3 , and the CHCl_3 solution was washed with water, dried over Na_2SO_4 , filtered, and concentrated. The residue was purified by column chromatography with CHCl_3 -MeOH (50:1) to give **15** (240 mg, 62%). *Rf* 0.61 (10:1 CHCl_3 -MeOH); $[\alpha]_{\text{D}}^{24} + 15.3^\circ$ ($c=1.0$, CHCl_3). $^1\text{H-NMR}$ (CD_3OD) δ : 7.31–7.23 (m, 25H, Ar), 5.10–5.04 (m, 6H, 2 \times Bn- CH_2 , Z- CH_2), 4.79 (dd, 1H, $J=5.5$, 7.3 Hz, Asp α), 4.29 (t, $J=5.5$ Hz, 1H, Ser α), 4.13 (m, 1H, Thr β), 3.62 (s, 3H, OMe), 2.92 (dd, 1H, $J=5.5$, 16.5 Hz, Asp β -Ha), 2.81 (dd, 1H, $J=7.3$, 16.5 Hz, Asp β -Hb), 2.50 (m, 2H, Glu γ), 2.18, 2.04 (each m, 2H, Glu β), 1.18 (d, 3H, $J=6.7$ Hz, Thr γ). $^{13}\text{C-NMR}$ (CD_3OD) δ : 58.3, 55.8 (Ser α), 70.1, 63.4 (Ser β), 44.2 (Gly), 51.4 (Asp α), 36.9 (Asp β), 54.2 (Glu α), 28.3 (Glu β), 31.4 (Glu γ), 58.5 (Thr α), 75.6 (Thr β), 16.6 (Thr γ), 52.8 (OMe), 74.4, 72.1, 68.1, 67.5 (Bn- CH_2), 67.8 (Z- CH_2). *Anal.* Calcd for $\text{C}_{58}\text{H}_{66}\text{N}_6\text{O}_{16}$: C, 63.15; H, 6.03; N, 7.62. Found: C, 62.96; H, 5.88; N, 7.51.

N-(Carbobenzoxy)-O-(3,4,6-tri-O-acetyl-2-azido-2-deoxy- α -D-galactopyranosyl)-L-seryl-O-benzyl-L-seryl-glycine Methyl Ester (17 α) and N-(carbobenzoxy)-O-(3,4,6-tri-O-acetyl-2-azido-2-deoxy- β -D-galactopyranosyl)-L-seryl-O-benzyl-L-seryl-glycine Methyl Ester (17 β) A suspension of 3,4,6-tri-O-acetyl-2-azido-2-deoxy- α -D-galactopyranosyl trichloroacetimidate (**16 α**) (220 mg, 0.45 mmol), compound **11** (220 mg, 0.45 mmol) and molecular sieves (AW 300) (250 mg) in dry CH_2Cl_2 (5.0 ml) was stirred at room temperature for 1 h, then 10% TMSOTf/ CH_2Cl_2 (0.1 ml) was added at -30°C . The mixture was stirred under an argon atmosphere for 14 h, then diluted with CHCl_3 and filtered through a pad of Celite. The filtrate was washed with water, and then dried over Na_2SO_4 , filtered, and concentrated. The residue was chromatographed on silica gel with CHCl_3 -MeOH (100:1) to provide an anomeric mixture of **17 α** and **17 β** (230 mg, 63%).

The same procedure from 4,6-tri-O-acetyl-2-azido-2-deoxy- β -D-galactopyranosyl trichloroacetimidate (**16 β**) and compound **11** gave an anomeric mixture of **17 α** and **17 β** (160 mg, 52%). *Rf* 0.82 (8:1 CHCl_3 -MeOH). Compound **17 α** : $^1\text{H-NMR}$ (CDCl_3) δ : 7.36–7.26 (m, 10H, Ar), 5.11 (br s, 1H, H-1), 4.56 (s, 2H, Bn- CH_2), 3.73 (s, 3H, OMe), 2.08, 2.04, 2.01 (each s, 9H, 3 \times OAc). Compound **17 β** : $^1\text{H-NMR}$ (CDCl_3) δ : 7.36–7.26 (m, 10H, Ar), 4.45 (d, $J=7.9$ Hz, 1H, H-1), 4.55 (d, 2H, $J=2.5$ Hz, Bn- CH_2), 3.73 (s, 3H, OMe), 2.14, 2.01, 2.00 (each s, 9H,

3 × OAc).

N-(Carbobenzoxy)-O-(3,4,6-tri-O-acetyl-2-acetamido-2-deoxy- α -D-galactopyranosyl)-L-seryl-O-benzyl-L-seryl-glycine Methyl Ester (18 α) and N-(Carbobenzoxy)-O-(3,4,6-tri-O-acetyl-2-acetamido-2-deoxy- β -D-galactopyranosyl)-L-seryl-O-benzyl-L-seryl-glycine Methyl Ester (18 β) A solution of compound **17 α** and **17 β** (230 mg, 0.29 mmol), NiCl₂·6H₂O (3.1 g, 12.9 mmol) and H₃BO₃ (560 mg, 9.5 mmol) in a mixture of EtOH (16 ml) and AcOEt (4.0 ml) was treated with an EtOH solution of NaBH₄ with stirring until the solution became black. Stirring was continued at room temperature for 1 h. Then, Ac₂O (2.3 ml) was added and the mixture was stirred for 4 h, then filtered through the pad of Celite, and CHCl₃ and water were added to the filtrate. The organic layer was separated, washed with water, dried over Na₂SO₄, filtered, and concentrated. The residue was chromatographed on silica gel with CHCl₃-MeOH (80:1) to provide **18 α** (150 mg, 64%) and **18 β** (77 mg, 33%).

Data for **18 α** ; *Rf* 0.64 (10:1 CHCl₃-MeOH); [α]_D²⁵ +95.1° (*c*=0.4, CHCl₃); ¹H-NMR (CDCl₃) δ : 7.37–7.28 (m, 10H, Ar), 5.11 (s, 2H, Z-CH₂), 4.98 (d, 1H, *J*=2.9 Hz, H-1), 4.58 (d, 2H, *J*=4.8 Hz, Bn-CH₂), 3.84 (s, 3H, OMe), 2.13, 2.00, 1.96, 1.94 (each s, 12H, 4 × Ac). *Anal.* Calcd for C₃₈H₄₈N₄O₁₆: C, 55.88; H, 5.92; N, 6.86. Found: C, 55.54; H, 5.63; N, 6.90.

Data for **18 β** ; *Rf* 0.56 (10:1 CHCl₃-MeOH); [α]_D²⁵ +0.9° (*c*=0.5, CHCl₃). ¹H-NMR (CDCl₃) δ : 7.36–7.24 (m, 10H, Ar), 5.09 (s, 2H, Z-CH₂), 4.76 (d, 1H, *J*=8.6 Hz, H-1), 4.56 (br s, 2 H, Bn-CH₂), 3.72 (s, 3H, OMe), 2.09, 2.02, 1.98, 1.84 (each s, 12H, 4 × Ac). *Anal.* Calcd for C₃₈H₄₈N₄O₁₆: C, 55.88; H, 5.92; N, 6.86. Found: C, 55.58; H, 5.70; N, 6.79.

3,4,6-Tri-O-acetyl-2-acetamido-2-deoxy- α -D-galactopyranosyl-L-seryl-L-seryl-glycine Methyl Ester (19 α) A solution of compound **18 α** (150 mg, 0.18 mmol) in 2:1 MeOH-AcOH (2.0 ml) was stirred with 10% Pd-C (75 mg) for 12 h under an H₂ atmosphere, then filtered through a pad of Celite and concentrated to dryness. The residue was chromatographed on silica gel with CHCl₃-MeOH (5:1) to provide **19 α** (87 mg, 79%). *Rf* 0.35 (5:1 CHCl₃-MeOH); [α]_D²⁵ +73.2° (*c*=0.4, MeOH). ¹H-NMR (CD₃OD) δ : 5.00 (d, 1H, *J*=3.6 Hz, H-1), 3.37 (s, 3H, OMe), 2.14, 2.03, 1.98, 1.94 (each s, 12H, 4 × Ac). *Anal.* Calcd for C₂₃H₃₆N₄O₁₄: C, 46.62; H, 6.12; N, 9.46. Found: C, 46.76; H, 5.82; N, 9.53.

3,4,6-Tri-O-acetyl-2-acetamido-2-deoxy- β -D-galactopyranosyl-L-seryl-L-seryl-glycine Methyl Ester (19 β) A solution of compound **18 β** (77 mg, 0.094 mmol) in 2:1 MeOH-AcOH (2.0 ml) was stirred with 10% Pd-C (38 mg) for 12 h under an H₂ atmosphere, then filtered through a pad of Celite and concentrated to dryness. The residue was chromatographed on silica gel with CHCl₃-MeOH (5:1) to provide **19 β** (46 mg, 82%). *Rf* 0.25 (8:1 CHCl₃-MeOH); [α]_D²⁵ -5.7° (*c*=0.3, MeOH). ¹H-NMR (CDCl₃) δ : 4.66 (d, 1H, *J*=8.0 Hz, H-1), 3.75 (s, 3H, OMe), 2.16, 2.05, 1.99, 1.97 (each s, 12H, 4 × Ac). *Anal.* Calcd for C₂₃H₃₆N₄O₁₄: C, 46.62; H, 6.12; N, 9.46. Found: C, 46.64; H, 5.82; N, 9.00.

2-Acetamido-2-deoxy- α -D-galactopyranosyl-L-seryl-L-seryl-glycine (20 α , Supprescin A) A solution of compound **19 α** (53 mg, 0.089 mmol) in 3:1 MeOH-H₂O (2.0 ml) was treated with NaOMe (30 mg) at room temperature for 12 h. The mixture was neutralized with Amberlite IR-120 (H⁺), filtered, and concentrated. The residue was chromatographed over Sephadex LH-20 with MeOH-H₂O (3:1) provide **20 α** (24 mg, 60%). *Rf* 0.47 (1:3:1 CHCl₃-MeOH-H₂O); [α]_D²⁵ +8.9° (*c*=0.5, H₂O). ¹H-NMR (D₂O) δ : 4.92 (d, 1H, *J*=3.7 Hz, H-1), 4.58 (t, 1H, *J*=5.2 Hz, Ser α), 4.24 (t, 1H, *J*=4.3 Hz, Ser α). FAB-MS *m/z*: 453 (M+H)⁺. *Anal.* Calcd for C₁₆H₂₈N₄O₁₁: C, 42.48; H, 6.24; N, 12.38. Found: C, 42.66; H, 6.02; N, 12.05.

2-Acetamido-2-deoxy- β -D-galactopyranosyl-L-seryl-L-seryl-glycine (20 β) A solution of compound **19 β** (46 mg, 0.078 mmol) in 3:1 MeOH-H₂O (2.0 ml) was treated with NaOMe (26 mg) at room temperature for 12 h. The mixture was neutralized with Amberlite IR-120 (H⁺), filtered, and concentrated. The residue was chromatographed over Sephadex LH-20 with MeOH-H₂O (3:1) to provide **20 β** (28 mg, 80%). *Rf* 0.43 (1:3:1 CHCl₃-MeOH-H₂O); [α]_D²⁵ -2.5° (*c*=0.5, H₂O). ¹H-NMR (D₂O) δ : 4.53 (d, 1H, *J*=5.5 Hz, H-1), 4.51 (t, 1H, *J*=8.5 Hz, Ser α), 4.14 (t, 1H, *J*=5.4 Hz, Ser α). FAB-MS *m/z*: 453 (M+H)⁺. *Anal.* Calcd for C₁₆H₂₈N₄O₁₁: C, 42.48; H, 6.24; N, 12.38. Found: C, 42.02; H, 6.14; N, 12.11.

2,3,4,6-Tetra-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-1,2,3,6-tetra-O-benzoyl- α -D-galactopyranose (23) A solution of 2,3,4,6-tetra-O-acetyl- α -D-galactopyranosyl bromide (**21**) (1.4 g, 3.4 mmol) in dry toluene (8.0 ml) was added dropwise with stirring to a solution of 1,2,3,6-

tetra-O-benzoyl- α -D-galactopyranose (**22**) (1.0 g, 1.7 mmol), silver trifluoromethanesulfonate (1.3 g, 5.1 mmol), 2,4,6-trimethylpyridine (500 mg) and molecular sieves (AW 300) (1.0 g) in dry toluene (8.0 ml) at -10 °C in the dark. The mixture was allowed to attain room temperature, then stirred overnight. It was filtered through a pad of Celite, and CHCl₃ and water were added to the filtrate. The organic layer was separated, washed with water, dried over Na₂SO₄, filtered, and then concentrated. The residue was chromatographed on silica gel with benzene-acetone (40:1) to provide **23** (870 mg, 56%). *Rf* 0.57 (4:1 benzene-acetone); [α]_D²⁵ +25.0° (*c*=1.0, CHCl₃). ¹H-NMR (CDCl₃) δ : 8.11–7.25 (m, 20H, Ar), 6.76 (d, 1H, *J*=3.7 Hz, H-1), 4.75 (d, 1H, *J*=7.9 Hz, H-1'), 2.27, 2.16, 2.01, 1.92 (each s, 12H, 4 × OAc). *Anal.* Calcd for C₄₈H₄₆O₁₉: C, 62.20; H, 5.00. Found: C, 61.95; H, 4.92.

2,3,4,6-Tetra-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzoyl- α -D-galactopyranosyl bromide (24) A solution of compound **23** (680 mg, 0.73 mmol) in CHCl₃ (6.0 ml) was treated with 25% HBr/AcOH (5.0 ml). The mixture was stirred at 0 °C for 2 h, then diluted with CHCl₃, and the CHCl₃ solution was washed with water, dried over Na₂SO₄, filtered, and concentrated to give **24** (640 mg, 99%). *Rf* 0.59 (4:1 benzene-acetone); [α]_D²⁵ +19.8° (*c*=1.0, CHCl₃). ¹H-NMR (CDCl₃) δ : 8.09–7.33 (m, 15H, Ar), 6.76 (d, 1H, *J*=3.6 Hz, H-1), 4.68 (d, 1H, *J*=7.9 Hz, H-1'), 2.20, 2.16, 2.00, 1.94 (each s, 12H, 4 × OAc). *Anal.* Calcd for C₄₀H₃₉BrO₁₇: C, 55.12; H, 4.51. Found: C, 54.92; H, 4.51.

2,3,4,6-Tetra-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-1,5-anhydro-3,6-di-O-benzoyl-2-deoxy-D-lyxo-hex-1-enitol (25) A solution of compound **24** (910 mg, 1.0 mmol) in AcOEt (3.0 ml) was added with stirring to a Zn-Cu reagent that had been prepared by addition of Zn dust (1.9 g) to acetate buffer [AcONa·3H₂O (130 mg)/60% AcOH-H₂O (13 ml)] containing CuSO₄·5H₂O (14 mg). The mixture was stirred at room temperature for 3 h, then filtered through a pad of Celite, and AcOEt and water were added to the filtrate. The organic layer was separated, washed with water, dried over Na₂SO₄, filtered, and concentrated. The residue was chromatographed on silica gel with CHCl₃ to provide **25** (620 mg, 88%). *Rf* 0.59 (4:1 benzene-acetone); [α]_D²⁵ +15.5° (*c*=1.0, CHCl₃). ¹H-NMR (CDCl₃) δ : 8.09–7.33 (m, 15H, Ar), 6.47 (d, 1H, *J*=6.1 Hz, H-1), 4.77 (d, 1H, *J*=7.9 Hz, H-1'), 2.20, 2.16, 2.00, 1.94 (each s, 12H, 4 × OAc). *Anal.* Calcd for C₃₂H₃₀O₁₅: C, 59.65; H, 5.30. Found: C, 59.54; H, 5.31.

2,3,4,6-Tetra-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzoyl-2-azido-2-deoxy-D-galactopyranosyl Nitrate (26) A solution of compound **25** (26 mg, 0.038 mmol), NaN₃ (6.0 mg) and cerium(IV) ammonium nitrate (CAN) (60 mg) in CH₃CN (1.0 ml) was stirred at -15 °C for 2 h, and then at room temperature for 2 h. The mixture was diluted with CHCl₃, and the CHCl₃ solution was washed with water, dried with Na₂SO₄, filtered, and evaporated. The residue was chromatographed on silica gel with benzene-acetone (25:1) to provide an anomeric mixture of **26** (19 mg, 64%). *Rf* 0.59 (4:1 benzene-acetone). The α -anomer: ¹H-NMR (CDCl₃) δ : 8.17–7.33 (m, 10H, Ar), 6.43 (d, 1H, *J*=4.0 Hz, H-1), 4.57 (d, 1H, *J*=7.9 Hz, H-1'), 2.15, 2.13, 2.00, 1.93 (each s, 12H, 4 × OAc).

The β -anomer: ¹H-NMR (CDCl₃) δ : 8.17–7.33 (m, 10H, Ar), 5.64 (d, 1H, *J*=8.5 Hz, H-1), 4.76 (d, 1H, *J*=8.9 Hz, H-1'), 2.15, 2.14, 2.00, 1.94 (each s, 12H, 4 × OAc).

2,3,4,6-Tetra-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzoyl-2-azido-2-deoxy-D-galactopyranose (27) A solution of compound **26** (150 mg, 0.19 mmol) in 1,4-dioxane (2.0 ml) was added with stirring to an NaNO₂ (250 mg) solution in 0.4 ml of H₂O. The mixture was stirred at 80 °C for 22 h, then diluted with CHCl₃, and the CHCl₃ solution was washed with water, dried with Na₂SO₄, filtered, and evaporated. The residue was chromatographed on silica gel with CHCl₃-MeOH (100:1) to provide an anomeric mixture of **27** (77 mg, 54%). *Rf* 0.40 (8:1 CHCl₃-MeOH).

The α -anomer: ¹H-NMR (CDCl₃) δ : 8.19–7.40 (m, 10H, Ar), 5.51 (d, 1H, *J*=3.1 Hz, H-1), 4.63 (d, 1H, *J*=7.9 Hz, H-1'), 2.14, 2.13, 2.00, 1.92 (each s, 12H, 4 × OAc).

The β -anomer: ¹H-NMR (CDCl₃) δ : 8.19–7.40 (m, 10H, Ar), 4.73 (d, 1H, *J*=7.9 Hz, H-1), 4.64 (d, 1H, *J*=6.7 Hz, H-1'), 2.16, 2.15, 2.13, 1.91 (each s, 12H, 4 × OAc).

2,3,4,6-Tetra-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzoyl-2-azido-2-deoxy-D-galactopyranosyl Trichloroacetimidate (28) A solution of compound **27** (76 mg, 0.10 mmol) and K₂CO₃ (100 mg) in CH₂Cl₂ (1.5 ml) was treated with CCl₃CN (0.1 ml). Stirring was continued at 0 °C for 4 h. The mixture was diluted with CHCl₃, and the CHCl₃ solution was washed with water, dried with Na₂SO₄, filtered,

and evaporated. The residue was chromatographed on silica gel with benzene-acetone (50:1) to provide an anomeric mixture of **28** (77 mg, 54%). *Rf* 0.64 (8:1 CHCl₃-MeOH).

The α -anomer: ¹H-NMR (CDCl₃) δ : 8.19–7.41 (m, 10H, Ar), 6.61 (d, 1H, *J* = 3.7 Hz, H-1), 4.60 (d, 1H, *J* = 7.9 Hz, H-1'), 2.16, 2.14, 2.00, 1.96 (each s, 12H, 4 \times OAc).

The β -anomer: ¹H-NMR (CDCl₃) δ : 8.19–7.41 (m, 10H, Ar), 5.77 (d, 1H, *J* = 7.9 Hz, H-1), 4.65 (d, 1H, *J* = 7.3 Hz, H-1'), 2.15, 2.14, 2.00, 1.92 (each s, 12H, 4 \times OAc).

***N*-(Carbobenzoxy)-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-(3,6-di-*O*-benzoyl-2-deoxy- α -D-galactopyranosyl)-L-seryl-*O*-benzyl-L-seryl-glycyl-4-benzyl-L-aspartyl-5-benzyl-L-glutamyl-*O*-benzyl-L-threonine Methyl Ester (**29 α**) and ***N*-(Carbobenzoxy)-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-(3,6-di-*O*-benzoyl-2-azido-2-deoxy- β -D-galactopyranosyl)-L-seryl-*O*-benzyl-L-seryl-glycyl-4-benzyl-L-aspartyl-5-benzyl-L-glutamyl-*O*-benzyl-L-threonine Methyl Ester (**29 β**)** A solution of compound **28** (230 mg, 0.26 mmol), compound **15** (240 mg, 0.22 mmol) and molecular sieves (AW 300) (400 mg) in dry CH₂Cl₂ (5 ml) was stirred at room temperature for 5 h, then 10% TMSOTf/CH₂Cl₂ (0.2 ml) was added at -30°C. The whole was stirred under an argon atmosphere for 14 h, then diluted with CHCl₃ and filtered through the pad of Celite. The filtrate was washed with water, and then dried over Na₂SO₄, filtered, and concentrated. The residue was chromatographed on silica gel with CHCl₃-MeOH (100:1) to provide **29 α** (120 mg, 31%) and **29 β** (83 mg, 22%).**

Data for **29 α** : *Rf* 0.65 (10:1 CHCl₃-MeOH); $[\alpha]_D^{25} + 17.8^\circ$ (*c* = 1.0, CHCl₃). ¹H-NMR (CDCl₃) δ : 8.06–7.99, 7.47–7.21 (each m, 30H, Ar), 5.24 (d, 1H, *J* = 3.7 Hz, H-1), 5.06–5.01 (m, 6H, Z-CH₂, 2 \times Bn-CH₂), 4.86 (brs, 1H, Asp α), 4.52 (d, 1H, *J* = 7.8 Hz, H-1'), 3.81 (brs, 2H, Gly), 3.60 (s, 3H, OMe), 2.50 (m, 2H, Glu γ), 2.03, 1.98, 1.96, 1.94 (each s, 12H, 4 \times OAc), 1.14 (dd, 3H, *J* = 6.1, 9.2 Hz, Thr γ). *Anal.* Calcd for C₉₂H₁₀₁N₉O₃₁: C, 60.42; H, 5.57; N, 6.89. Found: C, 60.22; H, 5.65; N, 6.75.

Data for **29 β** : *Rf* 0.62 (10:1 CHCl₃-MeOH); $[\alpha]_D^{25} + 15.9^\circ$ (*c* = 0.5, CHCl₃). ¹H-NMR (CDCl₃) δ : 8.06–7.99, 7.36–7.24 (each m, 30H, Ar), 5.10–5.03 (m, 6H, Z-CH₂, 2 \times Bn-CH₂), 4.87 (brs, 1H, Asp α), 4.60 (d, 1H, *J* = 7.7 Hz, H-1), 4.45 (d, 1H, *J* = 7.5 Hz, H-1'), 3.62 (s, 3H, OMe), 2.50 (m, 2H, Glu γ), 2.08, 2.03, 1.97, 1.96 (each s, 12H, 4 \times OAc), 1.16 (brs, 3H, Thr γ). *Anal.* Calcd for C₉₂H₁₀₁N₉O₃₁: C, 60.42; H, 5.57; N, 6.89. Found: C, 60.28; H, 5.73; N, 6.86.

***N*-(Carbobenzoxy)-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-(3,6-di-*O*-benzoyl-2-acetamido-2-deoxy- α -D-galactopyranosyl)-L-seryl-*O*-benzyl-L-seryl-glycyl-4-benzyl-L-aspartyl-5-benzyl-L-glutamyl-*O*-benzyl-L-threonine Methyl Ester (**30 α**)** A stirred solution of compound **29 α** (74 mg, 0.040 mmol), NiCl₂·6H₂O (360 mg, 1.5 mmol) and H₃BO₃ (180 mg, 2.9 mmol) in EtOH (15 ml) was treated dropwise with NaBH₄ in EtOH at 0°C until the color changed to black. After 1 h, Ac₂O (10 ml) was added, then after 4 h, the mixture was diluted with CHCl₃, washed with water, dried over Na₂SO₄, filtered, and concentrated. The residue was chromatographed on silica gel with CHCl₃-MeOH (50:1) to provide **30 α** (64 mg, 86%). *Rf* 0.56 (10:1 CHCl₃-MeOH); $[\alpha]_D^{25} - 23.1^\circ$ (*c* = 0.4, CHCl₃). ¹H-NMR (CDCl₃) δ : 8.04–8.01, 7.47–7.22 (each m, 30H, Ar), 5.23 (d, 1H, *J* = 3.5 Hz, H-1), 5.10–5.03 (m, 6H, Z-CH₂, 2 \times Bn-CH₂), 4.87 (brs, 1H, Asp α), 4.77 (d, 1H, *J* = 8.0 Hz, H-1'), 3.62 (s, 3H, OMe), 2.50 (m, 2H, Glu γ), 2.15, 2.08, 2.03, 1.97, 1.96 (each s, 15H, 5 \times Ac), 1.16 (brs, 3H, Thr γ). *Anal.* Calcd for C₉₄H₁₀₅N₇O₃₂: C, 61.20; H, 5.74; N, 5.31. Found: C, 61.15; H, 5.55; N, 5.11.

***N*-(Carbobenzoxy)-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-(3,6-di-*O*-benzoyl-2-acetamido-2-deoxy- β -D-galactopyranosyl)-L-seryl-*O*-benzyl-L-seryl-glycyl-4-benzyl-L-aspartyl-5-benzyl-L-glutamyl-*O*-benzyl-L-threonine Methyl Ester (**30 β**)** A solution of compound **29 β** (64 mg, 0.035 mmol), NiCl₂·6H₂O (360 mg, 1.5 mmol) and H₃BO₃ (180 mg, 2.9 mmol) in EtOH (15 ml) was treated dropwise with NaBH₄ in EtOH at 0°C until the color was changed to black. After 1 h, Ac₂O (10 ml) was added, then after 4 h, the mixture was diluted with CHCl₃, washed with water, dried over Na₂SO₄, filtered, and concentrated. The residue was chromatographed on silica gel with CHCl₃-MeOH (50:1) to provide **30 β** (60 mg, 93%). *Rf* 0.53 (10:1 CHCl₃-MeOH); $[\alpha]_D^{25} - 15.4^\circ$ (*c* = 0.5, CHCl₃). ¹H-NMR (CDCl₃) δ : 8.06–8.02, 7.48–7.20 (each m, 30H, Ar), 5.11–5.03 (m, 6H, Z-CH₂, 2 \times Bn-CH₂), 4.87 (brs, 1H, Asp α), 4.72 (d, 1H, *J* = 7.8 Hz, H-1'), 4.40 (d, 1H, *J* = 7.5 Hz, H-1), 3.62 (s, 3H, OMe), 2.50 (m, 2H, Glu γ), 2.14, 2.09, 2.00, 1.98, 1.97 (each s, 15H, 5 \times Ac), 1.17 (brs, 3H, Thr γ). *Anal.* Calcd for C₉₄H₁₀₅N₇O₃₂:

C, 61.20; H, 5.74; N, 5.31. Found: C, 61.33; H, 5.65; N, 5.22.

2,3,4,6-Tetra-*O*-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-(3,6-di-*O*-benzoyl-2-acetamido-2-deoxy- α -D-galactopyranosyl)-L-seryl-L-seryl-glycyl-L-aspartyl-L-glutamyl-L-threonine Methyl Ester (31 α**)** A solution of compound **30 α** (50 mg, 0.027 mmol) in 2:1 MeOH-AcOH (1.5 ml) was stirred with 10% Pd-C (25 mg) for 12 h under an H₂ atmosphere, then filtered through a pad of Celite and concentrated to dryness. The residue was chromatographed on silica gel with CHCl₃-MeOH (2:1) to provide **31 α** (24 mg, 65%). *Rf* 0.74 (5:4:1 CHCl₃-MeOH-H₂O); $[\alpha]_D^{25} - 10.2^\circ$ (*c* = 0.5, CHCl₃). ¹H-NMR (CD₃OD) δ : 8.04–8.00, 7.48–7.22 (each m, 10H, Ar), 5.23 (d, 1H, *J* = 3.7 Hz, H-1), 4.85 (brs, 1H, Asp α), 4.75 (d, 1H, *J* = 7.8 Hz, H-1'), 3.63 (s, 3H, OMe), 2.49 (m, 2H, Glu γ), 2.12, 2.05, 2.02, 1.98, 1.95 (each s, 15H, 5 \times Ac), 1.15 (brs, 3H, Thr γ). *Anal.* Calcd for C₅₈H₇₅N₇O₃₀: C, 51.59; H, 5.60; N, 7.26. Found: C, 51.67; H, 5.45; N, 7.35.

2,3,4,6-Tetra-*O*-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-(3,6-di-*O*-benzoyl-2-acetamido-2-deoxy- β -D-galactopyranosyl)-L-seryl-L-seryl-glycyl-L-aspartyl-L-glutamyl-L-threonine Methyl Ester (31 β**)** A solution of compound **30 β** (40 mg, 0.022 mmol) in 2:1 MeOH-AcOH (1.5 ml) was stirred with 10% Pd-C (20 mg) for 12 h under an H₂ atmosphere, then filtered through a pad of Celite and concentrated to dryness. The residue was chromatographed on silica gel with CHCl₃-MeOH (2:1) to provide **31 β** (20 mg, 68%). *Rf* 0.63 (5:4:1 CHCl₃-MeOH-H₂O); $[\alpha]_D^{25} - 5.4^\circ$ (*c* = 0.5, CHCl₃). ¹H-NMR (CD₃OD) δ : 8.06–8.00, 7.47–7.20 (each m, 10H, Ar), 4.87 (brs, 1H, Asp α), 4.70 (d, 1H, *J* = 8.0 Hz, H-1'), 4.42 (d, 1H, *J* = 7.6 Hz, H-1), 3.62 (s, 3H, OMe), 2.50 (m, 2H, Glu γ), 2.15, 2.05, 2.03, 1.99, 1.98 (each s, 15H, 5 \times Ac), 1.16 (brs, 3H, Thr γ). *Anal.* Calcd for C₅₈H₇₅N₇O₃₀: C, 51.59; H, 5.60; N, 7.26. Found: C, 51.38; H, 5.80; N, 7.13.

β -D-Galactopyranosyl-(1 \rightarrow 4)-(2-acetamido-2-deoxy- α -D-galactopyranosyl)-L-seryl-L-seryl-glycyl-L-aspartyl-L-glutamyl-L-threonine (32 α** , **Suppressin B**)** A solution of compound **31 α** (16 mg, 0.012 mmol) in 3:1 MeOH-H₂O (2 ml) was treated with NaOMe (8 mg) at room temperature for 1 h. The mixture was neutralized with Amberlite IR-120 (H⁺), filtered, and concentrated. The residue was chromatographed over Sephadex LH-20 with MeOH-H₂O (3:1) to provide **32 α** (8 mg, 71%). *Rf* 0.68 (1:3:1 CHCl₃-MeOH-H₂O); $[\alpha]_D^{25} - 5.4^\circ$ (*c* = 0.5, H₂O). ¹H-NMR (D₂O) δ : 4.87 (brs, 1H, Asp α), 4.84 (d, 1H, *J* = 3.7 Hz, H-1), 4.41 (d, 1H, *J* = 7.8 Hz, H-1'), 2.48 (m, 2H, Glu γ), 2.12 (s, 3H, Ac), 1.14 (brs, 3H, Thr γ). *Anal.* Calcd for C₃₅H₅₇N₇O₂₄: C, 43.80; H, 5.99; N, 10.21. Found: C, 43.58; H, 5.78; N, 10.46.

β -D-Galactopyranosyl-(1 \rightarrow 4)-(2-acetamido-2-deoxy- β -D-galactopyranosyl)-L-seryl-L-seryl-glycyl-L-aspartyl-L-glutamyl-L-threonine (32 β**)** A solution of compound **31 β** (12 mg, 0.0089 mmol) in 3:1 MeOH-H₂O (2 ml) was treated with NaOMe (6 mg) at room temperature for 1 h. The mixture was neutralized with Amberlite IR-120 (H⁺), filtered, and concentrated. The residue was chromatographed over Sephadex LH-20 with MeOH-H₂O (3:1) to provide **32 β** (6 mg, 71%). *Rf* 0.68 (1:3:1 CHCl₃-MeOH-H₂O); $[\alpha]_D^{25} - 5.4^\circ$ (*c* = 0.5, H₂O). ¹H-NMR (D₂O) δ : 4.85 (brs, 1H, Asp α), 4.45 (d, 1H, *J* = 7.9 Hz, H-1'), 4.28 (d, 1H, *J* = 7.8 Hz, H-1), 2.50 (m, 2H, Glu γ), 2.15 (s, 3H, Ac), 1.16 (brs, 3H, Thr γ). *Anal.* Calcd for C₃₅H₅₇N₇O₂₄: C, 43.80; H, 5.99; N, 10.21. Found: C, 43.58; H, 6.12; N, 9.99.

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References

- 1) a) Oku H., Shiraishi T., Ouchi S., *Naturwissenschaften*, **64**, 643 (1977); b) Shiraishi T., Oku H., Yamashita M., Ouchi S., *Ann. Phytopathol. Soc. Jpn.*, **44**, 659–665 (1978).
- 2) a) Darvill A.G., Albersheim P., *Annu. Rev. Plant Physiol.*, **35**, 243–275 (1984); b) Yamada T., Hashimoto H., Shiraishi T., Oku H., *Plant-Microbe Interact.*, **2**, 256–261 (1989).
- 3) Shiraishi T., Saitoh K., Kim H. M., Kato T., Tahara M., Oku H., Yamada T., Ichinose Y., *Plant Cell Physiol.*, **33**, 663–667 (1992).
- 4) Yoshioka H., Shiraishi T., Yamada T., Ichinose Y., Oku H., *Plant Cell Physiol.*, **31**, 1139–1146 (1990).
- 5) Toyoda K., Shiraishi T., Yoshioka H., Yamada T., Ichinose Y.,

- Oku H., *Plant Cell Physiol.*, **33**, 445—452 (1990).
- 6) Shioiri T., Yokoyama Y., Kasai Y., Yamada S., *Tetrahedron*, **32**, 2211—2217 (1976).
- 7) a) Belleau B., Malek G., *J. Am. Chem. Soc.*, **90**, 1651—1652 (1968);
b) Yajima H., Kawatani H., *Chem. Pharm. Bull.*, **19**, 1905—1908 (1971).
- 8) Grundler G., Schmidt R. R., *Liebigs Ann. Chem.*, 1826—1847 (1984).
- 9) Paulsen H., Sinnwell V., *Chem. Ber.*, **111**, 869—889 (1978).
- 10) Conchie J., Levvy G. A., *Methods in Carbohydr. Chem.*, **2**, 335—337 (1963).
- 11) Garegg P. J., Hultberg H., *Carbohydr. Res.*, **110**, 261—266 (1982).
- 12) a) Lemieux R. U., Takeda T., Chung B. Y., *Am. Chem. Soc. Symp. Ser.*, **39**, 90—115 (1976); b) Hanessian S., Banoub J., *Carbohydr. Res.*, **53**, c13—c16 (1977).
- 13) Bax A., Summers M. F., *J. Am. Chem. Soc.*, **108**, 2093—2094 (1986).
- 14) Lemieux R. U., Ratcliffe R. M., *Can. J. Chem.*, **57**, 1244—1251 (1979).