## A New Oleanene Glucuronide Having a Branched-Chain Sugar from *Melilotus officinalis*<sup>1)</sup>

Manabu Udayama, Junei Kinjo, Naotoshi Yoshida, and Toshihiro Nohara\*, a

Faculty of Pharmaceutical Sciences, Kumamoto University,<sup>a</sup> 5-1 Oe-honmachi, Kumamoto 862–0973, Japan and Medicinal Plant Garden, Faculty of Pharmaceutical Sciences, Hokkaido University,<sup>b</sup> Nishi 6, Kita 12, Kita-ku, Sapporo 060–0812 Japan. Received September 19, 1997; accepted November 5, 1997

A new oleanene glucuronide called melilotus-saponin  $O_1$  (1) was isolated together with three known ones from the roots of *Melilotus officinalis* (L.) Pallas (Leguminosae). The structure of 1 was determined to be 3-O- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 2)$ - $\alpha$ -L-arabinopyranosyl- $(1 \rightarrow 3)$ ]- $\beta$ -D-galactopyranosyl- $(1 \rightarrow 2)$ - $\beta$ -D-glucuronopyranosyl soyasapogenol B by spectroscopic and chemical methods.

Melilotus officinalis (L.) PALLAS is distributed worldwide and is known in English as common yellow melilot or medicinal sweet clover.<sup>2)</sup> This plant is used not only as a food and forage but also as a medicine. The preventive effect of its extract on experimental atherosclerosis in rabbits was reported.3) The effect of a medical preparation (Esberiven) using its extract was also evaluated on dermatological disease. 4) Earlier researchers found that the extract of aerial parts showed potent inhibitory activity on leucocyte migration and one of the constituents responsible for the action was azukisaponin V.5) During our course of studies on leguminous plants, 1) we have investigated the oleanene-type triterpene glucuronides (oleanene glucuronides) of the roots of Japanese Melilotus officinalis. This paper deals with the structural elucidation and identification of these oleanene glucuronides.

A methanolic extract of the aerial parts of M. officinalis was first separated by Sephadex LH-20 column chromatography to get a crude saponin fraction. A combination of MCI gel and silica gel chromatographies resulted in the isolation of four saponins (1—4). Saponins 2—4 were identified as soyasaponin I (2),<sup>6)</sup> dehydrosoyasaponin I (3),<sup>6b,7)</sup> and acetyl-soyasaponin I (4)<sup>8)</sup> by direct comparison with the authentic samples.

Melilotus-saponin  $O_1$  (1) was obtained as a white amorphous powder,  $[\alpha]_D^{25} + 5.7^{\circ}$  (MeOH). In the negative FAB-MS, 1 showed an  $[M-H]^-$  ion at m/z 1073. Fragment ion peaks at m/z 941 [M-pentose] and 927 [M-methylpentose] were also observed. The exact measurement under high resolution (HR) conditions showed that the composition is  $C_{53}H_{86}NaO_{22}$  at m/z $1097.5519 [M+Na]^+$  in the HR/positive FAB-MS. By acid hydrolysis, 1 gave soyasapogenol B as the sapogenol. The monosaccharide mixture obtained by acid hydrolysis revealed the presence of glucuronic acid, galactose, rhamnose and arabinose by TLC. Their absolute configurations were determined to be the D-form (glucuronic acid, galactose) and the L-form (arabinose, rhamnose). according to the procedure developed by Hara et al.9) In the sugar region of the <sup>13</sup>C-NMR spectrum for 1, signals based upon the terminal rhamnopyranosyl and the terminal arabinopyranosyl residues were observed. Since the carbon signals due to the sapogenol moiety were superimposable on those of 2,6b these sugars were

concluded to be composed of a branched-chain sugar which attached at C-3. The combination analyses of  ${}^{1}H^{-1}H$  shift correlation spectroscopy (COSY), heteronuclear multiple bond correlation (HMBC) and heteronuclear multiple quantum coherence (HMQC) spectra of 1 gave the correlations shown in Fig. 1.

Consequently, the structure of **1** was determined to be  $3-O-\alpha-L$ -rhamnopyranosyl- $(1\rightarrow 2)-\alpha-L$ -arabinopyranosyl- $(1\rightarrow 3)$ ]- $\beta$ -D-galactopyranosyl- $(1\rightarrow 2)$ - $\beta$ -D-glucuronopyranosyl soyasapogenol B.

Meanwhile, we clarified that oleanene glucuronides are effective for experimental hepatitis. <sup>10)</sup> Since Kang *et al.* previously reported the potent inhibitory activity on leucocyte migration of oleanene glucuronides obtained from the titled plant, <sup>5)</sup> some of these could show anti-inflammatory actions on not only hepatitis but various other inflammations.

## Experimental

The instruments and reagents used in this study were the same as those described.  $^{8)}$ 

Extraction and Isolation The dried roots (580 g) of Melilotus officinalis collected in the medicinal garden of Hokkaido University were extracted with MeOH, and the extract (18 g) was separated by Sephadex LH-20 column chromatography to give crude saponin fraction. After MCI gel CHP 20P column chromatography using 50%  $\rightarrow$ 100% MeOH to afford fractions 1 to 9, fractions 2, 3, 5 and 8 were separated by silica gel

HOOC 
$$Glc\ A\ OH$$

$$HO\ OR_1$$

$$Gal$$

$$1: ara \quad \beta\text{-OH}, \alpha\text{-H} \quad melilotus\text{-saponin } O_1$$

$$2: \quad H \quad \beta\text{-OH}, \alpha\text{-H} \quad soyasaponin I \quad dehydrosoyasaponin I \quad acetyl\text{-soyasaponin I}$$

$$4: \quad H \quad \beta\text{-OAc}, \alpha\text{-H} \quad acetyl\text{-soyasaponin I}$$

\* To whom correspondence should be addressed.

© 1998 Pharmaceutical Society of Japan

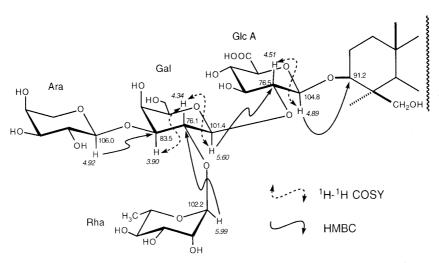


Fig. 1. <sup>1</sup>H-<sup>1</sup>H COSY and HMBC Connectivities for Sugar Moiety of 1

 $(CHCl_3: MeOH: H_2O=6:4:1)$  to provide compounds 1 (0.005%), 2 (0.02%), 3 (0.0009%) and 4 (0.0009%), respectively.

**Compound 1 (Melilotus-Saponin O<sub>1</sub>)** A white amorphous powder,  $[\alpha]_D^{25} + 5.7^\circ (c = 0.50, \text{MeOH})$ . HR positive ion FAB-MS m/z: 1097.5519 ( $C_{53}H_{86}\text{NaO}_{22}$ , Calcd for 1097.5510). Negative ion FAB-MS m/z: 1073  $[M-H]^-$ , 941  $[M-H-Ara]^-$ , 927  $[M-H-Rha]^-$ . <sup>1</sup>H-NMR (in pyridine- $d_5$ ): 0.72, 0.96, 1.01, 1.22, 1.26, 1.26, 1.44 (each 3H, s, *tert-Me* × 7), 1.72 (3H, d, J = 4.3 Hz, Rha  $H_3$ -6), 4.89 (1H, d, J = 5.9 Hz, Glc A H-1), 4.92 (1H, d, J = 6.7 Hz, Ara H-1), 5.30 (1H, s, H-12), 5.60 (1H, d, J = 7.7 Hz, Gal H-1), 5.99 (1H, s, Rha H-1). <sup>13</sup>C-NMR (in pyridine- $d_5$ ): 38.6, 26.3, 91.2, 43.7, 56.0, 18.4, 33.0, 39.7, 47.6, 36.3, 23.9, 122.4, 144.5, 42.2, 26.1, 28.7, 37.8, 45.4, 46.6, 30.6, 41.8, 76.1, 22.8, 63.3, 15.7, 16.9, 25.3, 20.7, 32.8, 28.6 (C-1—30), 104.8, 76.5, 73.5, 73.9, 77.9, 175.3 (Glc A C-1—6), 101.4, 76.1, 83.5, 68.7, 76.5, 61.5 (Gal C-1—6), 102.2, 71.8, 72.0, 73.7, 69.0, 18.6 (Rha C-1—6), 106.0, 71.9, 75.9, 70.6, 66.1 (Ara C-1—5).

Characterization of Sapogenol and Sugars for 1 A small amount of 1 was dissolved in 2 n HCl/H<sub>2</sub>O (2 ml) and heated at 90 °C for 2 h. After addition of CHCl<sub>3</sub>, the organic layer was identified to be soyasapogenol B<sup>11)</sup> by TLC. R/s: 0.34 [CHCl<sub>3</sub>–MeOH (19:1)], 0.48 [n-hexane–acetone (2:1)]. The aqueous layer was neutralized with 2 n KOH/H<sub>2</sub>O. The sugar mixture was subjected to TLC analysis [TLC, Kieselgel 60 F<sub>254</sub> (Merck Art 5554), n-PrOH: acetone: H<sub>2</sub>O = 5:3:1, R/s: 0.06 (glucuronic acid), 0.44 (galactose), 0.58 (arabinose), 0.79 (rhamnose).

**D, L Determination of Sugars of 1** A small amount of **1** was methylated with ethereal  $CH_2N_2$ . To a solution of the methylated sample of **1** was added  $NaBH_4$ , and the mixture was kept at room temperature for 30 min. The reaction mixture was worked up with MCI gel CHP 20P. The MeOH eluate was evaporated and heated in  $2 \,\mathrm{N}$  HCl/ $H_2O$  at  $90\,^{\circ}C$  for  $3 \,\mathrm{h}$ . The hydrolysate was subjected to MCI gel CHP 20P and Amberlite IRA-400 to give a sugar fraction. This fraction was dissolved in pyridine (0.1 ml), then the solution was added to a pyridine solution (0.2 ml) of L-cysteine methyl ester hydrochloride (0.1 mol/l) and warmed at  $60\,^{\circ}C$  for  $2 \,\mathrm{h}$ . The solvent was evaporated under  $N_2$  stream and dried *in vacuo*. The remaining syrup was trimethylsilylated with trimethylsilylimidazole (0.1 ml) at  $60\,^{\circ}C$  for  $1 \,\mathrm{h}$ . After addition of *n*-hexane and  $H_2O$ , the *n*-hexane layer was taken out and checked by GC. The retention times ( $t_R$ ) of the peaks were  $15.9 \,\mathrm{min}$  (D-glucose),  $9.0 \,\mathrm{min}$  (L-arabinose),  $10.8 \,\mathrm{min}$  (L-rhamnose) and  $16.9 \,\mathrm{min}$  (D-galactose).

**Soyasaponin I (2)**<sup>6)</sup> White amorphous powder,  $[\alpha]_D^{25} - 12.0^{\circ}$  [c = 0.5, MeOH]. Positive ion FAB-MS m/z: 965 [M+Na]<sup>+</sup>. HPLC, conditions see ref. 6b, ( $t_R$ : 31.4 min) and TLC, Kieselgel 60 F<sub>254</sub> (Merck Art 5554), CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O (6:4:1), Rf: 0.48; n-BuOH–AcOH–H<sub>2</sub>O (4:1:5, upper), Rf: 0.31].

**Dehydrosoyasaponin I (3)**<sup>7)</sup> White amorphous powder,  $[\alpha]_D^{25} - 11.7^{\circ}$  [c = 0.5, MeOH]. Negative ion FAB-MS m/z: 939 [M-H]<sup>-</sup>. HPLC, conditions see ref. 6b, ( $t_R$ : 33.5 min) and TLC, Kieselgel 60 F<sub>2.54</sub> (Merck Art 5554), CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O (6:4:1), Rf: 0.52; n-BuOH-AcOH-H<sub>2</sub>O

(4:1:5, upper), Rf: 0.30].

Acetyl-Soyasaponin I (4)<sup>8)</sup> White amorphous powder,  $[\alpha]_D^{25} - 3.4^{\circ}$  [c = 0.5, MeOH]. Negative ion FAB-MS m/z: 983 [M-H]<sup>-</sup>. HPLC, conditions see ref. 6b, ( $t_R$ : 40.1 min) and TLC, Kieselgel 60 F<sub>254</sub> (Merck Art 5554), CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O (6:4:1), Rf: 0.54; n-BuOH-AcOH-H<sub>2</sub>O (4:1:5, upper), Rf: 0.30].

**Acknowledgements** We are grateful to Prof. H. Okabe and Mr. H. Hanazono of the Faculty of Pharmaceutical Sciences, Fukuoka University for measurement of the HR-MS.

## References and Notes

- 1) Part 58 in a series of studies on leguminous plants.
- Bisby F. A., Buckingham J., Harborne J. B. (ed.), "Phytochemical Dictionary of the Leguminosae," Chapman & Hall, London, 1994, pp. 472—474.
- Sakamoto W., Ohashi K., Nishikaze O., Oyo Yakuri, 15, 1—14 (1978).
- Okuma M., Yakuri To Chiryo, 10, 581—584 (1978); Higo S., Nakamura Y., Kawasaki M., Oyo Yakuri, 15, 231—240 (1978).
- Kang S. S., Lim C.-H., Lee S. Y., Arch. Pharm, Res., 10, 9—13 (1987); Kang S. S., Lee Y. S., Lee E. B., Kor. J. Pharmacognosy, 18, 89—93 (1987); Kang S. S., Woo W. S., J. Nat. Prod., 51, 335—338 (1988); Kang S. S., Lee Y. S., Lee E. B., Arch. Pharm, Res., 11, 197—202 (1988).
- a) Kitagawa I., Wang H. K., Taniyama T., Yoshikawa M., Chem. Pharm. Bull., 36, 153—161 (1988);
   b) Kinjo J., Kishida F., Watanabe K., Hashimoto F., Nohara T., ibid., 42, 1874—1878 (1994).
- Kitagawa I., Taniyama T., Murakami T., Yoshihara M., Yoshikawa M., Yakugaku Zasshi, 108, 547—554 (1988).
- Arao T., Idzu T., Kinjo J., Nohara T., Isobe R., Chem. Pharm. Bull., 44, 1970—1972 (1996).
- Hara S., Okabe H., Mihashi K., Chem. Pharm. Bull., 35, 501—506 (1987).
- Niiho Y., Yamasaki T., Nakajima Y., Itoh H., Takeshita T., Kinjo J., Nohara T., Yakugaku Zasshi, 109, 424—431 (1989); idem, ibid., 110, 604—611 (1990); Takeshita T., Ito Y., Sakai Y., Nohara T., Yasuhara M., Saito H., Kitagawa I., Ariga T., Irino N., Takaoka T., J. Pharmacobio-Dyn., 13, s-54 (1990); Kinjo J., Natural Medicines, 50, 79—85 (1996); Miyao H., Arao T., Udayama M., Kinjo J., Nohara T., Planta Med., in press; Arao T., Udayama M., Kinjo J., Funakoshi T., Kojima S., Nohara T., Biol. Pharm. Bull., 20, 988—991 (1997).
- Kinjo J., Miyamoto I., Murakami K., Kida K., Tomimatu T., Yamazaki M., Nohara T., Chem. Pharm. Bull., 33, 1293—1296 (1985).