

Synthesis of Four Possible Intermediates after Secologanin on the Biosynthesis of the Oleoside-, 10-Hydroxyoleoside- and Ligustaloside-Type Glucosides in Oleaceous Plants^{1a,b)}

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Received November 18, 1997; accepted February 16, 1998

To examine the biosynthetic pathway from secologanin to three types of secoiridoid glucosides characteristic of oleaceous plants, synthesis of two respective stereoisomers at C-8 of 8,10-epoxysecologanin and -secoxyloganin was established and their deuterium-labeled analogues were prepared.

Key words synthesis; epoxysecologanin; epoxysecoxyloganin; oleoside; 10-hydroxyoleoside; ligustaloside

Oleaceous plants contain a wide variety of secoiridoid glucosides,²⁾ e.g. oleuropein (**1**),^{3,4)} jasminin (**2**)⁵⁾ (oleoside-type), and 10-acetoxyoleuropein (**3**)⁶⁾ (10-hydroxyoleoside-type). In the course of our continuing examinations of the iridoid glucosides of oleaceous plants, we have isolated, along with 10-hydroxyoleuropein (**4**), ligustalosides A (**5**) and B (**6**)⁷⁾ which are secoiridoid glucosides with an aldehyde group at C-10 (ligustaloside-type). The systematic explanation of the biosynthesis of these three types of glucosides led us propose a pathway from secologanin (**7**) via epoxide (**8a**) as depicted in Chart 1. The configuration at C-8 of epoxide (**8a**) was presumed to be based on the *E* configuration of the ethylidene group in oleoside-type glucosides, and this was in agreement with the configuration formed by *trans* elimination of the corresponding (8*S*)-hydroxy derivative in the chemical reaction.⁴⁾ Another important clue for the stereochemistry was provided by the isolation of 8-epikingside (**9**) with the same configuration (8*S*) as that of **8a** from a few plants of this family.⁸⁾ Since oleaceous secoiridoid glucosides usually have a carboxy group at C-7 which forms an ester linkage with various alcohols, however, carboxylic acid (**10a**), the oxidation derivative at C-7 of the epoxide (**8a**) also was assumed to be another putative intermediate in this pathway.

Based on the above presumption, the (*S*)-isomer **8a** or **10a** seemed to best satisfy the stereochemical requirement. However, in order to examine the stereospecificity of this pathway, the stereoisomers (**8b**, **10b**) were also prepared to be used for the feeding experiments.

We report here the syntheses of (8*S*)-8,10-epoxysecologanin (**8a**) and its oxidative derivative at C-7, (8*S*)-8,10-epoxysecoxyloganin (**10a**), and their (8*R*)-isomers (**8b**, **10b**). The preparation of their deuterium-labeled analogues is described.

Synthesis of 8,10-Epoxysecologanin (8) and 8,10-Epoxysecoxyloganin (10) Taking into consideration the presence of the epoxy ring and the glucosidic linkage in the target compounds (**8**, **10**), *o*-nitrophenylethylene acetal⁹⁾ which could be removed readily by light irradiation under neutral condition was selected for the protection of the aldehyde group in secologanin tetraacetate (**7a**). Protection of the aldehyde in **7a** with *o*-nitrophenylethylene glycol

gave a mixture of two stereoisomeric acetals **11**. Based on the intensity of the anomeric signals of the glucose moiety observed at δ 4.85 (1H, *J*=8.0 Hz) and 4.86 (1H, *J*=8.0 Hz) in the ¹H-NMR spectrum of **11**, the ratio of the two isomers¹⁰⁾ in this mixture was estimated to be 1 : 1. The mixture was used for further reaction without separation for a series of reactions leading to epoxide (**15**). This mixture was subjected to oxidation with OsO₄ to give diol **12**, which was then tosylated to afford tosylate **13**. Treatment of **13** with 1,5-diazabicyclo[4.3.0]non-5-ene (DBN) gave a separable mixture of two stereoisomeric epoxides (**14a**, **14b**) in a ratio of 1 : 1. Deprotection of the two compounds by photolysis yielded aldehydes **15a** and **15b**, respectively. The configurations at C-8 of these two isomers could not be determined from their ¹H-NMR spectra, so we tried to solve the problem by elucidating

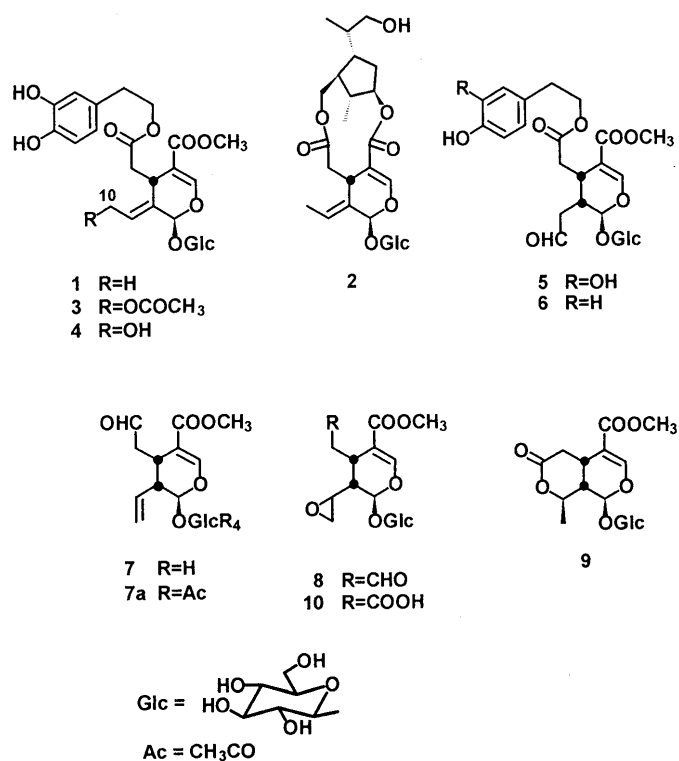


Fig. 1

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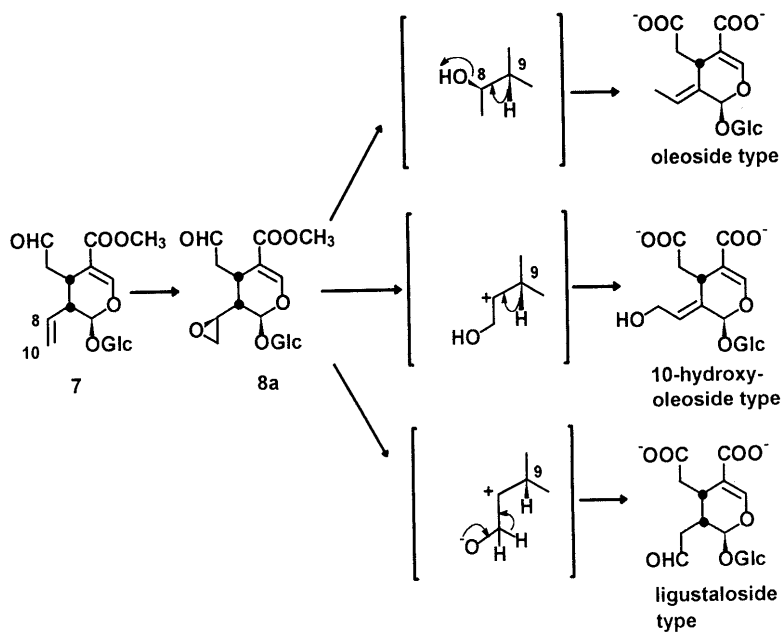
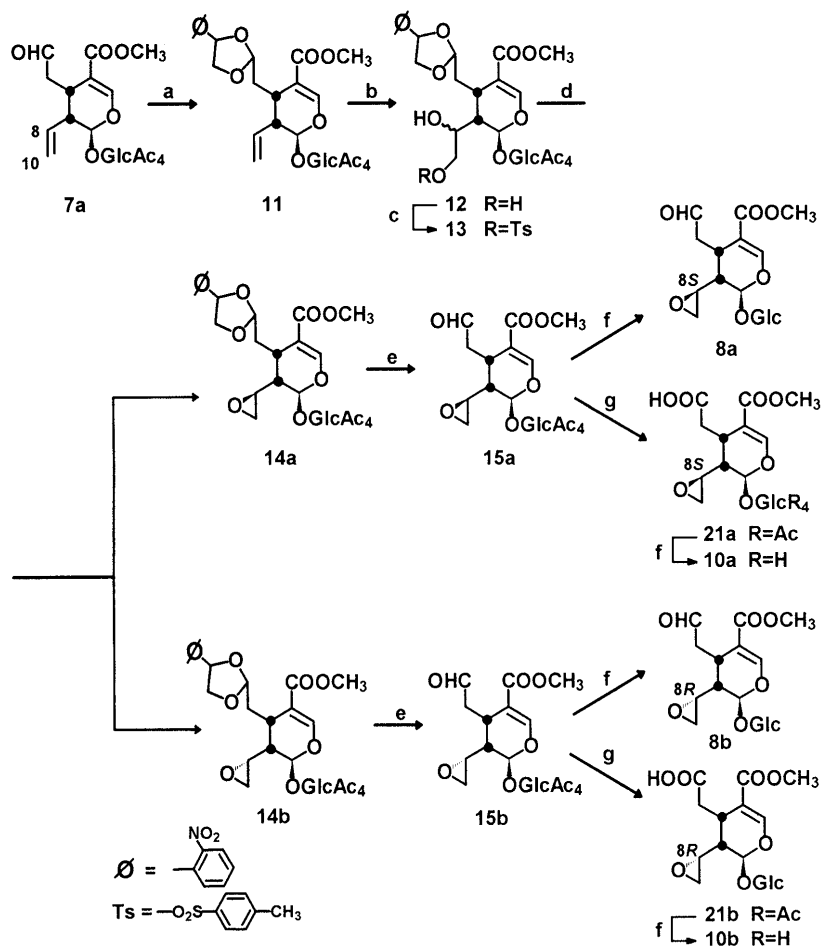
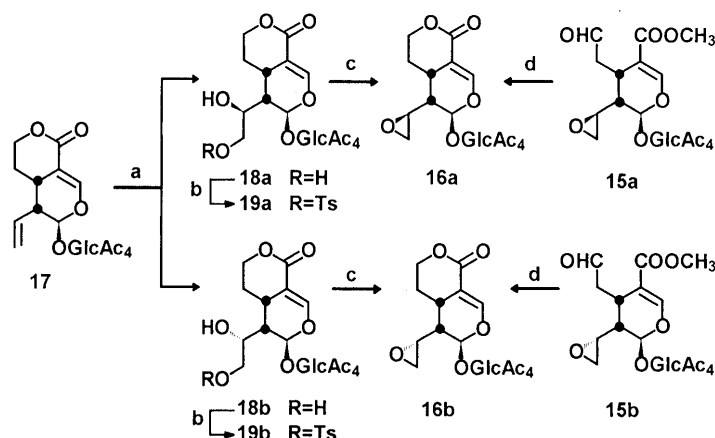


Chart 1



a) *o*-nitrophenylethylene glycol, *p*-TsOH / benzene; b) i: OsO₄, *N*-methylmorpholine-*N*-oxide / acetone-*tert*-BuOH-H₂O, ii: aqueous NaHSO₃; c) *p*-TsCl, molecular sieves / pyridine
d) DBN / benzene; e) hv / benzene; f) NaOMe / MeOH; g) Jones reagent / acetone

Chart 2



a) i: OsO₄, *N*-methylmorpholine-*N*-oxide / acetone-*tert*-BuOH-H₂O, ii: aqueous NaHSO₃;
 b) *p*-TsCl, molecular sieves / pyridine; c) DBN / benzene; d) NaBH₄ / EtOH, 0 °C

Chart 3

the relationship of the two isomers **15a** and **15b** to two epimers (**16a**, **16b**) of 8,10-epoxyswersoside tetraacetates,^{11,12} in which the stereochemistry at C-8 was established. Conversion of **15a** and **15b** into **16a** and **16b** was achieved by the reduction with NaBH₄, respectively.

The authentic samples **16a** and **16b** were prepared from sweroside tetraacetate (**17**). Compound **17** was dihydroxylated with OsO₄ to give a separable mixture of epimeric diols **18a** and **18b**.¹³ **18a** was converted to monotosylate **19a** which was treated with DBN to give the epoxide **16a** as colorless needles, mp 158–159 °C, whose chemophysical data were in agreement with those reported previously on (8*S*)-8,10-epoxyswersoside tetraacetate.¹² **18b** was derived in the same way *via* tosylate **19b** to epoxide **16b** (colorless needles, mp 199–200 °C), the chemophysical data of which were identical with those reported on (8*R*)-8,10-epoxyswersoside tetraacetate.^{11,12} Thus, it was concluded that **15a** was (8*S*)-8,10-epoxysecologanin tetraacetate and **15b** was its (8*R*)-epimer. Zemplén reaction of **15a** and **15b** gave the target compounds **8a** and **8b**, respectively. Further, Jones oxidation of **15a** and **15b** followed by Zemplén reaction gave (8*S*)-8,10-epoxysecoxyloganin (**10a**) and (8*R*)-8,10-epoxysecoxyloganin (**10b**), respectively.

Synthesis of [8-²H,10]-Epoxysecologanin (8) and [8-²H,10]-Epoxysecoxyloganin (10) As a preliminary experiment of the radioisotopic labelling, attempts to prepare compounds **15a** and **15b** labeled with deuterium on C-8 were made. The ketone **20**, which was obtained by Jones oxidation of tosylate **13**, was subjected to NaB²H₄ reduction in dioxane to afford [8-²H]-tosylate **13**. This compound was then treated with DBN followed by photolysis as described above to give [8-²H]-epoxyaldehydes (**15a**, **15b**) in a ratio of 2:1. In the ¹H-NMR spectrum of [8-²H]-**15a** and **15b**, three signals due to H-9, H₂-10 of each compound lost the coupling with H-8 (see Experimental), indicating deuterium-labeling at C-8. Accordingly, the synthetic method of 8-labeled 8,10-epoxides was established. Syntheses of tritium labeled epoxides **8a**, **8b**, **10a** and **10b**, and experiments on their administration

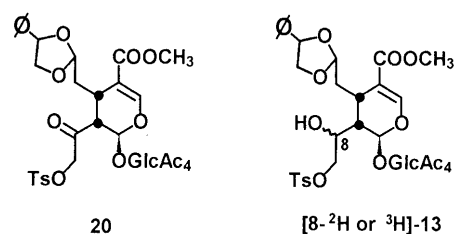


Fig. 2

will be reported elsewhere.

Experimental

Melting points were measured on Yanagimoto microapparatus and are uncorrected. Optical rotations were taken with a Union PM 101 automatic digital polarimeter. UV spectra were recorded on a Hitachi Model 323 S spectrophotometer, and IR spectra on a Hitachi Model 260-30 IR spectrophotometer. NMR spectra were taken on a JEOL JNM FX-200 unless otherwise noted or JEOL JNM-GX 500 with tetramethylsilane as the internal standard. FAB-MS spectra were recorded on a JEOL JMS-HX 100 mass spectrometer using 3-nitro-benzyl alcohol unless otherwise noted or glycerol as the matrix. Column chromatography was performed on Silica gel 60 (70–230 mesh, Merck) and on highly porous polymer Diaion HP-21 (Mitsubishi Chemical Industries). For TLC, Silica gel 60 GF₂₅₄ (Merck) was used and spots were visualized by irradiation under UV light, or by exposure to I₂ vapor. For preparative TLC (PTLC), Silica gel 60 PF₂₅₄ (Merck) was used and bands were detected under UV light. The ratios of solvents are expressed by volume. All extracts were dried over anhydrous MgSO₄.

Acetal 11 *p*-TsOH (230 mg) was added to a solution of **7a** (4.4 g), which was prepared from secologanin (**7**) with pyridine-Ac₂O, and *o*-nitrophenylethylene glycol (1.5 g) in dry benzene (500 ml), and the mixture was heated under reflux for 2 h with a Dean-Stark separator. It was then washed successively with saturated aqueous NaHCO₃ and brine, dried and concentrated *in vacuo*. The residue (5.5 g) was chromatographed on silica gel (66 g) column with CHCl₃ as an eluent, and 200 ml of each fraction was collected. Fractions 2–4 were combined and concentrated *in vacuo* to give acetal **11** (4.3 g, 75%) as a pale yellow powder. UV λ_{max}^{EtOH} nm (log ε): 228 (4.16), 262 inf (3.83). IR ν_{max}^{KBr} cm⁻¹: 1760, 1710, 1630, 1530. ¹H-NMR (CDCl₃) δ: 1.91–2.10 (12H, each s, -OCOCH₃), 2.40–2.60 (2H, m, H₂-6), 2.80–2.88 (1H, m, H-9), 2.88–3.18 (1H, m, H-5), 3.60–3.84 (5H, m, 5'-H, -CO₂CH₃, -C₆H₄CHCHH-O-), 4.15–4.80 (3H, m, H₂-6', -C₆H₄CHCHH-O-), 4.80, 4.85 (1H, each d, *J* = 8.0, H-1'), 4.95–5.80 (9H, m, H-1, -7, -8, -2', -3', -4', H₂-10, -C₆H₄CH-O-), 7.43–8.10 (4H, m, arom. H), 7.38, 7.39 (1H, each s, H-3). FAB-MS *m/z*: 722 (M+H)⁺. Anal. Calcd for C₃₃H₃₉NO₁₇: C,

54.92; H, 5.45; N, 1.94. Found: C, 54.94; H, 5.42; N, 1.89.

Diol 12 A solution of *N*-methylmorpholine-*N*-oxide·2H₂O (160 mg) in H₂O (10 ml) was added to a solution of **11** (730 mg) in acetone (50 ml) under argon. To the stirred solution was added dropwise a solution of OsO₄ (30 mg) in *tert*-BuOH (1 ml) and the mixture was stirred for 96 h at room temperature in the dark. After addition of 10% aqueous NaHSO₃ (1 ml), stirring was continued for an additional 1 h at room temperature. The solution was then diluted with H₂O (50 ml), neutralized with 1 N HCl, and extracted with CHCl₃ (50 ml × 4). The CHCl₃ layer was washed successively with brine, H₂O and dried. Removal of the solvent *in vacuo* gave the residue (633 mg), which was chromatographed on silica gel (20 g) column, and eluted successively with benzene-Et₂O (9:1) (150 ml) and Et₂O (100 ml). Fractions of 10 ml were collected. Fractions 14–33 were combined and concentrated *in vacuo* to give diol **12** (234.8 mg, 31%) as a white powder. UV λ_{max}^{EtOH} nm (log ε): 235 (4.19), 264 inf (3.80). IR ν_{max}^{KBr} cm⁻¹: 3480, 1755, 1705, 1640, 1530. ¹H-NMR (CDCl₃) δ: 1.70–1.98 (2H, m, H₂-6), 2.01–2.10 (12H, each s, -OCOCH₃), 2.26–2.64 (1H, m, H-9), 3.18–3.30 (1H, m, H-5), 3.58–4.02 (8H, m, H-8, -5', H₂-10, -CO₂CH₃, -C₆H₄CHCHH-O-), 5.02–5.64 (6H, m, H-1, -7, -2', -3', -4', -C₆H₄CH-O-), 4.10–4.80 (3H, m, H₂-6', -C₆H₄CHCHH-O-), 4.94, 4.96 (1H, each d, J = 8.0 Hz, H-1'), 7.41, 7.42 (1H, each s, H-3), 7.43–8.08 (4H, m, arom. H). FAB-MS *m/z*: 756 (M+H)⁺. Anal. Calcd for C₃₃H₄₁NO₁₉: C, 52.45; H, 5.47; N, 1.85. Found: C, 52.32; H, 5.42; N, 1.89. Fractions 1–3 were identified as the starting material (267 mg).

Tosylate 13 *p*-TsCl (130 mg) and freshly activated molecular sieves type 4A (500 mg) were added to a solution of **12** (497 mg) in dry pyridine (20 ml). After stirring for 40 h at room temperature in the dark, the mixture was filtered through a Celite layer, and the layer was washed with CHCl₃ (75 ml). The combined filtrate and washings were diluted with H₂O (100 ml), and extracted with CHCl₃ (100 ml × 4). The CHCl₃ layer was washed successively with 1 N HCl, saturated aqueous NaHCO₃ and H₂O, dried and concentrated *in vacuo*. The resulting residue (422 mg) was subjected to PTLC (CHCl₃:MeOH = 50:1, 3 developments) to give tosylate **13** (413.8 mg, 69%) as a white powder. UV λ_{max}^{EtOH} nm (log ε): 226 (4.37), 271 sh (3.60). IR ν_{max}^{KBr} cm⁻¹: 3480, 1755, 1705, 1640. ¹H-NMR (CDCl₃) δ: 1.60–1.98 (2H, m, H₂-6), 2.02–2.10 (12H, each s, -OCOCH₃), 2.24–2.63 (4H, m, H-9, arom. CH₃), 3.02–3.18 (1H, m, H-5), 3.47–3.82 (5H, m, H-5', -CO₂CH₃, -C₆H₄CHCHH-O-), 4.01–4.78 (6H, m, H-8, H₂-10, -6', -C₆H₄CHCHH-O-), 4.94–5.63 (7H, m, H-1, -7, -1', -2', -3', -4', -C₆H₄CH-O-), 7.38, 7.40 (1H, each s, H-3), 7.24–8.18 (8H, m, arom. H). FAB-MS *m/z*: 910 (M+H)⁺. Anal. Calcd for C₄₀H₄₇NO₂₁S: C, 52.80; H, 5.21; N, 1.54. Found: C, 52.69; H, 5.19; N, 1.60.

Epoxides 14a and 14b To a solution of **13** (79.2 mg) in dry benzene (5 ml) was added DBN (0.059 ml) and the mixture was stirred for 20 h at room temperature in the dark. After successive washings with 1 N HCl, brine and H₂O, the benzene layer was dried and concentrated *in vacuo* to give the residue (64.1 mg), which was subjected to PTLC (benzene:EtOAc = 5:1, 8 developments). Of the three bands, the least polar one yielded epoxide **14b** (19.5 mg, 30%) and the second polar one gave **14a** (20.2 mg, 32%) as a white powder, respectively. The most polar one was identified as the starting material (1.3 mg).

14a: UV λ_{max}^{EtOH} nm (log ε): 228 (4.13), 260 inf (3.76). IR ν_{max}^{KBr} cm⁻¹: 1760, 1710, 1630, 1530. ¹H-NMR (CDCl₃) δ: 1.60–1.90 (2H, m, H₂-6), 2.00–2.10 (12H, each s, -OCOCH₃), 2.48–2.90 (3H, m, H-8, H₂-10), 3.10–3.40 (2H, m, H-5, -9), 3.60–3.88 (5H, m, H-5', -CO₂CH₃, -C₆H₄CHCHH-O-), 4.00–4.84 (3H, m, H₂-6', -C₆H₄CHCHH-O-), 4.95–5.75 (7H, m, H-1, -7, -1', -2', -3', -4', -C₆H₄CH-O-), 7.30–8.16 (5H, m, H-3, arom. H). FAB-MS *m/z*: 738 (M+H)⁺. Anal. Calcd for C₃₃H₃₉NO₁₈: C, 53.73; H, 5.33; N, 1.90. Found: C, 53.65; H, 5.30; N, 1.91.

14b: UV λ_{max}^{EtOH} nm (log ε): 227 (4.13), 258 sh (3.75). IR ν_{max}^{KBr} cm⁻¹: 1755, 1710, 1630, 1530. ¹H-NMR (CDCl₃) δ: 1.50–1.80 (2H, m, H₂-6), 1.92–2.10 (12H, each s, -OCOCH₃), 2.64–2.96 (3H, m, H-8, H₂-10), 3.00–3.30 (2H, m, H-5, -9), 3.64–3.92 (5H, m, H-5', -CO₂CH₃, -C₆H₄CHCHH-O-), 4.02–4.81 (3H, m, H₂-6', -C₆H₄CHCHH-O-), 4.90–5.70 (7H, m, H-1, -7, -1', -2', -3', -4', -C₆H₄CH-O-), 7.40–8.18 (5H, m, H-3, arom. H). FAB-MS *m/z*: 738 (M+H)⁺. Anal. Calcd for C₃₃H₃₉NO₁₈: C, 53.73; H, 5.33; N, 1.90. Found: C, 53.65; H, 5.30; N, 1.91.

Epoxy-aldehydes 15a and 15b A solution of **14a** (20.1 mg) in benzene (2.5 ml) was stirred at intervals of 10 cm under irradiation through a high-pressure mercury lamp (λ_{max} 365 nm, 500 W, Eikosha Co., Ltd.) in a Pyrex filter for 1 h at room temperature. Removal of the solvent *in*

vacuo gave a residue, which was purified by PTLC (benzene:EtOAc = 7:3, 4 developments) to afford epoxy-aldehyde **15a** (9.3 mg, 57%) as a white powder. [α]_D²⁵ -67.50° (c = 0.800, CHCl₃). UV λ_{max}^{EtOH} nm (log ε): 233 (3.98). IR ν_{max}^{KBr} cm⁻¹: 1760, 1705, 1630. ¹H-NMR (CDCl₃) δ: 1.81 (1H, ddd, J = 8.0, 6.0, 4.0 Hz, H-9), 1.94, 2.01, 2.03, 2.10 (12H, each s, -OCOCH₃), 2.56 (1H, dd, J = 5.0, 3.0 Hz, H-10a), 2.73–2.81 (1H, m, H-8), 2.78 (2H, m, H-10b, ddd, J = 17.6, 8.0, 1.1 Hz, H-6a), 3.05 (1H, ddd, J = 17.6, 5.5, 1.1 Hz, H-6b), 3.39 (1H, dddd, J = 8.0, 6.0, 5.5, 1.8 Hz, H-5), 3.71 (3H, s, -CO₂CH₃), 3.74 (1H, ddd, J = 9.5, 4.4, 2.6 Hz, H-5'), 4.16 (1H, dd, J = 12.5, 2.6 Hz, H-6'a), 4.29 (1H, dd, J = 12.5, 4.4 Hz, H-6'b), 4.89 (1H, d, J = 8.1 Hz, H-1'), 5.01 (1H, dd, J = 9.2, 8.1 Hz, H-2'), 5.11 (1H, dd, J = 9.5, 9.2 Hz, H-4'), 5.23 (1H, t, J = 9.2 Hz, H-3'), 5.38 (1H, d, J = 4.0 Hz, H-1), 7.43 (1H, d, J = 1.8 Hz, H-3), 9.78 (1H, t like, J = 1.1 Hz, H-7). FAB-MS *m/z*: 573 (M+H)⁺. Anal. Calcd for C₂₃H₃₂O₁₅: C, 52.45; H, 5.63. Found: C, 52.65; H, 5.55.

14b (18.4 mg) was deprotected as described above to give **15b** (8.4 mg, 59%) as a white powder. [α]_D²⁵ -62.59° (c = 0.735, CHCl₃). UV λ_{max}^{EtOH} nm (log ε): 232 (3.94). IR ν_{max}^{KBr} cm⁻¹: 1760, 1710, 1630. ¹H-NMR (CDCl₃) δ: 1.81 (1H, ddd, J = 9.0, 6.0, 2.6 Hz, H-9), 1.91, 2.01, 2.03, 2.10 (12H, each s, -OCOCH₃), 2.52 (1H, ddd, J = 17.6, 8.0, 1.8 Hz, H-6a), 2.53 (1H, dd, J = 5.0, 2.8 Hz, H-10a), 2.73 (12H, ddd, J = 9.0, 4.0, 2.8 Hz, H-8), 2.82 (1H, dd, J = 5.0, 4.0 Hz, H-10b), 3.38 (1H, dddd, J = 8.0, 6.0, 5.5, 2.2 Hz, H-5), 3.70 (3H, s, -CO₂CH₃), 3.75 (1H, ddd, J = 9.5, 4.8, 2.6 Hz, H-5'), 4.13 (1H, dd, J = 12.5, 2.6 Hz, H-6'a), 4.30 (1H, dd, J = 12.5, 4.8 Hz, H-6'b), 4.90 (1H, d, J = 8.1 Hz, H-1'), 5.01 (1H, dd, J = 9.2, 8.1 Hz, H-2'), 5.09 (1H, dd, J = 9.5, 9.2 Hz, H-4'), 5.23 (1H, t, J = 9.2 Hz, H-3'), 5.58 (1H, d, J = 2.6 Hz, H-1), 7.48 (1H, d, J = 2.2 Hz, H-3), 9.75 (1H, t like dd, J = 1.8, 1.5 Hz, H-7). FAB-MS *m/z*: 573 (M+H)⁺. Anal. Calcd for C₂₃H₃₂O₁₅: C, 52.45; H, 5.63. Found: C, 52.63; H, 5.70.

Epoxy-lactones 16a and 16b A solution of NaBH₄ (2 mg) in EtOH (1 ml) was added dropwise to a stirred solution of **15a** (22.8 mg) in EtOH (1 ml) under ice-cooling and the mixture was stirred for 2 h at room temperature. After decomposition of the excess reagent by adding two drops of AcOH under ice-cooling, the mixture was diluted with H₂O (5 ml) and extracted with CHCl₃ (5 ml × 4). The CHCl₃ layer was washed successively with saturated aqueous NaHCO₃ and H₂O, dried and concentrated *in vacuo*. The residue (19.4 mg) was subjected to PTLC (CHCl₃:MeOH = 50:1) to yield colorless needles (14.1 mg, 65%) of epoxy-lactone **16a** on recrystallization from EtOH. mp 158–159°C. [α]_D²⁵ -150.08° (c = 0.653, CHCl₃). UV λ_{max}^{EtOH} nm (log ε): 241 (3.92). IR ν_{max}^{KBr} cm⁻¹: 1755, 1740, 1710 sh, 1620. ¹H-NMR (CDCl₃, 500 MHz) δ: 1.70 (1H, ddd, J = 9.0, 6.5, 2.0 Hz, H-9), 1.95–2.07 (2H, m, H₂-6, covered with acetyl group), 1.98, 2.01, 2.04, 2.10 (12H, each s, -OCOCH₃), 2.60 (1H, dd, J = 4.5, 2.5 Hz, H-10a), 2.74 (1H, ddd, J = 9.0, 4.0, 2.5 Hz, H-8), 2.80 (1H, dd, J = 4.5, 4.0 Hz, H-10b), 2.94 (1H, dtd, J = 12.5, 6.0, 2.5 Hz, H-5), 3.75 (1H, ddd, J = 9.5, 4.0, 2.0 Hz, H-5'), 4.16 (1H, dd, J = 12.5, 2.0 Hz, H-6'a), 4.30 (1H, dd, J = 12.5, 4.0 Hz, H-6'b), 4.34 (1H, ddd, J = 12.5, 11.5, 2.5 Hz, H-7ax), 4.54 (1H, ddd, J = 11.5, 4.5, 2.0 Hz, H-7eq), 4.90 (1H, d, J = 8.0 Hz, H-1'), 4.98 (1H, dd, J = 9.5, 8.0 Hz, H-2'), 5.09 (1H, t, J = 9.5 Hz, H-4'), 5.24 (1H, t, J = 9.5 Hz, H-3'), 5.43 (1H, d, J = 2.0 Hz, H-1), 7.57 (1H, d, J = 2.5 Hz, H-3). FAB-MS *m/z*: 543 (M+H)⁺. Anal. Calcd for C₂₄H₃₀O₁₄: C, 53.14; H, 5.57. Found: C, 53.08; H, 5.62. This compound was identical with (8S)-8,10-epoxy-sweroside tetraacetate derived from sweroside tetraacetate (**17**) (mixed mp, IR and ¹H-NMR).

15b (20.1 mg) was reduced with NaBH₄ (2 mg) as described above and PTLC followed by recrystallization of the crude product (16.2 mg) gave colorless needles (13.6 mg, 71%) of epoxy-lactone **16b**. mp 199–200°C. [α]_D²⁵ -150.99° (c = 0.404, CHCl₃). UV λ_{max}^{EtOH} nm (log ε): 242 (3.98). IR ν_{max}^{KBr} cm⁻¹: 1755, 1710, 1630. ¹H-NMR (CDCl₃, 500 MHz) δ: 1.67 (1H, ddd, J = 9.5, 5.5, 2.0 Hz, H-9), 1.83 (1H, dddd, J = 12.5, 5.5, 3.2, 2.5 Hz, H-6eq), 1.88 (1H, qd, J = ca. 12.5, 4.5 Hz, H-6ax), 1.98, 2.01, 2.03, 2.10 (12H, each s, -OCOCH₃), 2.57 (1H, dd, J = 5.2, 2.5 Hz, H-10a), 2.72 (1H, ddd, J = 9.5, 4.0, 2.5 Hz, H-8), 2.88 (1H, dd, J = 5.2, 4.0 Hz, H-10b), 2.96 (1H, dtd, J = 12.5, 5.5, 2.5 Hz, H-5), 3.77 (1H, ddd, J = 9.5, 4.5, 2.0 Hz, H-5'), 4.12 (1H, dd, J = 12.5, 2.0 Hz, H-6'a), 4.31 (1H, dd, J = 12.5, 4.5 Hz, H-6'b), 4.34 (1H, td, J = 11.5, 3.2 Hz, H-7ax), 4.50 (1H, ddd, J = 11.5, 4.2, 2.5 Hz, H-7eq), 4.93 (1H, d, J = 8.0 Hz, H-1'), 4.98 (1H, dd, J = 9.5, 8.0 Hz, H-2'), 5.07 (1H, t, J = 9.5 Hz, H-4'), 5.25 (1H, t, J = 9.5 Hz, H-3'), 5.66 (1H, d, J = 2.0 Hz, H-1), 7.62 (1H, d, J = 2.5 Hz, H-3). FAB-MS *m/z*: 543 (M+H)⁺. Anal. Calcd for C₂₄H₃₀O₁₄: C, 53.14; H, 5.57. Found: C, 53.21; H, 5.47. This compound was identical with (8R)-8,10-epoxy-sweroside tetraacetate derived from sweroside tetraacetate (**17**) (mixed mp, IR and ¹H-NMR).

Preparation of Epoxides 16a and 16b from Sweroside Tetraacetate (17) i) Diols **18a** and **18b**: In the same way as described above for the preparation of **12**, **17** (300 mg) was dihydroxylated with *N*-methylmorpholine-*N*-oxide (140 mg) in acetone and OsO₄ (20 mg) in *tert*-BuOH for 6 d. The crude product (260 mg) was subjected to PTLC (Et₂O: MeOH=95:5, 4 developments). Of the two major bands, the less polar one gave a residue (38.3 mg), which yielded on recrystallization from Et₂O–MeOH, colorless needles (36.4 mg, 11%) of diol **18a**. mp 109–110 °C (lit.,¹³) mp 111–113 °C. [α]_D¹⁵ –142.05° (*c*=0.352, CHCl₃). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 243 (3.92). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3425, 1750, 1700, 1620. ¹H-NMR (CDCl₃) δ : 1.93, 1.99, 2.01, 2.08 (12H, each s, –OCOCH₃), 2.24 (1H, td, *J*=6.0, 1.9 Hz, H-9), 2.92–3.08 (1H, m, H-5), 3.72 (1H, ddd, *J*=9.9, 4.3, 2.5 Hz, 5'-H), 3.56–3.68 (3H, m, H-8, H₂-10), 4.10 (1H, dd, *J*=12.5, 2.5 Hz, H-6'a), 4.29 (1H, dd, *J*=11.0, 2.5 Hz, H-7a), 4.33 (1H, dd, *J*=12.5, 4.3 Hz, H-6'b), 4.50 (1H, dt, *J*=11.0, 3.0 Hz, H-7b), 4.86 (1H, d, *J*=8.0 Hz, H-1'), 4.98 (1H, dd, *J*=9.3, 8.0 Hz, H-2'), 5.08 (1H, dd, *J*=9.6, 9.3 Hz, H-4'), 5.23 (1H, t, *J*=9.3 Hz, H-3'), 5.51 (1H, d, *J*=1.9 Hz, H-1), 7.52 (1H, d, *J*=2.6 Hz, H-3). FAB-MS *m/z*: 561 (M+H)⁺. Anal. Calcd for C₂₄H₃₂O₁₅: C, 51.43; H, 5.75. Found: C, 51.23; H, 5.87.

The more polar one gave a residue (50.7 mg), which yielded colorless needles (48.7 mg, 15%) of diol **18b** on recrystallization from Et₂O–MeOH. mp 92–93 °C (lit.,¹³) mp 92–93 °C. [α]_D¹⁵ –108.70° (*c*=0.092, CHCl₃). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 246 (3.81). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3450, 1760, 1740 sh, 1710, 1622. ¹H-NMR (CDCl₃) δ : 1.68–1.84 (2H, m, H₂-6), 1.94, 1.99, 2.02, 2.08 (12H, each s, –OCOCH₃), 2.14 (1H, t like, H-9), 2.84–3.04 (1H, m, H-5), 3.72 (1H, ddd, *J*=9.6, 4.5, 2.5 Hz, H-5'), 3.44–3.70 (3H, m, H-8, H₂-10), 4.15 (1H, dd, *J*=12.5, 2.5 Hz, H-6'a), 4.31 (1H, dd, *J*=12.5, 4.5, H-6'b), 4.49 (1H, dt like, H-7), 4.90 (1H, d, *J*=8.0 Hz, H-1'), 4.99 (1H, dd, *J*=9.3, 8.0 Hz, H-2'), 5.08 (1H, dd, *J*=9.6, 9.3 Hz, H-4'), 5.24 (1H, t, *J*=9.3 Hz, H-3'), 5.81 (1H, d, *J*=2.0 Hz, H-1), 7.64 (1H, d, *J*=2.5 Hz, H-3). FAB-MS *m/z*: 561 (M+H)⁺. Anal. Calcd for C₂₄H₃₂O₁₅: C, 51.43; H, 5.75. Found: C, 51.30; H, 5.67.

ii) Tosylates **19a** and **19b**: In the same way as described above for the preparation of **13**, **18a** (63 mg) in dry pyridine (1 ml) was tosylated with *p*-TsCl (21.6 mg) and molecular sieves type 4A (100 mg). After stirring for 40 h, *p*-TsCl (10.3 mg) was added to the mixture and stirred for further 15 h and worked-up as described above. PTLC (1st; Et₂O: MeOH=19:1, 2nd; CHCl₃: MeOH=50:1, 3 developments) followed by recrystallization from EtOH of the crude product (100 mg) gave colorless needles (56.2 mg, 70%) of tosylate **19a**. mp 87–88 °C. [α]_D¹⁵ –96.93° (*c*=0.619, CHCl₃). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 227 (4.28), 240 inf (3.96), 271 inf (2.78). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3475, 1750, 1710, 1620. ¹H-NMR (CDCl₃) δ : 1.80–1.95 (2H, m, H₂-6), 1.95, 2.01, 2.04, 2.10 (12H, each s, –OCOCH₃), 2.28 (1H, td, *J*=6.5, 1.5 Hz, H-9), 2.48 (3H, s, arom. CH₃), 3.00 (1H, m, H-5), 3.77 (2H, ddd, *J*=9.6, 4.5, 2.0 Hz, H-5' and m, H-8), 4.10–4.24 (3H, m, H-7a, H₂-10), 4.14 (1H, dd, *J*=12.5, 2.0 Hz, H-6'a), 4.32 (1H, dd, *J*=12.5, 4.5 Hz, H-6'b), 4.45 (1H, dt, *J*=11.0, 3.0 Hz, H-7b), 4.84 (1H, d, *J*=8.0 Hz, H-1'), 4.96 (1H, dd, *J*=9.3, 8.0 Hz, H-2'), 5.08 (1H, dd, *J*=9.6, 9.3 Hz, H-4'), 5.24 (1H, t, *J*=9.3 Hz, H-3'), 5.51 (1H, d, *J*=1.5 Hz, H-1), 7.36 (2H, AA'BB' pattern, *J*_{ortho}=8.3 Hz, arom. H), 7.47 (1H, d, *J*=2.5 Hz, H-3), 7.78 (2H, *J*_{ortho}=8.3 Hz, arom. H). FAB-MS *m/z*: 715 (M+H)⁺. Anal. Calcd for C₃₁H₃₈O₁₇S: C, 52.10; H, 5.36. Found: C, 52.08; H, 5.29.

Likewise, diol **18b** (62 mg) was tosylated with pyridine (1 ml)–*p*-TsCl (22.9 mg) as described above and the crude product (100 mg) was subjected to PTLC followed by recrystallization from EtOH to give colorless needles (52.3 mg, 66%) of tosylate **19b**. mp 92 °C. [α]_D¹⁵ –109.30° (*c*=0.677, CHCl₃). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 227 (4.26), 240 inf (3.94), 271 inf (3.00). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3500, 1750, 1710, 1620. ¹H-NMR (CDCl₃) δ : 1.95, 2.00, 2.04, 2.09 (12H, each s, –OCOCH₃), 2.47 (3H, s, arom. CH₃), 2.85–3.04 (1H, m, H-5), 3.75 (1H, ddd, *J*=10.0, 4.5, 2.5 Hz, H-5'), 3.80 (1H, dd, *J*=7.5, 3.0 Hz, H-8), 3.93–4.21 (3H, m, H-7a, H₂-10), 4.18 (1H, dd, *J*=12.6, 2.0 Hz, H-6'a), 4.30 (1H, dd, *J*=12.6, 4.5 Hz, H-6'b), 4.44 (1H, dt, *J*=11.0, 3.0 Hz, H-7b), 4.86 (1H, d, *J*=8.0 Hz, H-1'), 4.96 (1H, dd, *J*=9.3, 8.0 Hz, H-2'), 5.07 (1H, dd, *J*=10.0, 9.3 Hz, H-4'), 5.23 (1H, t, *J*=9.3 Hz, H-3'), 5.75 (1H, d, *J*=1.9 Hz, H-1), 7.36 (2H, AA'BB' pattern, *J*_{ortho}=8.3 Hz, arom. H), 7.59 (1H, d, *J*=2.6 Hz, H-3), 7.78 (2H, AA'BB' pattern, *J*_{ortho}=8.3 Hz, arom. H). FAB-MS *m/z*: 715 (M+H)⁺. Anal. Calcd for C₃₁H₃₈O₁₇S: C, 52.10; H, 5.36. Found: C, 52.21; H, 5.48.

iii) Epoxides **16a** and **16b**: Tosylate **19a** (28.7 mg) in dry benzene (1 ml) was epoxidated with DBN (0.021 ml) under stirring for 15 h as described above. The crude product (23.1 mg) was subjected to PTLC (CHCl₃:

MeOH=50:1, 4 developments) followed by recrystallization from EtOH to give (8*S*)-8,10-epoxyswerside tetraacetate (**16a**) (13.0 mg, 60%) as colorless needles.

Tosylate **19b** (30.4 mg) was treated with DBN (0.025 ml) as described above and purification of the product (32.8 mg) gave (8*R*)-8,10-epoxyswerside tetraacetate (**16b**) (13.0 mg, 56%).

Aldehydes 8a and 8b Methanolic NaOMe (1 N, 0.01 ml) was added to a solution of **15a** (31.6 mg) in dry MeOH (0.2 ml) and the mixture was stirred for 30 min at room temperature. After neutralization of the mixture by adding Amberlite IR-120 B (H⁺ form), the resin was filtered off. The filtrate was concentrated *in vacuo* to give a residue (26.2 mg), which was purified by PTLC (CHCl₃: MeOH=5:1, 5 developments) to yield a white powder (16.8 mg, 75%) of (8*S*)-8,10-epoxide **8a**. [α]_D¹⁸ –136.59° (*c*=0.410, MeOH). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 231 (4.08). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3380, 1710, 1635. ¹H-NMR (CD₃OD) δ : 1.74 (1H, ddd, *J*=8.0, 5.0, 2.0 Hz, H-9), 1.85 (1H, ddd, *J*=12.0, 6.4, 1.0 Hz, H-6a), 2.60 (1H, dd, *J*=4.5, 2.5 Hz, H-10a), 2.84 (1H, dd, *J*=4.5, 3.5 Hz, H-10b), 3.01 (1H, ddd, *J*=12.0, 4.8, 1.2 Hz, H-6b), 3.70 (3H, s, –CO₂CH₃), 4.70 (1H, d, *J*=7.0 Hz, H-1'), 5.63 (1H, d, *J*=2.0 Hz, H-1), 7.54 (1H, d, *J*=2.0 Hz, H-3), 9.72 (1H, t like, H-7). FAB-MS (glycerol) *m/z*: 403 (M–H)[–]. Anal. Calcd for C₁₇H₂₄O₁₁·1/2H₂O: C, 49.39; H, 6.10. Found: C, 49.42; H, 5.98.

Aldehyde **15b** (15.0 mg) was worked-up as described above to give a white powder (9.8 mg, 93%) of (8*R*)-8,10-epoxide **8b**. [α]_D¹⁸ –120.00° (*c*=0.175, MeOH). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 233 (4.02). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3350, 1690, 1620. ¹H-NMR (CD₃OD) δ : 1.70 (1H, ddd, *J*=7.5, 5.0, 2.0 Hz, H-9), 2.63 (1H, dd, *J*=4.5, 2.5 Hz, H-10), 3.69 (3H, s, –CO₂CH₃), 4.67 (1H, d, *J*=7.0 Hz, H-1'), 5.62 (1H, d, *J*=4.5 Hz, H-1), 7.57 (1H, d, *J*=1.2 Hz, H-3), 9.73 (1H, t like, H-7). FAB-MS *m/z*: 405 (M+H)⁺. Anal. Calcd for C₁₇H₂₄O₁₁·1/2H₂O: C, 49.39; H, 6.10. Found: C, 49.21; H, 6.00.

Acids 21a and 21b Jones reagent (0.02 ml) was added dropwise to a stirred solution of **15a** (25.4 mg) in acetone (1.0 ml) under ice-cooling and stirring was continued for 25 min. After decomposition of the excess reagent by adding a drop of MeOH, the reaction mixture was diluted with H₂O (4.0 ml) and extracted with CHCl₃ (5 ml × 4). The CHCl₃ layer was washed with brine, dried and concentrated *in vacuo*. The residue (22.1 mg) was purified by PTLC (CHCl₃: EtOAc=5:2, 2 developments) to give acid **21a** (15.2 mg, 58%) as a white powder. [α]_D¹⁸ –77.61° (*c*=0.335, CHCl₃). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 232 (3.96). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3450, 1740, 1720, 1630. ¹H-NMR (CDCl₃) δ : 1.87 (1H, ddd, *J*=8.0, 5.0, 4.0 Hz, H-9), 1.94, 2.01, 2.03, 2.11 (12H, each s, –OCOCH₃), 2.59 (1H, dd, *J*=4.2, 2.4 Hz, H-10a), 2.70 (1H, dd, *J*=10.8, 8.0, H-6a), 2.82 (1H, dd, *J*=4.2, 3.2 Hz, H-10b), 2.86 (1H, ddd, *J*=8.0, 3.2, 2.4 Hz, H-8), 3.12 (1H, dd, *J*=10.8, 4.5 Hz, H-6b), 3.28 (1H, dddd, *J*=8.0, 5.0, 4.5, 2.0 Hz, H-5), 3.72 (3H, s, –CO₂CH₃), 3.75 (1H, dd, *J*=9.0, 4.0, 2.0 Hz, H-5'), 4.16 (1H, dd, *J*=11.5, 2.0 Hz, H-6'a), 4.29 (1H, dd, *J*=11.5, 4.0 Hz, H-6'b), 4.90 (1H, d, *J*=7.2 Hz, H-1'), 5.01 (1H, dd, *J*=8.8, 7.2 Hz, H-2), 5.11 (1H, d, *J*=8.8, 8.2 Hz, H-4'), 5.23 (1H, dd, *J*=9.0, 8.2 Hz, H-3'), 5.41 (1H, d, *J*=4.0 Hz, H-1), 7.42 (1H, d, *J*=2.0 Hz, H-3). FAB-MS *m/z*: 589 (M+H)⁺. Anal. Calcd for C₂₅H₃₂O₁₆: C, 51.02; H, 5.48. Found: C, 50.97; H, 5.20.

15b (23.1 mg) was oxidized as described above to give acid **21b** (10.5 mg, 44%) as a white powder. [α]_D¹⁸ –54.01° (*c*=0.500, CHCl₃). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 229 (3.59). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3450, 1750, 1740. ¹H-NMR (CDCl₃) δ : 1.86 (1H, ddd, *J*=9.0, 6.0, 2.7 Hz, H-9), 1.91, 2.00, 2.03, 2.10 (12H, each s, –OCOCH₃), 2.45 (1H, dd, *J*=16.5, 10.0, H-6a), 2.52 (1H, dd, *J*=5.0, 2.9 Hz, H-10a), 2.75 (1H, ddd, *J*=9.1, 4.0, 2.9 Hz, H-8), 2.81 (1H, dd, *J*=5.0, 4.0 Hz, H-10b), 3.18 (1H, dd, *J*=16.5, 4.9 Hz, H-6b), 3.71 (3H, s, –CO₂CH₃), 4.12 (1H, dd, *J*=12.5, 2.9 Hz, H-6'a), 4.30 (1H, dd, *J*=12.5, 4.5 Hz, H-6'b), 4.90 (1H, d, *J*=8.1 Hz, H-1'), 5.00 (1H, dd, *J*=9.5, 8.1 Hz, H-2'), 5.09 (1H, t, *J*=10.0 Hz, H-4'), 5.22 (1H, t, *J*=10.0 Hz, H-3'), 5.57 (1H, d, *J*=2.7 Hz, H-1), 7.44 (1H, d, *J*=2.0 Hz, H-3). FAB-MS *m/z*: 589 (M+H)⁺. Anal. Calcd for C₂₅H₃₂O₁₆: C, 51.02; H, 5.48. Found: C, 51.22; H, 5.35.

Acids 10a and 10b Methanolic NaOMe (1 N, 0.03 ml) was added to a solution of **21a** (10.7 mg) in dry MeOH (0.5 ml) and the mixture was stirred for 30 min at room temperature. The reaction mixture was treated in the usual manner and the resulting crude product (9.2 mg) was purified by PTLC (CHCl₃: MeOH=85:15, 2 developments) to give a white powder (7.3 mg, 96%) of acid **10a**. [α]_D¹⁸ –89.52° (*c*=0.190, MeOH). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 235 (3.95). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3400, 1710, 1690, 1630. ¹H-NMR (CD₃OD) δ : 1.78 (1H, ddd, *J*=8.0, 5.0, 4.8 Hz, H-9), 2.58 (1H, dd, *J*=15.0, 8.0 Hz, H-6a), 2.63 (1H, dd, *J*=4.5, 2.0 Hz, H-10a),

2.79 (1H, dd, $J=4.5, 3.5$, H-10b), 2.92 (1H, ddd, $J=8.0, 3.5, 2.0$ Hz, H-8), 3.70 (3H, s, $-\text{CO}_2\text{CH}_3$), 4.65 (1H, d, $J=7.5$ Hz, H-1'), 5.65 (1H, d, $J=4.8$ Hz, H-1), 7.47 (1H, d, $J=1.2$ Hz, H-3). FAB-MS (glycerol) m/z : 421 (M+H)⁺. Anal. Calcd for $\text{C}_{17}\text{H}_{24}\text{O}_{12} \cdot 1/2\text{H}_2\text{O}$: C, 47.55; H, 5.87. Found: C, 47.82; H, 5.66.

21b (9.5 mg) was deacetylated with methanolic NaOMe (1 N, 0.02 ml) and worked-up as described above to give a white powder (5.0 mg, 74%) of acid **10b**. $[\alpha]_D^{25} -48.92^\circ$ ($c=0.450$, MeOH). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm ($\log \epsilon$): 232 (3.50). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3400, 1720, 1710. $^1\text{H-NMR}$ (CD_3OD) δ : 1.73 (1H, ddd, $J=9.0, 6.1, 4.0$ Hz, H-9), 2.54 (1H, dd, $J=16.0, 8.9$ Hz, H-6a), 2.57 (1H, dd, $J=4.6, 2.5$ Hz, H-10a), 2.79 (1H, dd, $J=4.6, 4.0$, H-10b), 3.20 (1H, dd, $J=16.0, 5.0$ Hz, H-6b), 3.69 (3H, s, $-\text{CO}_2\text{CH}_3$), 4.68 (1H, d, $J=8.0$ Hz, H-1'), 5.66 (1H, d, $J=4.0$ Hz, H-1), 7.52 (1H, d, $J=2.0$ Hz, H-3). FAB-MS (glycerol) m/z : 421 (M+H)⁺. Anal. Calcd for $\text{C}_{17}\text{H}_{24}\text{O}_{12} \cdot 1/2\text{H}_2\text{O}$: C, 47.55; H, 5.87. Found: C, 47.42; H, 5.95.

Ketone 20 Jones reagent (1 ml) was added dropwise to a solution of **13** (187.6 mg) in acetone (4 ml) under ice-cooling and the mixture was stirred for 20 h in the dark. The reaction mixture was worked-up in the usual way and the product (172.1 mg) was subjected to PTLC (CHCl_3 : MeOH = 50:1). Of the two major bands, the band around R_f 0.45 gave ketone **20** (116.4 mg, 62%) as a white powder and the band around R_f 0.03 afforded the starting material (19.4 mg). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm ($\log \epsilon$): 227 (4.37), 256 inf (3.83), 274sh (3.74). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 1755, 1740 sh, 1640, 1530. $^1\text{H-NMR}$ (CDCl_3) δ : 2.00–2.10 (12H, each s, $-\text{OCOCH}_3$), 2.44 (3H, s, arom. CH_3), 3.12–3.34 (1H, m, H-5), 3.44–3.84 (5H, m, H-5', $-\text{CO}_2\text{CH}_3$, $-\text{C}_6\text{H}_4\text{CHCH}_2\text{H}-\text{O}-$), 4.08–4.86 (3H, m, H₂-6', $-\text{C}_6\text{H}_4\text{CH}-\text{CH}_2\text{H}-\text{O}-$), 4.92–5.72 (7H, m, H-1, -7, -1', -2', -3', -4', $-\text{C}_6\text{H}_4\text{CH}-\text{O}-$), 7.30–8.16 (9H, m, H-3, arom. H). FAB-MS m/z : 909 (M+H)⁺. Anal. Calcd for $\text{C}_{46}\text{H}_{45}\text{NO}_{21}\text{S}$: C, 52.92; H, 5.00; N, 1.54. Found: C, 52.89; H, 4.86; N, 1.63.

Reduction of Ketone 20 with NaBH₄ A solution of NaBH₄ (6.3 mg) in H₂O (1 ml) was added to a stirred solution of **20** (101.4 mg) in dioxane (2.5 ml) and stirring was continued for 30 min at room temperature. The reaction mixture was worked-up in the usual way and the crude product (92.5 mg) was subjected to PTLC (CHCl_3 : MeOH = 50:1, 2 developments). Of the two major bands, the more polar one gave tosylate **13** (63.1 mg, 62%) and the less polar one the starting material (9.6 mg).

Reduction of Ketone 20 with NaB²H₄ Followed by Photolysis A solution of NaB²H₄ (20 mg) in H₂O (5 ml) was added dropwise to a solution of **20** (300 mg) in dioxane and the mixture was worked-up as described above to give tosylate [8-²H]-**13** (118.8 mg, 40%). To a solution of [8-²H]-**13** (135 mg) in dry benzene (10 ml) was added DBN (0.1 ml) and stirring was continued for 20 h at room temperature. The reaction mixture was washed successively with 1 N HCl, brine and H₂O, dried and concentrated *in vacuo*. The residue (110.2 mg) was subjected to PTLC (benzene: EtOAc = 5:1, 8 developments). Of the two major bands, the

more polar one gave [8-²H]-**14a** (47.5 mg, 43%) and the less polar one afforded [8-²H]-**14b** (27.1 mg, 25%).

A solution of [8-²H]-**14a** in benzene (6 ml) was irradiated with a high pressure mercury lamp for 1 h at room temperature at intervals of 10 cm with stirring. The reaction product was purified by PTLC (benzene: EtOAc = 7:3) to yield aldehyde [8-²H]-**15a** (22.1 mg, 60%). Only the proton signals which were changed through the introduction of deuterium were shown. $^1\text{H-NMR}$ (CDCl_3) δ : 1.81 (1H, dd, $J=6.0, 4.0$ Hz, H-9), 2.56 (1H, d, $J=5.0$ Hz, H-10a), 2.78 (1H, d, $J=5.0$ Hz, H-10b). FAB-MS m/z : 574 (M+H)⁺.

Likewise, a solution of [8-²H]-**14b** in benzene (3 ml) was irradiated as described above to afford aldehyde [8-²H]-**15b** (12.4 mg, 59%). $^1\text{H-NMR}$ (CDCl_3) δ : 1.81 (1H, dd, $J=6.0, 2.6$ Hz, H-9), 2.51 (1H, d, $J=5.0$ Hz, H-10a), 2.82 (1H, d, $J=5.0$ Hz, H-10b). FAB-MS m/z : 574 (M+H)⁺.

References and Notes

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