Dehydroxy Substitution Reactions of the Anomeric Hydroxy Groups in Some Protected Sugars Initiated by Anodic Oxidation of Triphenylphosphine

Hatsuo Maeda,* Sayaka Matsumoto, Takashi Koide, and Hidenobu Ohmori

Faculty of Pharmaceutical Sciences, Osaka University, 1-6 Yamada-oka, Suita, Osaka 565-0871, Japan. Received January 9, 1998; accepted February 16, 1998

The anodic transformation of 2,3:5,6-di-O-isopropylidene- α -D-mannofuranose (4) and 2,3,4,6-tetra-O-benzyl-D-glucopyranose (5) to the corresponding alkoxy phosphonium ions induces dehydroxy substitution of the sugars at the anomeric positions. Their dehydroxy fluorination and chlorination has been achieved by constant-current electrolysis with CH_2Cl_2-1 ,2-dimethoxyethane/ $Ph_3P/Ph_3PH \cdot BF_4$ and with $CH_2Cl_2/Ph_3P/Et_4N \cdot Cl$, respectively. The electrolysis in the presence of Ph_3P has proved to serve O-glycosylation with 4 or 5 as a glycosyl donor, provided that an aliphatic alcohol as a glycosyl acceptor is a weaker nucleophile, such as $(CF_3)_2CHOH$, CF_3CH_2OH , and tert-BuOH, than the protected sugar toward Ph_3P^+ generated anodically from Ph_3P . The present electrochemical reactions for 2,3,4,6-tetra-O-acetyl-D-glucopyranose were unsuccessful, except for the dehydroxy chlorination.

Key words anodic oxidation; dehydroxy substitution; anomeric hydroxy group; glycosyl halide; glycosylation; triphenylphosphine

We recently reported that alkoxy triphenylphosphonium ions $(Ph_3P^+-OR\cdot X^-; X=ClO_4 \text{ or } BF_4)$ are effectively prepared by the reaction of Ph_3P^+ , anodically generated from Ph_3P , with primary or secondary aliphatic alcohols. The isolated phosphonium ions have proved to function as alkylating reagents for soft nucleophiles such as Br^- , Cl^- , SCN^- , and $PhSH.^1$ In addition, it was of great interest to find that the thermal decomposition of electrochemically formed $Ph_3P^+-OR\cdot BF_4^-$ in tetrahydrofuran (THF) or dioxane affords the corresponding alkyl fluorides (R-F), where a fluorine atom from the tetrafluoroborate anion attacks the carbon α to the oxygen atom in the salt from the side opposite the oxy phosphonium moiety *via* an Sn2 mechanism. 2)

In our continued efforts to develop synthetically useful reactions initiated by anodic oxidation of phosphorus compounds, we turned our attention to the electrochemical reaction of Ph₃P in the presence of a lactol 1 in place of a simple aliphatic alcohol (Chart 1). This is because a phosphonium ion 2, anodically generated from 1, will undergo *in situ* decomposition into an oxonium ion 3, which is expected to behave as a much more reactive electrophile than 2 itself. Thus, we have examined the possibility of anodic oxidation of Ph₃P as a tool to replace anomeric hydroxy groups in protected sugars with nucleophiles not stepwise but in a single stage. In this paper, we describe the dehydroxy substitution reactions of some

* To whom correspondence should be addressed.

protected sugars at their anomeric positions, initiated by anodic formation of the corresponding alkoxy triphenylphosphonium ions.

Results and Discussion

To examine whether or not dehydroxy substitution reactions of anomeric hydroxy groups in protected sugars can be induced by anodic oxidation of Ph_3P , 2,3:5,6-di-O-isopropylidene- α -D-mannofuranose (4), 2,3,4,6-tetra-O-benzyl-D-glucopyranose (5), and 2,3,4,6-tetra-O-acetyl-D-glucopyranose (6 α , 6 β) (Chart 2) were utilized as model substrates. The electrochemical reactions of the sugars were carried out by constant-current electrolysis (CCE) at room temperature under N_2 atmosphere. An undivided cell, equipped with a graphite plate anode and a Pt foil cathode, was used for the CCE throughout. Table 1 summarizes the results of dehydroxy fluorination and chlorination of 4—6 by the electrolysis in the presence of Ph_3P .

When a mixture of Ph₃P, 4, and Ph₃PH·BF₄ in CH₂Cl₂

4:
$$X = OH$$
7: $X = F$
8: $X = CI$

8: $X = CI$

9

6 α : $A = AC$, $A = AC$ and $A = AC$ and

© 1998 Pharmaceutical Society of Japan

Table 1. Dehydroxy Halogenation of 4—6 by Constant-Current Electrolysis in the Presence of Ph₃P^a)

Run	Sub-	Electrolysis conditions	Products (%)		Recovered substrate (%)
	strate				
1	4	CH ₂ Cl ₂ /Ph ₃ PH·BF ₄ /40	7 (56)	9 (25)	4 (4)
2	4	CH ₂ Cl ₂ -DME (5:1)/Ph ₃ PH·BF ₄ /40	7 (72)	9 (12)	
3	4	CH ₂ Cl ₂ /Bu ₄ N·ClO ₄ /40	8 (68)	9 (8)	4 (9)
4	4	CH ₂ Cl ₂ /Et ₄ N·Cl/80	8 (94)		
5	5	CH_2Cl_2 -DME (5:1)/ $Ph_3PH \cdot BF_4/40$	10α	(16)	5 (36)
6	5	$CH_2Cl_2/Et_4N \cdot Cl/80$	11a	(88)	-
7	6α	CH_2Cl_2 -DME (5:1)/Ph ₃ PH·BF ₄ /40	12	(0)	6 (89) ^{b)}
8	6α	$CH_2Cl_2/Et_4N \cdot Cl/80$	13α	(25)	6 $(42)^{b}$
9	6β	CH_2Cl_2 -DME (5:1)/ $Ph_3PH \cdot BF_4/40$	12	(0)	$6 (100)^{b}$
10	6β	CH ₂ Cl ₂ /Et ₄ N·Cl/80	13α	(29)	$6 (50)^{b}$

a) The electrolysis was carried out in an undivided cell, where 3 F/mol of electricity on a substrate had been allowed to be consumed. b) The configuration at C-1 was not determined.

was subjected to CCE, a glycosyl fluoride 7 was obtained in moderate yield, along with a dimer 9 (Table 1, run 1). The result was quite encouraging, indicating that an alkoxy triphenylphosphonium ion 2, generated anodically from a lactol such as 4, smoothly enters the reaction course depicted in Chart 1 even at room temperature. This finding is a fine contrast with the previous observation that similar decomposition of the alkoxy phosphonium ions derived from simple primary and secondary aliphatic alcohols requires heating under reflux in dioxane and THF, respectively.²⁾ No formation of its β -anomer suggested that the dehydroxy fluorination proceeds not via an SN2 but an SN1 process, where a fluorine atom arising from tetrafluoroborate attacks an oxonium ion such as 3 from a sterically less hindered α -side. When the electrolysis for a mixture of Ph₃P, 4, and Ph₃PH·BF₄ was conducted in CH₂Cl₂ containing 1,2-dimethoxyethane (DME) as a co-solvent, the formation of 7 was improved, the yield being 72% (run 2). It was reported that 4 was converted into 7 in 57% yield probably via the same alkoxy triphenylphosphonium tetrafluoroborate, which was chemically formed by a modified Mitsunobu reaction with Ph₃P, diethyl azodicarboxylate, and triethyloxonium tetrafluoroborate.³⁾ However, the present electrochemical method seems more useful as an effective and simple tool for the preparation of 7 from 4 than the chemical method.

The dehydroxy chlorination of 4 initiated by anodic oxidation of Ph₃P was realized just by changing a supporting electrolyte from Ph₃PH·BF₄ to Bu₄N·ClO₄. Thus, CCE for a mixture of 4, Ph₃P, and the ammonium salt in CH₂Cl₂ afforded 8 in 68% (run 3). From the observed stereochemical outcome, it was suggested that the reaction involves an SNI mechanism similar to the case for the formation of 7. As for the source of the chlorine atom incorporated into 8 in the electrolysis, cathodic reduction of CH₂Cl₂ seems to play an important role. When CCE was carried out in CH₂Cl₂ containing Ph₃PH· BF₄, the cathodic reaction consisted of an evolution of H₂ gas as well as liberation of free Ph₃P. However, it seems likely that replacing Ph₃PH·BF₄ with Bu₄N·ClO₄ forces CH₂Cl₂ to undergo reductive cleavage of C-Cl bond, generating chloride anion. Since the presence of thus

generated chloride anion seemed to cause no problem in the anodic formation of an alkoxy phosphonium ion from Ph_3P and 4, it was expected that the deliberate addition of the nucleophilic ion would improve the yield for the electrochemical dehydroxy chlorination of 4. This was the case. When $Et_4N \cdot Cl$ was used as a supporting electrolyte instead of $Bu_4N \cdot ClO_4$, the formation of 8 took place almost quantitatively (run 4).

The dehydroxy fluorination and chlorination of 5, 6α . and 6β were attempted by CCE under the conditions used in runs 2 and 4, respectively. The results are also included in Table 1. The stereoselective transformation of 5 into a glycosyl fluoride 10α was brought about by anodic oxidation of Ph₃P as in the case of 4, although the yield was not satisfactory (run 5). The electrolysis under the same conditions proved unsuccessful for the dehydroxy fluorination of 6α or 6β , and merely resulted in recovering the protected sugar in a large amount (runs 7, 9). Substitution of the anomeric hydroxy group in 5 with chloride ion was effectively performed by CCE using Et₄N·Cl as a supporting electrolyte (run 6). Both anomers of 6 also underwent the electrochemical dehydroxy chlorination in contrast to the case of the fluorination, giving the same product 13\(\alpha\) in around 25\% yields (runs 8, 10).

Based on the results described so far, the reactivity of anomeric hydroxy groups in 4-6 seems to be a determinant of how successfully the dehydroxy substitution reactions at the anomeric positions are induced by anodic oxidation of Ph₃P. In general, etherified sugars show much higher anomeric reactivities as glycosyl acceptors as well as glycosyl donors than their esterified counterparts.4) These well-known phenomena are responsible for the changes in the effectiveness for the electrochemical transformation of 4, 5, and 6. Namely, the formation of the corresponding alkoxy triphenylphosphonium ion such as 2 depicted in Chart 1 will be favored in the following order: 4>5>6, which must be reflected in the results of the present dehydroxy halogenations. In addition, it is likely that a protic supporting electrolyte Ph₃PH·BF₄ decreases the nucleophilic reactivities of the sugars more than those in the presence of an aprotic salt Bu₄N·ClO₄ or Et₄N·Cl. This is probably because to a certain extent, the protonation by the protic salt prevents the anomeric hydroxy group from attacking Ph₃P⁺ ', to form the corresponding phosphonium ion. Thus, the present dehydroxy halogenation, especially for 5 and 6 with weaker anomeric reactivities, seemed to proceed more effectively in the electrolysis with the ammonium salts rather than $Ph_3PH \cdot BF_4$. In the electrolysis with $Et_4N \cdot Cl$ as a supporting electrolyte, a reaction sequence in which Ph₃PCl₂ is initially formed at the anode and then reacts with the sugars cannot be ruled out, although our previous work^{1,5,6)} suggests that Ph₃P⁺ reacts with the anomeric hydroxy groups in the electrolysis with Ph₃PH·BF₄ or Bu₄N·ClO₄ as a supporting electrolyte.

The possibility of the dehydroxy substitution of 4-6 initiated by anodic oxidation of Ph_3P as a tool for O-glycosylation was also examined. The study seemed intriguing for the following reasons: (i) the electrochemical method might provide a novel methodology for O-glycosylation, which is crucial for the synthesis of glycoconju-

June 1998 941

gates of biological importance⁷⁾; (ii) the electrophilic reactivity of Ph₃P⁺ against various alcohols will be elucidated, which will be helpful for further development of synthetically useful reactions with the electrochemically formed alkoxy phosphonium ions. In order to achieve a desired O-glycosylation, the electrolysis of Ph₃P must be carried out in the presence of both the protected sugar and R1OH. Two routes affording a glycoside from the two components can be envisaged as shown in Chart 3: route A via the formation of 2 (cf. Chart 1) from the protected sugar; route B including the transformation of R¹OH into the corresponding phosphonium ion (Ph₃P⁺-OR¹) (14). However, it has been found that 14 with ClO₄ or BF₄ as a counter anion is unable to work as an alkylating reagent for aliphatic alcohols, or even for their alkoxides.8) Accordingly, it is conceivable that a glycoside is afforded through the present electrochemical reaction only when Ph₃P⁺ initially reacts with the protected sugar, leading to the formation of an alkylating reagent such as 3 (cf. Chart 1), which is active enough for R¹OH to react with. Thus, analysis of the electrolysis products will compare the reactivities of the protected sugar and R¹OH toward Ph_3P^+ : the sugar $> R^1OH$ by the formation of a desired glycoside, the sugar < R1OH through no formation of a desired glycoside or the formation of 14.

To establish the scope and limitation of the present route A

Ph₃P⁺

Ph₃P⁺

Ph₃P⁺-OR¹

Ph₃P⁺-OR¹

14

R¹OH: (CF₃)₂CH₂OH (15), CF₃CH₂OH (16), t-BuOH, i-PrOH, EtOH

Chart 3

Table 2. Condensation of 4-6 with R¹OH by Constant-Current Electrolysis in the Presence of Ph₃Pa¹

electrolysis in the presence of the protected sugar as a tool for O-glycosylation, the reactivity of 4 with Ph₃P⁺ · was first compared with those of various aliphatic alcohols (R¹OH) based on the above argument. Taking electronic and steric effects into consideration, (CF₃)₂CHOH (15), CF₃CH₂OH (16), tert-BuOH, iso-PrOH, and EtOH were used as R¹OH (Chart 3). A mixture of Ph₃P, 4, and R¹OH was subjected to CCE in CH₂Cl₂ containing Ph₃PH·ClO₄ (method A) or Bu₄N·ClO₄ (method B) as a supporting electrolyte using an undivided cell. The electrolysis was carried out at room temperature under N₂ atmosphere. The results are summarized in Table 2.

When the perfluorinated alcohols and *tert*-BuOH were utilized as R¹OH, the electrochemical condensation of **4** with R¹OH was successfully achieved, giving the corresponding glycosides **17a**—**c** in fair to good yields (runs 2—4). Accordingly, the protected sugar seemed to have a higher nucleophilicity than these alcohols in the reaction with Ph₃P⁺, allowing route A in Chart 3 to prevail. The lack of formation of their β -isomers in these electrolyses

17a: $X=\alpha$ -OCH(CF₃)₂

Chart 4

Run	Substrate	R¹OH (ml)	Electrolysis method ^{b)}	Products (%)	Recovered substrate (%
1	4	15 (3)	A	17a (6) 9 (6)	4 (73)
2	4	15 (3)	В	17a (57) 9 (25)	4 (4)
3	4	16 (2)	В	17b (92)	_
4	4	tert-BuOH (1)	Α	17c (79)	
5	4	tert-BuOH (1)	В	17c (20) 8 (42)	
6	4	iso-PrOH (2)	A	17d (6) 18d (4) 14d (67)	4 (25)
7¢)	4	iso-PrOH (2)	В		4 (56)
8	4	EtOH (2)	Α	17e (2) 14e (75)	4 (53)
9 c)	4	EtOH (2)	В	17e (9)	4 (82)
10	5	16 (2)	В	19b + 20b $(82, 70:30)^{d,e}$	_
11	5	tert-BuOH (1)	A	19c + 20c $(62, 46:54)^{d,f}$	5 (17)
12	6α	16 (2)	В	22b + 23b $(57, 94:6)^{d,g}$	$6 (32)^{h}$
13	6α	tert-BuOH (1)	A	21c (7)	$6 (80)^{h}$
14	6β	16 (2)	В	22b + 23b $(87, 92:8)^{d,g}$	

a) The electrolysis was carried out in CH_2Cl_2 using an undivided cell, where 3 F/mol of electricity on a substrate had been allowed to be consumed. b) Method A: $Ph_3PH \cdot ClO_4/40 \,\text{mA}$; method B: $Bu_4N \cdot ClO_4/20 \,\text{mA}$. c) ³¹P-NMR spectra indicated that no **14** existed in crude products. d) Obtained as a mixture of both isomers. e-g) The ratio between the two isomers was determined with ¹H-, ¹³C-, and ¹⁹F-NMR spectra, respectively. h) The configuration at C-1 was not determined.

942 Vol. 46, No. 6

demonstrates that the glycosylation initiated by anodic oxidation of Ph₃P proceeds not via an S_N2 but an S_N1 mechanism, where an oxonium ion 3 is probably a crucial intermediate as in the case of the dehydroxy halogenation described above. The electrochemical glycosylation of iso-PrOH and EtOH with 4 was unsuccessful (runs 6-9). In CCE by method A, phosphonium ions 14d and 14e from iso-PrOH and EtOH, respectively, were obtained in good yields (runs 6, 8). In each of the electrolyses in the presence of iso-PrOH or EtOH, 4 was always recovered. The observations indicate that route B predominates over route A when iso-PrOH and EtOH are used as R¹OH. Further quantitative studies are needed for strict estimation of the reactivities of alcohols examined here toward Ph₃P⁺. The reactivities do, however, appear to roughly follow the order EtOH and iso-PrOH>4>15, 16, and tert-BuOH, based on the results.

It should be mentioned here that the stereoselectivity in the electrochemical glycosylation with 4 was disturbed when iso-PrOH was used as a glycosyl acceptor, leading to the formation of a mixture of α - and β -anomers (3:2) (run 6). Although the exact origin of the observed stereochemical outcome is not clear, the nucleophilicity and molecular size of iso-PrOH might be responsible for inducing an SN2 process. The anomeric configurations for all of the products 17a—e and 18d were easily established from the splitting patterns for the anomeric protons on 1 H-NMR spectra: 17 exhibits an H-1 signal as a singlet, whereas the H-1 of 18d is observed as a doublet.

Electrochemical glycosylation of **16** and *tert*-BuOH was also effectively performed when **5** was used as a glycosyl donor. In each case, glycosides **19** and **20** were obtained as a mixture in a good yield, although the stereoselectivity was not satisfactory (runs 10, 11). The structures for both **19** and **20** were determined by comparing the chemical shifts for the signals due to C-1 carbons on the ¹³C-NMR spectra, based on the argument that a β -D-glucopyranoside exhibits the corresponding signal at around 100 ppm, which is shifted to a lower magnetic field than that for its α -anomer. ⁹⁾

The ability of 6 as a glycosyl donor in the present electrochemical reaction was quite poor, as expected. When the electrolysis was carried out for 6α or 6β in the presence of 16, no formation of a desired glycoside was observed. The isolated products were 1.2-orthoacetates 22b and 23b, suggesting that an oxonium 3 anodically generated from 6 is transformed into a more stable cyclic acetoxonium ion through well-known neighboring-group participation. 4c,10) The structures of 22b and 23b were determined by the recognized characteristic of the methyl group in the orthoacetate moiety on ¹H-NMR spectra. 11,12) Utilizing tert-BuOH as a glycosyl acceptor in CCE for 6α resulted in the formation of the corresponding glycoside 21c in poor yield. The anomeric configuration was confirmed by the coupling constant between H-1 and H-2 on its ¹H-NMR spectrum.

In conclusion, it was demonstrated that the anodic oxidation of Ph₃P in the presence of lactols such as protected sugars leads to the formation of the corresponding alkoxy phosphonium ions, similar to the case of the electrolysis of primary and secondary aliphatic alcohols,

as long as steric and/or stereoelectronic effects do not attenuate the nucleophilicities of the anomeric hydroxy groups. Further, the phosphonium ions proved to decompose in situ to oxonium ions, which work as glycosyl donors for nucleophiles like BF_4^- , Cl^- , and aliphatic alcohols. Although a limitation was observed for aliphatic alcohols as a glycosyl acceptor in O-glycosylation by the present electrochemical reaction, this study has provided a criterion for the reactivities of various alcohols including protected sugars 4-6 toward Ph_3P^+ , which will further contribute to the design of the anodic oxidation of Ph_3P as a unique synthetic tool.

Experimental

Infrared (IR) spectra were taken on a JASCO VALOR-III spectrometer. ¹H-, ¹³C-, and ¹⁹F-NMR spectra were obtained in CDCl₃ at 200, 67.8, and 188.3 MHz on Varian VXR-200 and JEOL EX-270 spectrometers. Chemical sifts were expressed in parts per million (δ), where tetramethylsilane (TMS) was used as an internal standard for ¹H- and ¹³C-NMR spectra, and hexafluorobenzene for ¹⁹F-NMR spectra. Mass spectra (MS) were recorded at 70 eV with a direct inlet system on a JEOL JMS-HX100 spectrometer. For column chromatography, SiO₂ (Wakogel C-200) was used. CCE was carried out with a Hokuto Denko HA301, HA104, or HA105 potentiostat/galvanostat connected with a Hokuto Denko HF201 coulomb/amperehour meter.

Materials Protected sugars 6α and 6β were prepared by treatment of 1,2,3,4,6-penta-O-acetyl- α -D-glucose and 1,2,3,4,6-penta-O-acetyl- β -D-glucose with piperidine in THF,¹³⁾ respectively. CH₂Cl₂ for the electrolysis was distilled from P₂O₅. All other chemicals were of reagent grade and were used without further purification.

General Procedure for the Electrolysis A solution (30 ml) of a protected sugar (3 mmol), Ph_3P (4, 5 or 6 mmol), and a supporting electrolyte (4, 5 or 6 mmol) in an undivided cell equipped with a graphite plate anode (12.5 cm²) and a Pt foil cathode (4.0 cm²) was subjected to CCE (20, 40 or 80 mA) at room temperature under an N_2 atmosphere. After 3.0 F/mol (vs. a protected sugar) had been passed, the electrolyte was washed with water, and the aqueous layer was extracted with CH_2CI_2 (50 ml × 2). The combined organic layer was washed with brine, dried over MgSO₄, and then concentrated under reduced pressure, and the residue was subjected to silica gel column chromatography (n-hexane–ethyl acetate) to give the products. The products $7, ^{30}$ $8, ^{12}$ $10\alpha, ^{14}$ $11\alpha, ^{12}$ and $13\alpha, ^{15}$ were identified by comparison of their spectroscopic data with those described in the cited references. Other products gave satisfactory physical data as shown below.

2,3:5,6-Di-O-isopropylidene- α -D-mannofuranosyl 2,3:5,6-Di-O-isopropylidene- α -D-mannofuranoside (9): White powder, mp 185—186 °C.

¹H-NMR δ : 5.23 (1H, s, H-1), 4.79 (1H, dd, J=6.0, 3.8 Hz), 4.57 (1H, d, J=6.0 Hz), 4.44—4.35 (1H, m), 4.15—3.98 (2H, m), 3.91 (1H, dd, J=8.0, 3.8 Hz), 1.48 (3H, s), 1.45 (3H, s), 1.38 (3H, s), 1.34 (3H, s).

¹³C-NMR δ : 112.49 (s), 108.97 (s), 101.29 (d), 97.43 (d), 84.80 (d), 80.70 (d), 79.25 (d), 72.74 (d), 66.65 (t), 26.65 (q), 25.61 (q), 24.93 (q), 24.26 (q). Anal. Calcd for C₂₄H₃₈O₁₁: C, 57.36; H, 7.62. Found: C, 57.09; H, 7.39.

1,1,1,3,3,3-Hexafluoroisopropyl 2,3:5,6-Di-*O*-isopropylidene-α-D-mannofuranoside (17a): Pale yellow oil. 1 H-NMR δ : 5.24 (1H, s, H-1), 4.86 (1H, dd, J=5.9, 3.5 Hz), 4.78 (1H, d, J=5.9 Hz), 4.49—4.37 (2H, m), 4.15—3.93 (3H, m), 1.47 (3H, s), 1.44 (3H, s), 1.39 (3H, s), 1.34 (3H, s). 13 C-NMR δ : 121.67 (sq, J_{CF} =283.2 Hz), 121.15 (sq, J_{CF} =282.0 Hz), 113.28 (s), 109.54 (s), 107.65 (d), 84.74 (d), 82.07 (d), 79.26 (d), 72.70 (d), 71.23 (d of quintet, J_{CF} =32.9 Hz), 66.79 (t), 26.79 (q), 25.88 (q), 25.21 (q), 24.49 (q). 19 F-NMR δ : -74.79 (s, CF₃). *Anal*. Calcd for C₁₅H₂₀F₆O₆: C, 43.91; H, 4.91. Found: C, 44.13; H, 4.87.

2,2,2-Trifluoroethyl 2,3:5,6-Di-O-isopropylidene-α-D-mannofuranoside (17b): Colorless oil. 1 H-NMR δ : 5.08 (1H, s, H-1), 4.82 (1H, dd, J=6.0, 3.6 Hz), 4.69 (1H, d, J=6.0 Hz), 4.46—4.37 (1H, m) 4.16—3.81 (5H, m), 1.47 (3H, s), 1.46 (3H, s), 1.38 (3H, s), 1.33 (3H, s). 13 C-NMR δ : 123.79 (sq, $J_{\rm CF}$ =277.8), 112.97 (s), 109.34 (s), 106.65 (d), 84.87 (d), 81.13 (d), 79.37 (d), 72.99 (d), 66.81 (t), 64.02 (tq, $J_{\rm CF}$ =34.9 Hz), 26.85 (q), 25.88 (q), 25.19 (q), 24.49 (q). 19 F-NMR δ : -75.30 (t, J=8.4 Hz, CF₃). Anal. Calcd for C₁₄H₂₁F₃O₆: C, 49.12; H, 6.18. Found: C, 49.11; H, 6.03.

tert-Butyl 2,3:5,6-Di-O-isopropylidene-α-D-mannofuranoside (17c): Colorless oil. 1 H-NMR δ: 5.27 (1H, s, H-1), 4.79 (1H, dd, J= 5.8, 3.6 Hz), 4.53 (1H, d, J= 6.0 Hz), 4.42—4.33 (1H, m) 4.13—3.96 (3H, m), 1.47 (3H, s) 1.45 (3H, s), 1.38 (3H, s), 1.31 (3H, s), 1.23 (9H, s). 13 C-NMR δ: 112.27 (s), 109.18 (s), 101.80 (d), 86.32 (d), 79.80 (d), 79.75 (d), 75.02 (s), 73.28 (d), 66.99 (t), 28.70 (3C, q), 26.90 (q), 25.88 (q), 25.28 (q), 24.65 (q). Anal. Calcd for $C_{16}H_{28}O_6$: C, 60.74; H, 8.92. Found: C, 60.58; H, 8.76.

Isopropyl 2,3:5,6-Di-*O*-isopropylidene-α-D-mannofuranoside (17d): Colorless oil. ¹H-NMR δ: 5.10 (1H, s, H-1), 4.79 (1H, dd, J=6.0, 3.6 Hz), 4.56 (1H, d, J=5.9 Hz), 4.44—4.35 (1H, m) 4.15—3.80 (4H, m), 1.47 (3H, s) 1.45 (3H, s), 1.38 (3H, s), 1.32 (3H, s), 1.16 (3H, t, J=6.6 Hz), 1.14 (3H, d, J=6.5 Hz). ¹³C-NMR δ: 112.42 (s), 109.20 (s), 104.63 (d), 85.43 (d), 80.18 (d), 79.64 (d), 73.22 (d), 69.25 (d), 67.01 (t), 26.88 (q), 25.88 (q), 25.21 (q), 24.49 (q), 23.41 (q), 21.49 (q). *Anal.* Calcd for $C_{15}H_{26}O_6$: C, 59.58; H, 8.67. Found: C, 59.40; H, 8.40.

Isopropyl 2,3:5,6-Di-*O*-isopropylidene-β-D-mannofuranoside (**18d**): Colorless oil. ¹H-NMR δ: 4.79 (1H, d, J=3.5 Hz, H-1), 4.69 (1H, dd, J=6.0, 3.8 Hz), 4.57 (1H, dd, J=6.0, 3.5 Hz), 4.46 (1H, dt, J=7.8, 5.3 Hz), 4.09 (2H, d, J=5.3 Hz), 3.97 (1H, quint, J=6.2 Hz), 3.54 (1H, dd, J=7.8, 3.8 Hz), 1.55 (3H, s), 1.45 (3H, s), 1.38 (3H, s), 1.36 (3H, s), 1.27 (3H, d, J=6.2 Hz), 1.22 (3H, d, J=6.2 Hz). ¹³C-NMR δ: 113.53 (s), 109.18 (s), 101.26 (d), 79.95 (d), 79.01 (d), 76.80 (d), 73.37 (d), 72.43 (d), 66.92 (t), 27.03 (q), 25.71 (q), 25.32 (q), 25.16 (q), 23.14 (q), 21.85 (q). *Anal.* Calcd for C₁₅H₂₆O₆: C, 59.58; H, 8.67. Found: C, 59.29; H, 8.48.

Ethyl 2,3:5,6-Di-*O*-isopropylidene-α-D-mannofuranoside (17e): Colorless oil. ¹H-NMR δ: 4.99 (1H, s, H-1), 4.78 (1H, dd, J=6.0, 3.7 Hz), 4.58 (1H, d, J=5.9 Hz), 4.45—4.36 (1H, m) 4.15—4.00 (2H, m), 3.93 (1H, dd, J=7.6, 3.6 Hz), 3.76—3.61 (1H, m), 3.53—3.37 (1H, m), 1.47 (3H, s), 1.46 (3H, s) 1.38 (3H, s), 1.32 (3H, s), 1.19 (3H, t, J=7.0 Hz). ¹³C-NMR δ: 112.52 (s), 109.20 (s), 106.09 (d), 85.17 (d), 80.21 (d), 79.59 (d), 73.22 (d), 66.97 (t), 62.84 (t), 26.92 (q), 25.91 (q), 25.19 (q), 24.54 (q), 14.97 (q). *Anal.* Calcd for C₁₄H₂₄O₆: C, 58.31; H, 8.39. Found: C, 58.50; H, 8.18.

2,2,2-Trifluoroethyl 2,3,4,6-Tetra-*O*-benzyl-D-glucopyranoside Obtained as a Mixture of α- and β-Isomers (**19b** and **20b**, Respectively) (70:30): White powder. ¹³C-NMR (α-anomer) δ: 97.75 (d, C-1). ¹³C-NMR (β-anomer) δ: 103.59 (d, C-1). ¹⁹F-NMR (α-anomer) δ: -74.68 (t, J=9.2 Hz, CE_3). ¹⁹F-NMR (β-anomer) δ: -75.25 (t, J=8.5 Hz, CE_3). *Anal*. Calcd for $C_{36}H_{37}F_3O_6$: C, 69.44; H, 5.99. Found: C, 69.24; H, 5.93.

tert-Butyl 2,3,4,6-Tetra-*O*-benzyl-D-glucopyranoside Obtained as a Mixture of α- and β-Anomers (46:54): Colorless oil. ¹H-NMR (α-anomer) δ: 5.14 (1H, d, J=3.8 Hz, H-1). ¹³C-NMR (α-anomer) δ: 91.45 (d, C-1). ¹³C-NMR (β-anomer) δ: 97.81 (d, C-1). *Anal.* Calcd for $C_{38}H_{44}O_6$: C, 76.48; H, 7.43. Found: C, 76.37; H, 7.43.

3,4,6-Tri-O-acetyl-1,2-O-[1-(2,2,2-trifluoroethoxy)-ethylidene)- α -D-glucopyranoside Obtained as a Mixture of Exo and Endo Diastereo-isomers (22b and 23b, Respectively): White powder. ¹H-NMR (22b) δ :

5.75 (1H, d, J=5.4 Hz, H-1), 2.13 (3H, s), 2.11 (6H, s), 1.74 (3H, s). 1 H-NMR (23b) δ : 5.68 (1H, d, J=5.1 Hz, H-1). 13 C-NMR (22b) δ : 170.71 (s), 169.63 (s), 169.14 (s), 123.57 (sq, $J_{\rm CF}$ =277.0), 121.13 (s), 97.11 (d, C-1), 73.04 (d), 69.74 (d), 68.09 (d), 67.17 (d), 63.05 (t), 61.51 (tq, $J_{\rm CF}$ =35.3 Hz), 20.77 (3C, q), 20.29 (q). 13 C-NMR (23b) δ : 122.17 (s), 97.20 (d, C-1). 19 F-NMR (22b) δ : -75.49 (t, J=9.1 Hz, CF₃). 19 F-NMR (23b) δ : -74.98 (t, J=8.5 Hz, CF₃). Anal. Calcd for C₁₆H₂₁F₃O₁₀: C, 44.65; H, 4.92. Found: C, 44.72; H, 4.79.

tert-Butyl 2,3,4,6-Tetra-*O*-acetyl-β-D-glucopyranoside (21e): White powder. $^1\text{H-NMR}$ δ: 5.25 (1H, t, J=9.5 Hz), 5.08—4.89 (2H, m), 4.64 (1H, d, J=7.9 Hz, H-1), 4.22 (1H, dd, J=12.1, 5.7 Hz), 4.09 (1H, dd, J=12.1, 2.6 Hz), 3.73—3.64 (1H, m), 2.06 (3H, s) 2.03 (3H, s), 2.02 (3H, s), 2.00 (3H, s), 1.23 (9H, s). $^{13}\text{C-NMR}$ δ: 170.65 (s), 170.38 (s), 169.47 (s), 169.16 (s), 95.52 (d), 76.55 (s), 73.12 (d), 71.64 (d), 71.59 (d), 68.88 (d), 62.44 (t), 28.42 (3C, q), 20.72 (2C, q), 20.65 (2C, q). Anal. Calcd for $\text{C}_{18}\text{H}_{28}\text{O}_{10}$: C, 53.46; H, 6.98. Found: C, 53.23; H, 6.75.

Acknowledgments This paper was supported in part by a Grantin-Aid for Scientific Research (07772099) from the Ministry of Education, Science, Sports, and Culture, Japan.

References and Notes

- Maeda H., Koide T., Maki T., Ohmori H., Chem. Pharm. Bull., 43, 1076—1080 (1995).
- Maeda H., Koide T., Matsumoto S., Ohmori H., Chem. Pharm. Bull., 44, 1480—1483 (1996).
- 3) Kunz H., Sagar W., Helv. Chim. Acta, 68, 283-287 (1985).
- For example: a) Fraser-Reid B., Acc. Chem. Res., 29, 57—66 (1996);
 b) Schmidt R. R., Angew. Chem., Int. Ed. Engl., 25, 212—235 (1986);
 c) Paulsen H., ibid., 21, 155—173 (1982).
- Ohmori H., Maeda H., Ueda C., Masui M., Chem. Pharm. Bull., 36, 1865—1868 (1988).
- Ohmori H., Maeda H., Kikuoka M., Maki T., Masui M., Tetrahedron, 47, 767—776 (1991) and references therein.
- 7) Toshima K., Tatsuta K., Chem. Rev., 93, 1503—1531 (1993).
- 8) Unpublished results.
- 9) For example: Brewster K., Harrison J. M., Inch T. D., *Tetrahedron Lett.*, **1979**, 5051—5054; Mukaiyama T., Hashimoto Y., Shoda S., *Chem. Lett.*, **1983**, 935—938.
- 10) Ogawa T., Matsui M., Carbohydr. Res., 51, C13-C18 (1976).
- 11) Perlin A. S., Can. J. Chem., 41, 399-406 (1963).
- 12) Leroux J., Perlin A. S., Carbohydr. Res., 67, 163-178 (1978).
- 13) van Boeckel C. A. A., Beetz T., Vos J. N., de Jong A. J. M., van Aelst S. F., van den Bosch R. H., Mertens J. M. R., van der Vlugt F. A., *J. Carbohydr. Chem.*, **4**, 293—321 (1985).
- Caddick S., Gazzard L., Motherwell W. B., Wilkinson J. A., Tetrahedron, 52, 149—156 (1996).
- Jensen S. R., Kirk O., Nielsen B. J., Tetrahedron, 43, 1949—1954 (1987).