

## Constituents of the Leaves of *Aristolochia kaempferi*

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Two new phenanthrene derivatives, aristoliukine-A, -B, and a benzenoid, sodium (2*R*)-(p-hydroxyphenyl)lactate, together with thirty-three known compounds were isolated from the fresh leaves of *Aristolochia kaempferi*. Structures were elucidated by spectral analysis. Among the new compounds, sodium (2*R*)-(p-hydroxyphenyl)lactate was determined to be in the salt form by IR and <sup>1</sup>H-NMR methods.

**Key words** *Aristolochia kaempferi*; Aristolochiaceae; aristolactam; aristoliukine-A; aristoliukine-B

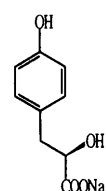
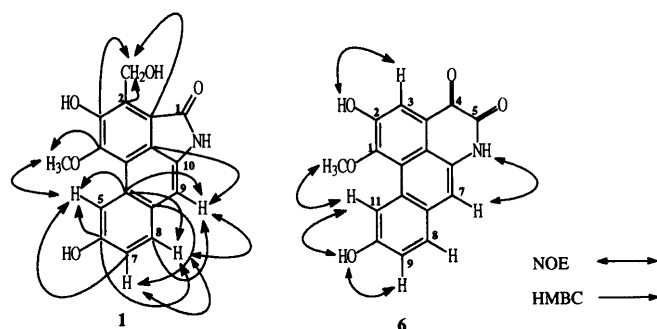
Plants of the genus *Aristolochia* (Aristolochiaceae) are known to number about 400 different species, and are found from the tropics to temperate zones. Five species are native to Taiwan, namely *Aristolochia cucurbitifolia*, *A. foveolata*, *A. heterophylla*, *A. zollingeriana* and *A. kaempferi*, and certain species have been used in folk medicine as anodynes, antiphlogistics and detoxicants.<sup>1)</sup> *A. kaempferi* (*A. liukiunensis*) is distributed in the southern Ryukyu islands and also in Taiwan,<sup>1)</sup> and several aristolochic acids and aristolactams have been isolated from this species.<sup>2–7)</sup> In this paper, we describe the isolation and structural elucidation of three new compounds, aristoliukine-A (**1**), aristoliukine-B (**6**), and sodium (2*R*)-(p-hydroxyphenyl)lactate (**17**), together with thirty-three known compounds from the leaves of *A. kaempferi*.

Aristololiukine-A (**1**) was isolated as yellowish needles, and exhibited an UV spectrum characteristic of a phenanthrene chromophore.<sup>8)</sup> The IR bands at 3380, 3300, 3170 and 1668 cm<sup>-1</sup> revealed the presence of hydroxyl OH, amido NH and lactam carbonyl groups. The <sup>1</sup>H-NMR spectrum of **1** showed the presence of an amide NH proton at δ 9.70 (1H, br s, exchangeable with D<sub>2</sub>O) and one methoxyl at δ 3.98 (3H, s). In the aromatic region, three mutually coupled signals at δ 8.46 (1H, d, *J*=2.8 Hz), 7.76 (1H, d, *J*=8.4 Hz) and 7.07 (1H, dd, *J*=8.4, 2.8 Hz) were attributed to H-5, H-8 and H-7, respectively. A singlet at δ 7.04 (1H, s) was assigned to H-9 and an oxygenated benzylic methylene signal appeared at δ 5.07 as a 2-proton singlet. This data resembled that of the 5,7,8,9-unsubstituted aristolactam. The methoxyl group was located at C-4 on the basis of a nuclear Overhauser effect (NOE) spectroscopy (NOESY) experiment, which showed NOE correlations between H-5 (δ 8.46) and the methoxyl signal (δ 3.98). The methylene group at C-2 was determined by heteronuclear multiple-bond correlation spectroscopy (HMBC) from correlations of the methylene protons at δ 5.07 with δ 119.3 (s, C-2), 131.4 (s, C-1a) and 149.6 (s, C-3). Based on these data, the structure **1** was assigned for aristoliukine-A.

Aristololiukine-B (**6**) was obtained as red needles. FAB-MS displayed a *pseudo*-molecular ion peak at *m/z* 310 (M<sup>+</sup>+H). The UV spectrum showed absorptions characteristic of a 4,5-dioxoaporphine derivative<sup>8)</sup> at 219, 235, 246, 275, 321, 332, 373 and 480 nm. IR revealed the presence of hydroxyl and amino groups between 3400 to 3100 cm<sup>-1</sup> and a strong carbonyl group absorption at 1676 cm<sup>-1</sup>. The <sup>1</sup>H-NMR spectrum of **6** showed three exchangeable proton signals at δ 11.93 (1H, s), 10.62 (1H, s) and 9.94 (1H, s) attributable to

amide NH and two phenolic hydroxyl protons. An ABX system for the aromatic protons was observed at δ 8.89 (1H, d, *J*=2.0 Hz) for H-11 and 7.76 (1H, d, *J*=8.8 Hz), 7.15 (1H, dd, *J*=8.8, 2.0 Hz) for H-8 and H-9, respectively. The remaining three singlet signals at δ 8.06 (1H), 7.41 (1H) and 4.08 (3H) could be assigned to H-3, H-7 and a methoxyl group. From this data, compound **6** was determined to be triangularine-A (1,10-dihydroxy-2-methoxy-4,5-dioxoaporphine).<sup>7)</sup> By comparison of their <sup>1</sup>H-NMR spectra, the positions of substituents were revealed. To confirm the position of the methoxyl, a NOESY experiment was conducted and the hydroxyl (δ 9.94) was found to be within NOE distance of H-11 (δ 8.89) and H-9 (δ 7.15), and the methoxyl group (δ 4.08) had NOE correlation with H-11 (δ 8.89), which indicated that the methoxyl must be located at C-1. Thus, structure **6** was proposed for aristoliukine-B.

Sodium (2*R*)-(p-hydroxyphenyl)lactate (**17**) was isolated as an optically active colorless powder. The UV spectrum showed absorptions at 225, 279 and 285 nm, which suggested it was a benzenoid. In the <sup>1</sup>H-NMR spectrum, a set of *ortho*-coupled protons appeared at δ 7.13 (2H, d, *J*=8.6 Hz) and 6.83 (2H, d, *J*=8.6 Hz) attributable to *para*-substituted aromatic protons. Three mutually coupled aliphatic protons at δ 3.82 (1H, dd, *J*=7.6, 5.2 Hz), 3.13 (1H, dd, *J*=14.4, 5.2 Hz)



**17**  
Chart 1

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and 2.93 (1H, dd,  $J=14.4, 7.6$  Hz) were assigned to the  $\alpha$  and  $\beta$ -protons of a  $\beta$ -hydroxyarylpropanoic acid moiety. The above data for compound **17** was similar to (*p*-hydroxyphenyl)lactic acid.<sup>9</sup> However, the IR spectrum of **17** displayed a carboxyl absorption at  $1590\text{ cm}^{-1}$ , indicative of a salt form. Acidification of **17** with HCl afforded sodium chloride, which was determined with an atomic absorption spectrometer and (2*R*)-(p-hydroxyphenyl)lactic acid ( $[\alpha]_D -13^\circ$ ,  $c=0.15$ , MeOH). On the basis of the above data, compound **17** was assigned to be sodium (2*R*)-(p-hydroxyphenyl)lactate.

In addition to **1**, **6** and **17**, cepharanone-A (**2**),<sup>10</sup> aristolactam-IIIa (**3**),<sup>11</sup> -AII (**4**),<sup>10</sup> -AIIIa (**5**),<sup>11</sup> 4,5-dioxodehydroasimilobine (**7**),<sup>12</sup> cepharadione-A (**8**),<sup>10</sup> aristolochic acid-I (**9**),<sup>10</sup> -II (**10**),<sup>10</sup> -IIIa (**11**),<sup>10</sup> -IVa (**12**),<sup>10</sup> sodium aristolochate-I (**13**),<sup>13</sup> aristolic acid (**14**),<sup>14</sup> aristofolin-A (**15**),<sup>7</sup> aristofolin-B (**16**),<sup>15</sup> *p*-hydroxybenzaldehyde (**18**),<sup>16</sup> benzoic acid (**19**),<sup>17</sup> *p*-hydroxybenzoic acid (**20**),<sup>18</sup> methylparaben (**21**),<sup>10</sup> vanillic acid (**22**),<sup>19</sup> methyl vanillate (**23**),<sup>10</sup> cinnamamide (**24**),<sup>20</sup> *p*-hydroxy cinnamic acid (**25**),<sup>10</sup> methyl *p*-hydroxy cinnamate (**26**),<sup>10</sup> ferulic acid (**27**),<sup>10</sup> methyl 3,4-dihydroxy cinnamate (**28**),<sup>21</sup> kaempferol (**29**),<sup>22</sup> kaempferol-3-*O*-glucoside (**30**),<sup>22</sup> tiliroside (**31**),<sup>23</sup> isorhamnetin 3-*p*-coumaroylglucoside (**32**),<sup>23</sup> 3-carboxy pyridine (**33**),<sup>24</sup> *N*-*p*-coumaroyltyramine (**34**),<sup>10</sup> pheophytin-a (**35**)<sup>25</sup> and  $\beta$ -sitossterol-3-*O*-glucoside (**36**)<sup>26</sup> were also isolated from the leaves of *A. kaempferi*. These known compounds were characterized by their spectral properties.

#### Experimental

UV spectra were recorded in MeOH, and IR spectra were determined as KBr discs. <sup>1</sup>H-NMR spectra were obtained on a Bruker NMR spectrometer, with tetramethylsilane (TMS) as internal standard. EI-MS was measured with a 70 eV direct inlet system on a VG70-250AS spectrometer. Melting points were uncorrected. Optical rotations were recorded on a Jasco DIP-370 digital polarimeter.

**Plant Material** *Aristolochia kaempferi* was collected at Nantou, Taiwan, in April, 1994 and verified by Prof. C.S. Kuoh. A voucher specimen is deposited in the Herbarium of Cheng Kung University, Taiwan.

**Extraction and Separation** The fresh leaves (1.3 kg) were extracted with MeOH at room temperature and concentrated to give a deep brown syrup (190 g). The MeOH extract was partitioned between H<sub>2</sub>O and CHCl<sub>3</sub>, and the H<sub>2</sub>O layer was filtered to give an H<sub>2</sub>O insoluble fraction and a H<sub>2</sub>O soluble layer. The CHCl<sub>3</sub> layer was directly chromatographed on silica gel column and eluted with CHCl<sub>3</sub> containing increasing proportions of MeOH (0→100%, stepwise elution with a 10% increase at each step) to give 9 fractions. Fraction 3 was rechromatographed on silica gel and eluted with *n*-C<sub>6</sub>H<sub>14</sub>:EtOAc (14:1) to obtain **19** (1 mg) and **35** (1 mg). Fraction 4 was also rechromatographed on silica gel with CHCl<sub>3</sub>:EtOAc (14:1) to give **2** (15 mg) and **8** (10 mg). Fraction 5 was treated in a similar way to fraction 4 to give **4** (3 mg), **5** (6 mg) and **24** (1 mg). Fraction 6 was also treated in the same manner as fraction 4 to obtain **7** (2 mg), **16** (0.5 mg), **21** (2 mg) and **36** (100 mg), respectively. Fraction 8 was filtered to give **9** (450 mg). The H<sub>2</sub>O soluble layer was chromatographed on Diaion HP-20 and eluted with H<sub>2</sub>O containing increasing proportions of MeOH (0→100%, stepwise elution with a 10% increase at each step) to give 9 fractions. Fraction 3 of the H<sub>2</sub>O soluble layer was filtered to give **17** (200 mg). Fraction 4 was chromatographed on silica gel with CHCl<sub>3</sub>:MeOH (9:1) to give **20** (4 mg), **22** (2 mg), **27** (3 mg) and **33** (5 mg). Fractions 5 and 7 were combined and rechromatographed on silica gel with CHCl<sub>3</sub>:EtOAc (7:1) to give **25** (10 mg) and **30** (30 mg). Fraction 6 was treated similarly to obtain **5** (2 mg), **15** (7 mg), **18** (1 mg), **21** (1 mg), **28** (1 mg), **31** (5 mg), **32** (2 mg) and **34** (2 mg). Fraction 8 was treated in a similar way as fraction 4 to give **3** (3 mg), **6** (1 mg), **10** (1.5 mg), **14** (7 mg), **26** (3 mg), **29** (1 mg) and **31** (1 mg), respectively.

The H<sub>2</sub>O insoluble portion was chromatographed on a Rp-18 column to give 13 fractions. Fraction 5 was rechromatographed on silica gel with CHCl<sub>3</sub>:MeOH (5:1) to obtain **11** (1.5 mg) and **23** (5 mg). Fraction 10 was treated in the same manner as fraction 5 to give **1** (7 mg), **5** (2 mg), **6** (1 mg)

and **12** (10 mg). Fractions 11 and 12 were combined and filtered to give **13** (5 mg).

**Aristolochine-A (1)**: Yellowish needles (MeOH), mp 274–276 °C. HR-FAB-MS (Pos.): Calcd for C<sub>17</sub>H<sub>14</sub>NO<sub>5</sub>,  $m/z$  312.0872 [M+H]<sup>+</sup>, Found 312.0876. UV  $\lambda_{\text{max}}$  nm: 239, 256, 280(sh), 321, 345(sh), 404. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 3380, 3300, 3170, 1668, 1651, 1377, 1238, 1224, 1147, 1043, 1001, 667. FAB-MS (Pos.)  $m/z$  (rel. int.): 312 (M<sup>+</sup>+1), 307 (24), 289 (11), 155 (29), 154 (100), 137 (70), 107 (23). <sup>1</sup>H-NMR (DMSO-*d*<sub>6</sub>, 400 MHz)  $\delta$ : 10.88 (1H, s, OH), 9.75 (1H, s, OH), 9.70 (1H, br s, NH), 8.46 (1H, d,  $J=2.8$  Hz, H-5), 7.76 (1H, d,  $J=8.8$  Hz, H-8), 7.07 (1H, dd,  $J=8.8, 2.8$  Hz, H-7), 7.04 (1H, s, H-9), 5.80 (1H, br s, OH), 5.07 (2H, s, CH<sub>2</sub>), 3.98 (3H, s, OCH<sub>3</sub>). <sup>13</sup>C-NMR (DMSO-*d*<sub>6</sub>, 100 MHz)  $\delta$ : 169.5 (C-1), 155.5 (C-2), 149.6 (C-3), 148.3 (C-4), 132.4 (C-10), 131.4 (C-1a), 130.2 (C-8), 127.4 (C-8a), 127.2 (C-9), 122.3 (C-10a), 119.3 (C-2), 118.9 (C-4a), 117.1 (C-7), 111.5 (C-5), 104.9 (C-9), 60.0 (OCH<sub>3</sub>), 56.1 (C-11).

**Aristolochine-B (6)**: Red needles; mp 249–250 °C (dec.); HR-FAB-MS (Pos.): Calcd for C<sub>17</sub>H<sub>12</sub>NO<sub>5</sub>,  $m/z$  310.0715 [M+H]<sup>+</sup>, Found 310.0723. UV  $\lambda_{\text{max}}$  nm: 219, 235, 246, 275(sh), 321(sh), 332, 373(sh) and 480 nm. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 3400-3100, 1676, 1569, 1524, 1400 and 1211. FAB-MS  $m/z$  (rel. int.): 310 (M<sup>+</sup>+1, 3), 289 (12), 219 (8), 154 (100), 137 (70), 124 (12), 107 (22). <sup>1</sup>H-NMR (DMSO-*d*<sub>6</sub>, 400 MHz)  $\delta$ : 11.93 (1H, s, NH), 10.62 (1H, s, OH), 9.94 (1H, s, OH), 8.89 (1H, d,  $J=2.0$  Hz, H-11), 8.06 (1H, s, H-3), 7.76 (1H, d,  $J=8.8$  Hz, H-8), 7.41 (1H, s, H-7), 7.15 (1H, dd,  $J=8.8, 2.0$  Hz, H-9), 4.08 (3H, s, OCH<sub>3</sub>).

**Sodium (2*R*)-3-(*p*-Hydroxyphenyl)lactate (17)**: Colorless powder (MeOH), mp >300 °C.  $[\alpha]_D -33^\circ$  ( $c=0.01$ , MeOH). UV  $\lambda_{\text{max}}$  nm: 225, 279, 285. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 3200, 1590, 1520, 1470, 1455, 1420, 1360, 1335, 1250. <sup>1</sup>H-NMR (D<sub>2</sub>O, 200 MHz)  $\delta$ : 7.13 (2H, d,  $J=8.6$  Hz, H-5,9), 6.83 (2H, d,  $J=8.6$  Hz, H-6,8), 3.82 (1H, dd,  $J=7.6, 5.2$  Hz, H-2), 3.13 (1H, dd,  $J=14.4, 5.2$  Hz, H-3), 2.93 (1H, dd,  $J=14.4, 7.6$  Hz, H-3).

Acidification of **17**: Compound **17** (2 mg) was dissolved in 5% HCl (aq) (1 ml). The solution was eluted in a Sephadex LH-20 column with H<sub>2</sub>O, then MeOH, to afford NaCl (0.5 mg) and (2*R*)-3-(*p*-hydroxyphenyl)lactic acid (1.6 mg) ( $[\alpha]_D -13^\circ$ ,  $c=0.15$ , MeOH).

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