## Synthesis and Cytotoxic Activity of 1-Alkoxy- and 1-Amino-2-hydroxy-1,2-dihydroacronycine Derivatives

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Sixteen new derivatives of the natural alkaloid acronycine, bearing 1-alkoxy or 1-amino and 2-hydroxy groups, were synthesized in order to clarify the role of the C-1 substitution. Studies on the cytotoxic activity of compounds 4—19 were carried out *in vitro* on L-1210 cells. Structure–activity relationships are discussed.

Key words acronycine; cytotoxicity; alkaloid; acridone

The acridone alkaloid acronycine (1), first isolated from Acronychia baueri SCHOTT (Rutaceae) in 1948<sup>1,2)</sup> exhibits a broad spectrum of activity against numerous solid tumors including sarcoma, myeloma, carcinoma and melanoma.<sup>3-5)</sup> Nevertheless, clinical trials gave only poor results,<sup>6)</sup> most probably due to the moderate potency of acronycine and its very low water-solubility which excludes parenteral formulation of the drug. Early structure-activity studies clearly indicated that the 1,2-double bond on the pyran ring was an essential requirement to observe cytotoxic activity in this series.<sup>3,7–11)</sup> Based on those data, we formulated a hypothesis of bioactivation of acronycine by transformation of the 1,2-double bond into the corresponding epoxide in vivo12) and synthesized a series of cis- and trans-1,2-dihydroxy-1,2-dihydroacronycine diesters.<sup>13,14)</sup> As expected, these compounds exhibited promising antitumor properties, with a broadened spectrum of activity and an increased potency when compared with acronycine itself on several tumor strains in vitro and *in vivo*.<sup>13,14</sup>) In a continuation of our studies on structure– activity relationships in the acronycine series, and with the aim to clarify the role of the C-1 substitution, we report here the synthesis and cytotoxic activity of 1-alkoxy and 1-amino-2-hydroxy-1,2-dihydroacronycine derivatives.

The high reactivity of the benzylic position of *cis*-1,2-dihydroxy-1,2-dihydroacronycine (**2**) and its diacetate (**3**)<sup>13)</sup> toward nucleophilic agents led us to consider these two compounds as suitable candidates for substitution reactions at C-1.

Treatment of *cis*-diol **2** with excess methanol in the presence of hydrochloric acid led to a 1:1 mixture of *cis*-2-hydroxy-1-methoxy-1,2-dihydroacronycine (**4**) and *trans*-2-hydroxy-1-methoxy-1,2-dihydroacronycine (**5**), which could be separated by column chromatography. These two methoxy compounds gave rise to the corresponding esters at the 2-position, **6**—**9**, upon treatment with acetic anhydride or chloroacetyl chloride in the presence of pyridine. It should be noted that *cis*-2-chloroacetoxy-1-methoxy-1,2-dihydroacronycine (**8**) is particularly unstable, most probably due to steric hindrance. For instance, it spontaneously reacts with trace amounts of water to give quantitatively alcohol **4**.

In a similar way, reaction of cis-1,2-dihydroxy-1,2-dihydroacronycine (2) with benzyl alcohol, carried out in tetrahydrofuran containing a catalytic amount of boron trifluoride,

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afforded a 3:7 isomeric mixture of cis-1-benzyloxy-2-hydroxy-1,2-dihydroacronycine (10) and trans-1-benzyloxy-2hydroxy-1,2-dihydroacronycine (11). The solvent plays a crucial role in the course of this latter reaction. When dimethylformamide or acetonitrile are used instead of tetrahydrofuran, 2-oxo-1,2-dihydroacronycine (12) is the only product which can be isolated from the reaction mixture. Esterification reactions, using either acetic anhydride or chloroacetyl chloride could be successfully performed on the trans- isomer 11, leading to trans-2-acetoxy-1-benzyloxy-1,2-dihydroacronycine (13) and to trans-1-benzyloxy-2-chloroacetoxy-1,2-dihydroacronycine (14), respectively. In contrast, acylation of the *cis*-isomer 10 seemed, like in the case of 4, more difficult. Indeed, treatment of 10 with chloroacetyl chloride, even when the reaction was carried out at low temperature led to a complicated mixture from which only the unexpected trans-ester 14 could be isolated and identified (which probably results from epimerization at C-1).

Amination reactions at position-1 of the 1,2-dihydroacronycine system could be easily performed using *cis*-1,2diacetoxy-1,2-dihydroacronycine (**3**).<sup>13)</sup> Indeed, treatment of **3** with methylamine in ethanolic solution at reflux led to *trans*-2-acetoxy-1-methylamino-1,2-dihydroacronycine (**15**), accompanied by its saponification product *trans*-2-hydroxy-1-methylamino-1,2-dihydroacronycine (**16**). Treatment of the latter with excess acetic anhydride gave access to *trans*-2acetoxy-1-methylacetamido-1,2-dihydroacronycine (**17**). Finally, reaction of **3** with hydrazine in ethanol gave *trans*-1hydrazino-2-hydroxy-1,2-dihydroacronycine (**18**).

The formation of two epimeric ethers upon treatment of diol **2** by methanol or benzyl alcohol is a consequence of the SN1 type reaction previously known to occur at the benzylic position of various pyranocoumarins.<sup>15,16)</sup> In contrast, amination of diester **3** proceeds with complete inversion of stereochemistry at C-1 *via* an SN2 type reaction. It is of interest to point out that treatment of diacetate **3** with excess of methanol gave the expected 1-alkoxy derivatives **6** and **7** in poor yield. The major compound of this reaction was 2-ace-toxy-acronycine (**19**). The formation of compound **19**, in this case, could be explained by an elimination reaction *via* an E1 mechanism.

The study of the cytotoxic properties of the new acronycine derivatives was carried out *in vitro* on L-1210

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v) CH<sub>3</sub>NH<sub>2</sub>, EtOH,80°C vi) NH<sub>2</sub>NH<sub>2</sub>, EtOH, 80°C vii)MeOH, 0°C

Chart 2

leukemia cells.<sup>17,18)</sup> The results (IC<sub>50</sub>) are summarized in Table 1. In contrast with *cis*- and *trans*-1,2-dihydroxy-1,2-dihydroacronycine diesters, most 1-alkoxy and 1-amino-2-hydroxy-1,2-dihydroacronycine derivatives only exhibit marginal cytotoxic activity. This lack of significant activity confirms our previous hypothesis that cytotoxicity in this series is correlated with the presence of a good leaving group at the benzylic position, able to ensure sufficient reactivity toward nucleophilic agents.<sup>13)</sup> It should nevertheless be noted that 1-alkoxy derivatives bearing a chloroacetyl ester group at position-2 such as 9 and 14 exhibit significant cytotoxic activity, within the same range of magnitude as the corresponding *cis* and *trans* diesters.<sup>13,14)</sup>

Tab	ole	1.	Cytoto	xic /	Activity <sup>a</sup>
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Compound	IC <sub>50</sub> (µм)	Compound	IC <sub>50</sub> (µм)
4	44.6	12	43.3
5	50.1	13	>100
6	43.6	14	5.2
7	54.0	15	61.3
8	NT	16	56.6
9	14.7	17	75.8
10	40.5	18	>100
11	48.2	19	>100

a) Inhibition of L1210 cell proliferation measured by the MMT assay (mean of 2 values obtained in independent experiments). NT: non tested.

## Experimental

**General Experimental Procedures** Spectra were recorded on the following apparatus: MS, Nermag R10-10C in disorption-chemical ionization, using NH<sub>3</sub> as reagent gas. NMR, Bruker AC 200, <sup>1</sup>H-NMR (200 MHz), <sup>13</sup>C-NMR (50 MHz) and a Bruker DRX400, <sup>1</sup>H-NMR (400 MHz). Chemical shifts are given in  $\delta$  with tetramethylsilane (TMS) as an internal standard. Coupling constants (*J*) are given in Hz. The signals of <sup>1</sup>H and <sup>13</sup>C spectra were unambiguously assigned by using 2D NMR techniques: <sup>1</sup>H–<sup>1</sup>H correlation spectroscopy (COSY), <sup>13</sup>C–<sup>1</sup>H HETCORR and heteronuclear multiple bond correlation (HMBC). These 2D experiments were performed using standard Bruker microprograms. Column chromatographies were conducted using flash silica gel 60 Merck (40–63 $\mu$ m), with an overpressure of 300 mbars. All new compounds gave satisfactory combustion analyses (C, H, N, within ±0.4% of calculated values).

(±)-trans-2-Hydroxy-1-methoxy-1,2-dihydroacronycine (5) To a solution of 2 (200 mg, 0.56 mmol) in MeOH (5 ml) was added HCl (10 N, 0.1 ml) and the reaction mixture was stirred for 48 h at 0 °C. Then the mixture was neutralized with resin IR-45 and the solvent was removed under reduced pressure. The remaining residue was purified by flash chromatography on Si gel with CH<sub>2</sub>Cl<sub>2</sub>-MeOH (99:1) to give compound 5 (60 mg, 29%) and 4 (60 mg, 29%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz) δ: 8.36 (1H, dd, J=8, 1.5 Hz, H-8), 7.62 (1H, td, J=8, 1.5 Hz, H-10), 7.39 (1H, d, J=8 Hz, H-11), 7.24 (1H, t, J=8 Hz, H-9), 6.26 (1H, s, H-5), 4.97 (1H, d, J=8 Hz, H-1), 3.98 (3H, s, OMe-C6), 3.82 (1H, d, J=8 Hz, H-2), 3.81 (3H, s, NMe), 2.75 (3H, s, OMe-C1), 1.59 (3H, s, Me), 1.43 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$ : 177.42 (C-7), 162.53 (C-6), 160.08 (C-4a), 148.21 (C-12a), 144.04 (C-11a), 132.48 (C-10), 126.89 (C-8), 125.17 (C-7a), 121.66 (C-9), 115.82 (C-11), 110.77 (C-6a), 99.17 (C-12b), 94.31 (C-5), 78.18 (C-3), 76.11 (C-1), 69.93 (C-2), 56.93 (OMe-C6), 49.92 (OMe-C1), 41.83 (NMe), 26.60 (Me), 17.48 (Me). MS-DCI m/z: 370 (M+H)<sup>+</sup>.

(±)-*cis*-2-Hydroxy-1-methoxy-1,2-dihydroacronycine (4) <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$ : 8.36 (1H, dd, J=8, 1.5 Hz, H-8), 7.62 (1H, td, J=8, 1.5 Hz, H-10), 7.39 (1H, d, J=8 Hz, H-11), 7.24 (1H, t, J=8 Hz, H-9), 6.27 (1H, s, H-5), 4.80 (1H, d, J=4.6 Hz, H-1), 4.05 (1H, d, J=4.6 Hz, H-2), 3.97 (3H, s, OMe-C6), 3.62 (3H, s, NMe), 3.55 (3H, s, OMe-C1), 1.60 (3H, s, Me), 1.44 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$ : 177.97 (C-7), 162.53 (C-6), 169.40 (C-4a), 149.32 (C-12a), 144.19 (C-11a), 132.47 (C-10), 127.01 (C-8), 125.25 (C-7a), 121.66 (C-9), 115.51 (C-11), 111.44 (C-6a), 98.14 (C-12b), 94.17 (C-5), 76.57 (C-3), 74.40 (C-1), 66.65 (C-2), 56.73 (OMe-C6), 55.45 (OMe-C1), 41.31 (NMe), 25.14 (Me), 21.73 (Me). MS-DCI *m/z*: 370 (M+H)<sup>+</sup>.

(±)-trans-2-Acetoxy-1-methoxy-1,2-dihydroacronycine (7) To a solution of 5 (20 mg, 0.05 mmol) in dry pyridine (1.5 ml) was added Ac<sub>2</sub>O (1.5 ml, 15 mmol). The reaction mixture was stirred for 24 h at room temperature and then the reagents were removed under reduced pressure (using a high vacuum pump). The residue was purified by flash chromatography on Si gel with cyclohexane-EtOAc (50:50) to give compound 7 (19 mg, 85%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$ : 8.33 (1H, dd, J=8, 1.5 Hz, H-8), 7.62 (1H, td, J=8, 1.5 Hz, H-10), 7.38 (1H, d, J=8 Hz, H-11), 7.25 (1H, t, J=8 Hz, H-9), 6.27 (1H, s, H-5), 5.30 (1H, d, J=9.5 Hz, H-2), 5.08 (1H, d, J=9.5 Hz, H-1), 3.99 (3H, s, OMe-C6), 3.82 (3H, s, NMe), 2.71 (3H, s, OMe-C1), 2.19 (3H, s, CH<sub>3</sub>CO), 1.46 (6H, s, 2Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz) δ: 177.65 (C-7), 169.97 (CH<sub>3</sub>CO), 162.74 (C-6), 159.84 (C-4a), 148.37 (C-12a), 144.72 (C-11a), 132.71 (C-10), 126.89 (C-8), 125.58 (C-7a), 121.87 (C-9), 115.92 (C-11), 111.40 (C-6a), 98.90 (C-12b), 94.37 (C-5), 77.00 (C-3), 73.40 (C-1), 69.90 (C-2), 56.24 (OMe-C6), 50.49 (OMe-C1), 42.04 (NMe), 26.20 (Me), 21.02 (<u>CH</u><sub>3</sub>CO), 18.93 (Me). MS-DCI m/z: 412 (M+H)<sup>+</sup>.

(±)-*cis*-2-Acetoxy-1-methoxy-1,2-dihydroacronycine (6) Treatment of 4 under conditions essentially the same as those described for the preparation of 7 afforded compound 6. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$ : 8.34 (1H, dd, *J*=8, 1.5 Hz, H-8), 7.62 (1H, td, *J*=8, 1.5 Hz, H-10), 7.35 (1H, d, *J*=8 Hz, H-11), 7.26 (1H, t, *J*=8 Hz, H-9), 6.27 (1H, s, H-5), 5.52 (1H, d, *J*=5 Hz, H-2), 4.91 (1H, d, *J*=5 Hz, H-1), 3.99 (3H, s, OMe-C6), 3.73 (3H, s, NMe), 3.45 (3H, s, OMe-C1), 2.10 (3H, s, CH<sub>3</sub>CO), 1.53 (3H, s, Me), 1.42 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$ : 177.60 (C-7), 171.12 (CH<sub>3</sub>CO), 162.70 (C-6), 158.80 (C-4a), 149.10 (C-12a), 144.41 (C-11a), 132.58 (C-10), 127.02 (C-8), 125.57 (C-7a), 121.72 (C-9), 115.72 (C-11), 111.90 (C-6a), 99.43 (C-12b), 94.26 (C-5), 77.00 (C-3), 73.04 (C-1), 68.44 (C-2), 56.44 (OMe-C6), 56.23 (OMe-C1), 41.50 (NMe), 25.25 (Me), 23.10 (Me), 20.96 (CH<sub>3</sub>CO). MS-DCI *m/z*: 412 (M+H)<sup>+</sup>.

( $\pm$ )-trans-2-Chloroacetoxy-1-methoxy-1,2-dihydroacronycine (9) To a solution of 5 (20 mg, 0.05 mmol) in dry Et<sub>2</sub>O (5 ml) was added CICOCH<sub>2</sub>Cl (0.25 ml) and dry pyridine (0.25 ml). The reaction mixture was stirred for 3 h at 0 °C and then the reagents were removed under reduced pressure (using a high vacuum pump). The residue was purified by flash chromatography on Si gel with cyclohexane–EtOAc (50:50) to give compound **9** (19 mg, 85%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$ : 8.32 (1H, dd, J= 8, 1.5 Hz, H-8), 7.62 (1H, td, J=8, 1.5 Hz, H-10), 7.35 (1H, d, J=8 Hz, H-11), 7.23 (1H, t, J=8 Hz, H-9), 6.25 (1H, s, H-5), 5.31 (1H, d, J=8 Hz, H-2), 5.08 (1H, d, J=8 Hz, H-1), 4.17 (2H, s, CICH<sub>2</sub>CO), 3.95 (3H, s, C6-OMe), 3.79 (3H, s, NMe), 2.65 (3H, s, C1-OMe), 1.46 (3H, s, Me), 1.45 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$ : 177.60 (C-7), 165.80 (CICH<sub>2</sub>CO), 162.80 (C-6), 159.83 (C-4a), 148.36 (C-12a), 144.69 (C-11a), 132.23 (C-10), 126.58 (C-8), 125.58 (C-7a), 121.62 (C-9), 115.57 (C-11), 111.43 (C-6a), 103.48 (C-12b), 94.00 (C-5), 78.97 (C-3), 73.07 (C-1), 71.55 (C-2), 56.02 (OMe), 50.27 (OMe-C1), 41.68 (NMe), 40.43 (CICH<sub>2</sub>CO), 25.86 (Me), 18.41 (Me). MS-DCI *m*/*z*: 448, 446 (M+H)<sup>+</sup>.

(±)-*cis*-2-Chloroacetoxy-1-methoxy-1,2-dihydroacronycine (8) Treatment of 4 under conditions essentially the same as those described for the preparation of 9 afforded compound 8. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz) δ: 8.33 (1H, dd, J=8, 1.5 Hz, H-8), 7.60 (1H, td, J=8, 1.5 Hz, H-10), 7.31 (1H, d, J=8 Hz, H-11), 7.21 (1H, t, J=8 Hz, H-9), 6.22 (1H, s, H-5), 5.53 (1H, d, J=4.5 Hz, H-2), 4.88 (1H, d, J=4.5 Hz, H-1), 4.03 (2H, s, ClCH<sub>2</sub>CO), 3.96 (3H, s, C6-OMe), 3.70 (3H, s, NMe), 3.45 (3H, s, C1-OMe), 1.52 (3H, s, Me), 1.45 (3H, s, Me). MS-DCI *m/z*: 448, 446 (M+H)<sup>+</sup>.

(±)-trans-1-Benzyloxy-2-hydroxy-1,2-dihydroacronycine (11) To a solution of 2 (100 mg, 0.28 mmol) in dry tetrahydrofuran (THF) (5 ml) was added benzyl alcohol (1 ml) and BF3 ether complex (0.5 ml) and the reaction mixture was stirred for 5 d at room temperature. Then the mixture was neutralized with resin IR-45 and the solvent was removed under reduced pressure (using a high vacuum pump). The remaining residue was purified by flash chromatography on Si gel with CH2Cl2-MeOH (99:1) to give compound 11 (26 mg, 21%) and 10 (12 mg, 10%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz) δ: 8.36 (1H, dd, J=8, 1.5 Hz, H-8), 7.50 (1H, td, J=8, 1.5 Hz, H-10), 7.20 (1H, t, J=8Hz, H-9), 7.15 (1H, d, J=8Hz, H-11), 7.02 (1H, t, J=8Hz, H-4'), 7.00 (2H, t, J=8 Hz, H-3', 5'), 6.82 (2H, d, J=8 Hz, H-2', 6'), 6.27 (1H, s, H-5), 5.10 (1H, d, J=8 Hz, H-1), 4.12 (1H, d, J=12 Hz, CH<sub>a</sub>-C<sub>6</sub>H<sub>5</sub>), 3.99 (3H, s, OMe-C6), 3.98 (1H, d, J=8Hz, H-2), 3.75 (3H, s, NMe), 3.62 (1H, d, J=12 Hz,  $CH_{b}-C_{6}H_{5}$ ), 1.59 (3H, s, Me), 1.45 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz) δ: 177.59 (C-7), 162.78 (C-6), 160.16 (C-4a), 148.22 (C-12a), 144.51 (C-11a), 137.45 (C-1'), 132.52 (C-10), 128.15 (C-2', 6'), 127.65 (C-3', 5'), 127.47 (C-4'), 126.76 (C-8), 125.30 (C-7a), 121.72 (C-9), 116.02 (C-11), 111.00 (C-6a), 99.30 (C-12b), 94.34 (C-5), 78.17 (C-3), 76.60 (C-1), 71.02 (C-2), 65.23 (CH2-C6H5), 56.20 (OMe-C6), 41.91 (NMe), 26.68 (Me), 17.39 (Me). MS-DCI m/z: 446 (M+H)<sup>+</sup>.

(±)-*cis*-1-Benzyloxy-2-hydroxy-1,2-dihydroacronycine (10) <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$ : 8.36 (1H, dd, *J*=8, 1.5 Hz, H-8), 7.60 (1H, td, *J*=8, 1.5 Hz, H-10), 7.24—7.20 (7H, m, H-9, 11, 2', 3', 4', 5', 6'), 6.27 (1H, s, H-5), 5.10 (1H, d, *J*=4.6 Hz, H-1), 4.90 (1H, d, *J*=12 Hz, CH<sub>a</sub>-C<sub>6</sub>H<sub>5</sub>), 4.71 (1H, d, *J*=12 Hz, CH<sub>b</sub>-C<sub>6</sub>H<sub>5</sub>), 4.15 (1H, d, *J*=8 Hz, H-2), 3.99 (3H, s, OMe-C6), 3.69 (3H, s, NMe), 1.62 (3H, s, Me), 1.50 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$ : 177.50 (C-7), 162.70 (C-6), 159.61 (C-4), 149.72 (C-12a), 144.29 (C-11a), 136.91 (C-1'), 132.33 (C-10), 128.61 (C-2', 6'), 127.92 (C-4'), 126.99 (C-8), 126.69 (C-3', 5'), 125.67 (C-7a), 121.76 (C-9), 15.94 (C-11), 112.00 (C-6a), 98.41 (C-12b), 94.34 (C-5), 76.79 (C-3), 73.74 (C-1), 69.80 (CH<sub>2</sub>-C<sub>6</sub>H<sub>5</sub>), 67.66 (C-2), 56.25 (OMe-C6), 41.89 (NMe), 25.23 (Me), 22.03 (Me). MS-DCI *m/z*: 446 (M+H)<sup>+</sup>.

(±)-trans-2-Acetoxy-1-benzyloxy-1,2-dihydroacronycine (13) Treatment of 11 under conditions essentially the same as those described for the preparation of 7 afforded compound 13. <sup>1</sup>H-NMR (CDCl<sub>2</sub>, 200 MHz)  $\delta$ : 8.33 (1H, dd, J=8, 1.5 Hz, H-8), 7.54 (1H, td, J=8, 1.5 Hz, H-10), 7.20 (1H, t, J=8Hz, H-9), 7.16 (1H, d, J=8Hz, H-11), 7.09 (1H, t, J=8Hz, H-4'), 7.00 (2H, t, J=8 Hz, H-3', 5'), 6.77 (2H, d, J=8 Hz, H-2', 6'), 6.29 (1H, s, H-5), 5.48 (1H, d, J=8Hz, H-2), 5.21 (1H, d, J=8Hz, H-1), 4.18 (1H, d, J=9 Hz, CH<sub>a</sub>-C<sub>6</sub>H<sub>5</sub>), 3.98 (3H, s, OMe-C6), 3.75 (3H, s, NMe), 3.37 (1H, d, J=9 Hz, CH<sub>b</sub>-C<sub>6</sub>H<sub>5</sub>), 1.48 (3H, s, Me), 1.46 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz) δ: 177.62 (C-7), 169.98 (COCH<sub>3</sub>), 162.87 (C-6), 159.83 (C-4a), 148.20 (C-12a), 144.60 (C-11a), 136.92 (C-1'), 132.47 (C-10), 128.58 (C-2', 6'), 128.04 (C-3', 5'), 127.49 (C-4'), 126.82 (C-8), 125.49 (C-7a), 121.84 (C-9), 116.13 (C-11), 111.43 (C-6a), 99.26 (C-12b), 94.38 (C-5), 77.00 (C-3), 73.67 (C-1), 70.48 (C-2), 65.39 (CH2-C6H5), 56.26 (OMe-C6), 41.98 (NMe), 26.26 (Me), 21.04 (COCH<sub>3</sub>), 18.65 (Me). MS-DCI m/z: 446  $(M+H)^{+}$ .

(±)-*trans*-2-Chloroacetoxy-1-benzyloxy-1,2-dihydroacronycine (14) Treatment of 11 under conditions essentially the same as those described for the preparation of 9 afforded compound 14. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$ : 8.36 (1H, dd, J=8, 1.5 Hz, H-8), 7.58 (1H, td, J=8, 1.5 Hz, H-10), 7.24 (1H, t, J=8 Hz, H-9), 7.13 (1H, d, J=8 Hz, H-11), 7.00 (1H, t, J=8 Hz, H-4'), 7.00 (2H, t, J=8 Hz, H-3', 5'), 6.72 (2H, d, J=8 Hz, H-2', 6'), 6.35 (1H, s, H-5), 5.55 (1H, d, J=8 Hz, H-2), 5.28 (1H, d, J=8 Hz, H-1), 4.20 (1H, d, J=9 Hz,  $C\underline{H}_{a}-C_{6}H_{5}$ ), 4.20 (2H, s,  $COC\underline{H}_{2}Cl$ ), 4.02 (3H, s, OMe-C6), 3.80 (3H, s, NMe), 3.52 (1H, d, J=9 Hz,  $C\underline{H}_{b}-C_{6}H_{5}$ ), 1.52 (3H, s, Me), 1.51 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$ : 177.59 (C-7), 166.59 (COCH<sub>2</sub>Cl), 16290 (C-6), 159.62 (C-4a), 148.17 (C-12a), 144.55 (C-11a), 136.66 (C-1'), 132.55 (C-10), 128.60 (C-2', 6'), 128.09 (C-3', 5'), 127.61 (C-4'), 126.83 (C-8), 126.50 (C-7a), 121.95 (C-9), 116.14 (C-11), 111.00 (C-6a), 98.83 (C-12b), 94.37 (C-5), 76.90 (C-3), 73.54 (C-1), 72.51 (C-2), 65.38 (CH<sub>2</sub>-C<sub>6</sub>H<sub>5</sub>), 56.31 (OMe-C6), 41.96 (NMe), 40.73 (COCH<sub>2</sub>Cl), 26.26 (Me), 21.04 (COCH<sub>3</sub>), 18.53 (Me). MS-DCI m/z: 512 (M+H)<sup>+</sup>.

(±)-trans-2-Hydroxy-1-methylamino-1.2-dihydroacronycine (16) To a solution of 3 (62 mg, 0.14 mmol) in EtOH (2 ml) was added CH<sub>3</sub>NH<sub>2</sub> (0.25 ml, 40% soln. in water). The reaction mixture was stirred for 16 h at 80 °C and then the reagents were removed under reduced pressure. The remaining residue was purified by flash chromatography on Si gel with  $\rm CH_2Cl_2\text{--}MeOH$ (99:1) to give compound 16 (20 mg, 38%) and 14 (6 mg, 10%). <sup>1</sup>H-NMR  $(\text{CDCl}_{3}, 200 \text{ MHz}) \delta$ : 8.32 (1H, dd, J=8, 1.5 Hz, H-8), 7.60 (1H, td, J=8, 1.5 Hz, H-10), 7.31 (1H, d, J=8 Hz, H-11), 7.22 (1H, t, J=8 Hz, H-9), 6.23 (1H, s, H-5), 4.02 (1H, d, J=8 Hz, H-2), 3.93 (3H, s, OMe-C6), 3.69 (3H, s, NMe), 3.59 (1H, d, J=8 Hz, H-1), 1.61 (3H, s, NMe-C1), 1.54 (3H, s, Me), 1.38 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz) δ: 177.52 (C-7), 161.82 (C-6), 160.14 (C-4a), 148.99 (C-12a), 144.97 (C-11a), 132.71 (C-10), 127.11 (C-8), 125.53 (C-7a), 122.06 (C-9), 116.18 (C-11), 111.00 (C-6a), 100.92 (C-12b), 94.80 (C-5), 77.00 (C-3), 70.71 (C-2), 57.66 (C-1), 56.14 (OMe-C6), 43.29 (NMe), 28.15 (NMe-C1), 26.70 (Me), 16.98 (Me). MS-DCI m/z:  $369 (M+H)^+$ .

(±)-*trans*-2-Acetoxy-1-methylamino-1,2-dihydroacronycine (15) <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$ : 8.32 (1H, dd, J=8, 1.5 Hz, H-8), 7.61 (1H, td, J=8, 1.5 Hz, H-10), 7.23 (1H, d, J=8 Hz, H-11), 7.23 (1H, t, J=8 Hz, H-9), 6.28 (1H, s, H-5), 5.98 (1H, d, J=8 Hz, H-2), 4.11 (1H, d, J=3 Hz, N-H), 3.95 (3H, s, OMe-C6), 3.63 (1H, dd, J=8, 3 Hz, H-1), 3.61 (3H, s, NMe), 2.71 (3H, s, OMe-C1), 2.01 (3H, s, CH<sub>3</sub>CO), 1.99 (3H, s, NMe-C1), 1.54 (3H, s, Me), 1.52 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$ : 177.34 (C-7), 175.51 (CH<sub>3</sub>CO), 162.55 (C-6), 160.44 (C-4a), 148.58 (C-12a), 144.97 (C-1a), 133.08 (C-10), 127.19 (C-8), 125.53 (C-7a), 122.25 (C-9), 115.69 (C-11), 110.00 (C-6a), 98.55 (C-12b), 94.99 (C-5), 78.10 (C-3), 75.38 (C-2), 56.28 (OMe-C6), 55.71 (C-1), 42.66 (NMe), 31.34 (NMe-C1), 26.12 (Me), 22.52 (CH<sub>3</sub>CO), 16.80 (Me). MS-DCI m/z: 411 (M+H)<sup>+</sup>.

(±)-*trans*-2-Acetoxy-1-methylacetamido-1,2-dihydroacronycine (17) Treatment of 16 under conditions essentially the same as those described for the preparation of 7 afforded compound 17. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$ : 8.30 (1H, dd, *J*=8, 1.5 Hz, H-8), 7.58 (1H, td, *J*=8, 1.5 Hz, H-10), 7.20 (1H, d, *J*=8 Hz, H-11), 7.20 (1H, t, *J*=8 Hz, H-9), 6.28 (1H, s, H-5), 6.20 (1H, d, *J*=8 Hz, H-1), 4.93 (1H, d, *J*=8 Hz, H-2), 3.96 (3H, s, OMe-C6), 3.63 (3H, s, NMe), 2.10 (3H, s, CH<sub>3</sub>COO-C<sub>2</sub>), 2.03 (3H, s, NMe-C1), 1.89 (3H, s, CH<sub>3</sub>CON-C1), 1.58 (3H, s, Me), 1.41 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$ : (C-6), 160.14 (C-4a), 148.74 (C-12a), 144.99 (C-11a), 132.96 (C-10), 127.04 (C-8), 125.44 (C-7a), 122.13 (C-9), 116.10 (C-11), 112.00 (C-6a), 99.35 (C-12b), 94.55 (C-5), 77.00 (C-3), 72.78 (C-2), 56.27 (OMe-C6), 52.67 (C-1), 42.97 (NMe), 30.83 (NMe-C1), 26.06 (Me), 22.60 (CH<sub>3</sub>CON-C1), 20.99 (CH<sub>3</sub>COO-C2), 18.15 (Me). MS-DCI *m/z*: 453 (M+H)<sup>+</sup>.

(±)-*trans*-2-Acetoxy-1-hydrazino-1,2-dihydroacronycine (18) To a solution of 3 (55 mg, 0.13 mmol) in EtOH (2 ml) was added NH<sub>2</sub>NH<sub>2</sub> hydrate (0.2 ml). The reaction mixture was stirred for 3 h at 80 °C and then the reagents were removed under reduced pressure. Compound 18 was purified by crystallization with MeOH (21 mg, 40%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$ : 8.90 (1H, br, N-H), 7.71 (1H, dd, *J*=8, 1.5 Hz, H-8), 7.55 (1H, td, *J*=8, 1.5 Hz, H-10), 7.33 (1H, d, *J*=8 Hz, H-11), 6.96 (1H, t, *J*=8 Hz, H-9), 5.90 (1H, s, H-5), 5.47 (1H, br, N-H), 5.12 (1H, br, N-H),4.12 (1H, d, *J*=8.8 Hz, H-2), 3.72 (3H, s, OMe-C6), 3.48 (1H, d, *J*=8.8 Hz, H-1), 3.45 (3H, s, NMe), 1.90 (3H, s, CH<sub>3</sub>CO), 1.60 (3H, s, Me), 1.38 (3H, s, Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$ : 177.03 (C-7), 170.64 (CH<sub>3</sub>CO), 161.96 (C-6), 160.20 (C-4a), 149.24 (C-12a), 145.05 (C-11a), 133.51 (C-10), 126.24 (C-8), 125.06 (C-

7a), 122.05 (C-9), 117.79 (C-11), 110.44 (C-6a), 101.61 (C-12b), 94.90 (C-5), 79.49 (C-3), 71.49 (C-2), 59.78 (C-1), 56.31 (OMe-C6), 43.40 (NMe), 27.20 (Me), 20.76 (CH<sub>3</sub>CO), 18.24 (Me). MS-DCI *m/z*: 412 (M+H)<sup>+</sup>.

**2-Acetoxy-acronycine (19)** A solution of **3** (55 mg, 0.13 mmol) in MeOH (5 ml) was stirred for 48 h at 0 °C. The solvent was removed under reduced pressure and the remaining residue was purified by flash chromatography on Si gel with  $CH_2Cl_2$ –MeOH (99.5 : 0.5) to give compound **19** (24 mg, 50%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz) & 8.39 (1H, dd, J=8, 1.5 Hz, H-8), 7.63 (1H, td, J=8, 1.5 Hz, H-10), 7.34 (1H, d, J=8 Hz, H-11), 7.23 (1H, t, J=8 Hz, H-9), 6.50 (1H, s, H-1), 6.38 (1H, s, H-5), 3.99 (3H, s, OMe-C6), 3.85 (3H, s, NMe), 2.27 (3H, s, CH<sub>3</sub>CO), 1.56 (3H, s, 2×Me). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 50 MHz) & 177.38 (C-7), 169.25 (CH<sub>3</sub>CO), 162.79 (C-6), 157.14 (C-4a), 146.57 (C-12a), 144.57 (C-11a), 140.86 (C-2), 132.72 (C-10), 127.24 (C-8), 125.48 (C-7a), 121.96 (C-9), 115.89 (C-11), 111.07 (C-6a), 109.82 (C-1), 102.18 (C-12b), 94.14 (C-5), 77.00 (C-3), 56.34 (OMe-C6), 44.23 (NMe), 24.04 (2×Me), 21.07 (CH<sub>3</sub>CO). MS-DCI m/z: 380 (M+H)<sup>+</sup>.

**Cell Culture and Cytotoxicity** L1210 cells were cultivated in RPMI 1640 medium (Gibco) supplemented with 10% fetal calf serum, 2 mM L-glutamine, 100 units/ml penicillin, 100  $\mu$ g/ml streptomycin, and 10 mM HEPES buffer (pH=7.4). Cytotoxicity was measured by the microculture tetrazolium assay as described.<sup>17)</sup> Cells were exposed to graded concentrations of drug (nine serial dilutions in triplicate) for 48 h. Results are expressed as IC<sub>50</sub>, the concentration needed to reduce by 50% the optical density of treated cells with respect to the optical density of untreated controls.

## **References and Notes**

- Hughes G. K., Lahey F. N., Price J. R., Webb L. J., *Nature* (London), 62, 223—224 (1948).
- Macdonald P. L., Robertson A. V., Aust. J. Chem., 19, 275–281 (1966).
- 3) Svoboda G. H., *Lloydia*, 29, 206–224 (1966).
- Svoboda G. H., Poore G. A., Simpson P. J., Boder G. B., J. Pharm. Sci., 55, 758—768 (1966).
- Dorr T. R., Liddil J. D., Von Hoff D. D., Soble M., Osborne C. K., Cancer Res., 49, 340–344 (1989).
- Scarffe H. J., Beaumont A. R., Crowther D., Cancer Treatment Reports, 67, 93—94 (1983).
- Schneider J., Evans E. L., Grunberg E., Fryer R. I., J. Med. Chem., 15, 266–270 (1972).
- Shieh H. L., Pezzuto J. M., Cordell G. A., Chem.-Biol. Interact., 81, 35—55 (1992).
- Elomri A., Michel S., Tillequin F., Koch M., *Heterocycles*, 34, 799– 806 (1992).
- Mitaku S., Skaltsounis A. L., Tillequin F., Koch M., Rolland Y., Pierre A., Atassi Gh., *Pharm. Res.*, 13, 939–943 (1996).
- Elomri A., Skaltsounis A. L., Michel S., Tillequin F., Koch M., Rolland Y., Pierre A., Atassi Gh., *Chem. Pharm. Bull.*, 44, 2165–2168 (1996).
- Brum-Bousquet M., Mitaku S., Skaltsounis A. L., Tillequin F., Koch M., *Planta Med.*, **54**, 470–471 (1988).
- 13) Elomri A., Mitaku S., Michel S., Skaltsounis A. L., Tillequin F., Koch M., Pierre A., Guilbaud N., Léonce S., Kraus-Berthier L., Rolland Y., Atassi Gh., *J. Med. Chem.*, **39**, 4762–4766 (1996).
- 14) Magiatis P., Mitaku S., Skaltsounis A. L., Tillequin F., Koch M., Pierré A., Atassi Gh., J. Nat. Prod., 61, 198–201 (1998).
- Shnoder H. D., Bencze W., Halpern O., Schmid H., Chem. Ber., 92, 2338—2363 (1959).
- 16) Lemmich J., Lemmich E., Nielsen B. E., Acta Chem. Scand., 20, 2497—2507 (1966).
- 17) Pierré A., Dunn T. A., Kraus-Berthier L., Léonce S., Saint-Dizier D., Regnier G., Dhainaut A., Berlion M., Bizzari J. P., Atassi Gh., *Invest. New Drug*, **10**, 137–148 (1992).
- Léonce S., Perez V., Casabianca-Pignede M. R., Anstett M., Bisagni E., Pierré A., Atassi Gh., *Invest. New Drug*, 14, 169–180 (1996).