

Structures of New Oplopane-Type Sesquiterpenoids from the Flower Buds of *Tussilago farfara* L.¹⁾

Yasunori YAOITA, Hirokazu KAMAZAWA, and Masao KIKUCHI*

Tohoku Pharmaceutical University, 4-1 Komatsushima 4-chome, Aoba-ku, Sendai, Miyagi 981-8558, Japan.

Received January 5, 1999; accepted February 4, 1999

Five new oplopane-type sesquiterpenoids, 7 β -seneciolyoxyoplopa-3(14)Z,8(10)-dien-2-one (**1**), 7 β -angeloyloxyoplopa-3(14)Z,8(10)-dien-2-one (**2**), 7 β -(4-methylseneciolyoxy)oplopa-3(14)E,8(10)-dien-2-one (**3**), 1 α -angeloyloxy-7 β -(4-methylseneciolyoxy)oplopa-3(14)Z,8(10)-dien-2-one (**4**) and 1 α ,7 β -di(4-methylseneciolyoxy)oplopa-3(14)Z,8(10)-dien-2-one (**5**), were isolated from the flower buds of *Tussilago farfara* L. (Compositae). The structures of these compounds were elucidated on the basis of spectroscopic evidence.

Key words *Tussilago farfara*; Compositae; oplopane-type sesquiterpenoid

The flower buds of *Tussilago* (*T.*) *farfara* L. (Compositae) have been widely used for the treatment of coughs, bronchitis and asthmatic disorders in China.²⁾ In previous papers, we reported the isolation and structural elucidation of the essential oil components,³⁾ sesquiterpenoids,⁴⁾ phenolic compounds⁵⁾ and triterpenoids¹⁾ from the plant. Here, we report the isolation and structural elucidation of five new oplopane-type sesquiterpenoids, 7 β -seneciolyoxyoplopa-3(14)Z,8(10)-dien-2-one (**1**), 7 β -angeloyloxyoplopa-3(14)Z,8(10)-dien-2-one (**2**), 7 β -(4-methylseneciolyoxy)oplopa-3(14)E,8(10)-dien-2-one (**3**), 1 α -angeloyloxy-7 β -(4-methylseneciolyoxy)oplopa-3(14)Z,8(10)-dien-2-one (**4**) and 1 α ,7 β -di(4-methylseneciolyoxy)oplopa-3(14)Z,8(10)-dien-2-one (**5**). Extraction and isolation were carried out as described in the Experimental section.

Compound **1** was isolated as a colorless oil, $[\alpha]_D -97.6^\circ$. The molecular formula was determined to be C₂₀H₂₈O₃ by high-resolution (HR)-MS. The IR spectrum showed absorption bands due to an α,β -unsaturated ester (1715 cm⁻¹), an α,β -unsaturated ketone (1715 cm⁻¹) and a double bond (1650 cm⁻¹). The ¹H- (*vide* Experimental) and ¹³C-NMR (Table 1) spectra were virtually identical to those of 7 β -(4-methylseneciolyoxy)oplopa-3(14)Z,8(10)-dien-2-one (**6**), recently isolated from the flower buds of *T. farfara*, as an inhibitor of platelet aggregation caused by platelet activating factor,²⁾ except for the presence of a senecieryl group⁴⁾ instead of a 4-methylsenecieryl group in **6**. In the ¹H-detected heteronuclear multiple bond correlation (HMBC) spectrum, a cross peak was observed between the H-7 at δ_H 5.53 and the C-1' at δ_C 165.7, confirming that the senecieryl group was attached to the oxygen at C-7. The stereochemistry of $\Delta^{3(14)}$ was shown to be *Z* by the nuclear Overhauser effect correlation spectroscopy (NOESY) spectrum, giving a cross peak between H-11 and H-14. The absolute configuration of oplopa-3(14)Z,8(10)-dien-2-one, which is the main skeleton of **1** has been established by Joseph-Nathan *et al.*⁶⁾ On the basis of this evidence, the structure of **1** was determined to be 7 β -seneciolyoxyoplopa-3(14)Z,8(10)-dien-2-one.

Compound **2** was isolated as a colorless oil, $[\alpha]_D -104.5^\circ$. The molecular formula was determined to be C₂₀H₂₈O₃ by HR-MS. The IR spectrum showed absorptions due to an α,β -unsaturated ester (1714 cm⁻¹), an α,β -unsaturated ketone (1714 cm⁻¹) and a double bond (1646 cm⁻¹). The ¹H- and ¹³C-NMR spectra of **2** were identical to those of **1** except for

the appearance of signals ascribable to an angeloyl group⁴⁾ instead of signals due to a senecieryl group in **1**. Thus, **2** was characterized as 7 β -angeloyloxyoplopa-3(14)Z,8(10)-dien-2-one.

Compound **3** was isolated as a colorless oil, $[\alpha]_D -33.3^\circ$. The molecular formula was determined to be C₂₁H₃₀O₃ by HR-MS. The IR spectrum showed absorptions due to an α,β -unsaturated ester (1720 cm⁻¹), an α,β -unsaturated ketone (1720 cm⁻¹) and a double bond (1646 cm⁻¹). The ¹H-NMR spectrum was similar to that of **6**,²⁾ although the ¹H-NMR spectrum showed that the olefinic methyl group (H₃-15) of **6** at δ_H 2.15 was shifted upfield to 1.90, and the trisubstituted olefinic proton (H-14) of **6** at δ_H 6.23 was shifted downfield to 6.53. The above results indicate that **3** is a $\Delta^{3(14)}$ *E* isomer of **6**.⁷⁾ Based on this evidence, the structure of **3** was deter-

Table 1. ¹³C-NMR Chemical Shifts of Compounds **1**–**5** (150 MHz, CDCl₃)

Carbon	1	2	3 ^{a)}	4	5
1	41.0	41.1	39.6	72.4	71.8
2	206.4	206.3	205.3	200.0	200.0
3	141.1	140.9	142.2	139.0	139.1
4	50.8	50.8	51.3	44.9	44.8
5	39.9	40.4	37.7	40.4	40.6
6	30.7	30.8	29.9	29.8	29.9
7	73.1	73.9	72.6	73.4	73.5
8	146.1	145.8	146.2	140.9	140.9
9	41.7	42.0	41.3	45.8	45.7
10	110.0	110.4	110.2	112.7	113.5
11	27.0	27.0	29.5	27.5	27.5
12	21.3	21.3	21.3	21.3	21.3
13	15.3	15.3	16.0	15.4	15.5
14	132.4	132.8	128.5	137.0	137.0
15	14.6	14.6	14.6	15.1	15.1
1'	165.7	167.0	166.1	165.9	166.0
2'	116.3	128.2	114.6	114.5	114.6
3'	156.8	137.8	162.1	162.1	162.2
4'	27.4	15.9	33.8	33.8	33.8
5'	20.3	20.7	12.0	11.9	11.9
6'			18.9	18.9	18.9
1''				166.5	166.0
2''				127.3	114.6
3''				139.1	164.1
4''				15.9	33.9
5''				20.6	11.9
6''					19.1

a) Measured at 100 MHz.

* To whom correspondence should be addressed.

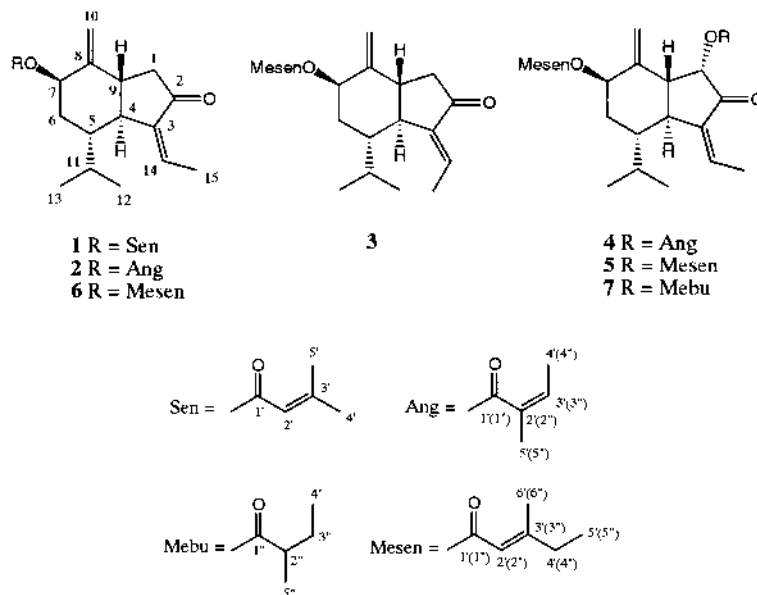


Chart 1

mined to be 7 β -(4-methylseneciyoxy)oplopa-3(14)*E*,8(10)-dien-2-one. This is the first report of the isolation of an oplopane derivative with a $\Delta^{3(14)}E$ configuration from the flower buds of *T. farfara*.

Compound **4** was isolated as a colorless oil, $[\alpha]_D -166.7^\circ$. The molecular formula was determined to be $C_{26}H_{36}O_5$ by HR-MS. The IR spectrum showed absorptions due to an α,β -unsaturated ester (1730 cm^{-1}), an α,β -unsaturated ketone (1708 cm^{-1}) and a double bond (1646 cm^{-1}). The ^1H - and ^{13}C -NMR spectra of **4** were similar to those of 1 α -(2-methylbutyryloxy)-7 β -(4-methylseneciyoxy)oplopa-3(14)*Z*,8(10)-dien-2-one (**7**),⁴⁾ except for the presence of an angeloyl group⁴⁾ in place of a 2-methylbutyryl group in **7**. The oxygenated methine proton at δ_{H} 5.58 was assigned to H-1 by a ^1H - ^1H shift correlation spectroscopy (^1H - ^1H COSY) correlation between δ_{H} 5.58 and δ_{H} 2.70 (H-9). The oxygenated quaternary carbon at δ_{C} 166.5 was assigned to C-1'' by the HMBC correlation between δ_{C} 166.5 and δ_{H} 1.86 (H₃-5''). Furthermore, in the HMBC spectrum, a cross peak was observed between the H-1 and the C-1'', confirming that the angeloyl group was attached to the oxygen at C-1. The configuration of the angeloyloxy group was shown to be α by the NOESY spectrum, giving a cross peak between H-1 β and H-9 β . On the basis of this evidence, the structure of **4** was determined to be 1 α -angeloyloxy-7 β -(4-methylseneciyoxy)oplopa-3(14)*Z*,8(10)-dien-2-one.

Compound **5** was isolated as a colorless oil, $[\alpha]_D -173.9^\circ$. The molecular formula was determined to be $C_{27}H_{38}O_5$ by HR-MS. The IR spectrum showed absorptions due to an α,β -unsaturated ester (1730 cm^{-1}), an α,β -unsaturated ketone (1710 cm^{-1}) and a double bond (1646 cm^{-1}). The ^1H - and ^{13}C -NMR spectra of **5** were similar to those of **4** except for the appearance of signals due to a 4-methylseneciyoxy group⁴⁾ instead of signals due to an angeloyl group in **4**. Therefore, **5** was characterized as 1 α ,7 β -di(4-methylseneciyoxy)oplopa-3(14)*Z*,8(10)-dien-2-one. Compounds **4** and **5** are the first examples of the isolation of oplopane derivatives with an angeloyloxy and 4-methylseneciyoxy groups at the C-1 positions, respectively, from the flower buds of *T. farfara*.

Experimental

General Procedures Optical rotations were determined with a JASCO DIP-360 digital polarimeter. IR spectra were recorded with a Perkin-Elmer FT-IR 1725X infrared spectrophotometer and UV spectra with a Beckman DU-64 spectrophotometer. ^1H - and ^{13}C -NMR spectra were recorded with JEOL JNM-LA 600 (600 and 150 MHz, respectively) and JEOL JMN-LA 400 (400 and 100 MHz, respectively) spectrometers. Chemical shifts are given on a δ (ppm) scale with tetramethylsilane as an internal standard (s, singlet; br s, broad singlet; d, doublet; dd, double doublet; ddd, double double doublet; t, triplet; q, quartet; dq, double quartet; qd, quartet doublet; qq, quartet quartet; m, multiplet). The electron ionization (EI)- and HR-MS were recorded on a JEOL JMS-DX 303 mass spectrometer. Column chromatography was carried out on Kieselgel 60 (Merck; 230–400 mesh). Preparative HPLC was carried out on a Tosoh HPLC system (pump, CCPD; detector, RI-8010) using a TSK gel ODS-120T (7.8 mm i.d. \times 30 cm) column (Tosoh).

Plant Material The dried flower buds of *T. farfara* were purchased from Tochimoto Tenkaido Co., Ltd., Osaka, Japan in 1990.

Extraction and Isolation The dried flower buds of *T. farfara* (5.0 kg) were extracted with Et_2O at room temperature for one week. The Et_2O extract was subjected to steam distillation to give an essential oil (7.7 g) and residue (64.5 g). A part of this residue (23.5 g) was placed on a silica gel column and developed with *n*-hexane–AcOEt (4 : 1–1 : 4), AcOEt and MeOH to afford 19 fractions (frs. 1–19). Fraction 5 was purified by preparative HPLC [MeOH– H_2O (4 : 1); flow rate, 1.5 ml/min; column temperature, 40 $^\circ\text{C}$] to give a mixture of **1** and **2**, **3** (0.3 mg), **4** (0.4 mg) and **5** (0.2 mg). The mixture of **1** and **2** was purified by preparative HPLC [MeOH– H_2O (5 : 2); flow rate, 1.6 ml/min; column temperature, 40 $^\circ\text{C}$] to give **1** (0.4 mg) and **2** (0.7 mg).

7 β -Seneciyoxyoplopa-3(14)*Z*,8(10)-dien-2-one (1**)** Colorless oil, $[\alpha]_D^{27} -97.6^\circ$ ($c=0.04$, CHCl_3). IR $\nu_{\text{max}}^{\text{CHCl}_3}\text{ cm}^{-1}$: 1715, 1650. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 223 (4.3). HR-MS m/z : 316.2032 (M^+ , Calcd for $\text{C}_{20}\text{H}_{28}\text{O}_5$; 316.2039). EI-MS m/z : 316 (M^+), 216 ($\text{M}^+ - \text{C}_5\text{H}_8\text{O}_2$), 173 ($\text{M}^+ - \text{C}_5\text{H}_8\text{O}_2 - \text{C}_3\text{H}_7$). ^1H -NMR (600 MHz, CDCl_3) δ : 0.81 (3H, d, $J=7.0$ Hz, H₃-13), 0.96 (3H, d, $J=7.0$ Hz, H₃-12), 1.89 (3H, d, $J=1.5$ Hz, H₃-4'), 2.12 (3H, d, $J=7.3$ Hz, H₃-15), 2.16 (3H, d, $J=1.1$ Hz, H₃-5'), 2.23 (1H, dd, $J=16.8$, 12.8 Hz, H-1 α), 2.34 (1H, m, H-11), 2.39 (1H, dd, $J=16.8$, 6.2 Hz, H-1 β), 2.59 (1H, m, H-9), 4.79 (1H, dd, $J=2.2$, 1.1 Hz, H-10a), 5.13 (1H, s, H-10b), 5.53 (1H, dd, $J=3.3$, 3.3 Hz, H-7), 5.67 (1H, dd, $J=1.5$, 1.1 Hz, H-2'), 6.22 (1H, q, $J=7.3$ Hz, H-14). ^{13}C -NMR (150 MHz, CDCl_3) see Table 1.

7 β -Angeloyloxyoplopa-3(14)*Z*,8(10)-dien-2-one (2**)** Colorless oil, $[\alpha]_D^{25} -104.5^\circ$ ($c=0.07$, CHCl_3). IR $\nu_{\text{max}}^{\text{CHCl}_3}\text{ cm}^{-1}$: 1714, 1646. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 224 (4.1). HR-MS m/z : 316.2027 (M^+ , Calcd for $\text{C}_{20}\text{H}_{28}\text{O}_5$; 316.2039). EI-MS m/z : 316 (M^+), 216 ($\text{M}^+ - \text{C}_5\text{H}_8\text{O}_2$), 173 ($\text{M}^+ - \text{C}_5\text{H}_8\text{O}_2 - \text{C}_3\text{H}_7$). ^1H -NMR (600 MHz, CDCl_3) δ : 0.82 (3H, d, $J=7.0$ Hz, H₃-13), 0.96 (3H, d, $J=7.0$ Hz, H₃-12), 1.88 (3H, dq, $J=1.5$, 1.5 Hz, H₃-5'), 1.98 (3H, dq, $J=7.3$, 1.5 Hz, H₃-4'), 2.13 (3H, dd, $J=7.0$, 2.2 Hz, H₃-15), 2.25 (1H, dd,

$J=17.2$, 13.2 Hz, H-1 α), 2.36 (1H, m, H-11), 2.40 (1H, dd, $J=17.2$, 6.2 Hz, H-1 β), 2.57 (1H, m, H-9), 4.82 (1H, dd, $J=2.2$, 1.1 Hz, H-10a), 5.16 (1H, s, H-10b), 5.61 (1H, dd, $J=3.3$, 2.9 Hz, H-7), 6.06 (1H, qq, $J=7.3$, 1.5 Hz, H-3'), 6.23 (1H, qd, $J=7.3$, 2.2 Hz, H-14). $^{13}\text{C-NMR}$ (150 MHz, CDCl_3): see Table 1.

7 β -(4-Methylseneciolyloxy)oplopa-3(14)E,8(10)-dien-2-one (3) Colorless oil, $[\alpha]_{\text{D}}^{25} -33.3^\circ$ ($c=0.03$, CHCl_3). IR $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$: 1720 , 1646 . UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 225 (4.0). HR-MS m/z : 330.2180 (M^+ , Calcd for $\text{C}_{21}\text{H}_{30}\text{O}_3$; 330.2195). EI-MS m/z : 330 (M^+), 216 ($\text{M}^+ - \text{C}_6\text{H}_{10}\text{O}_2$), 173 ($\text{M}^+ - \text{C}_6\text{H}_{10}\text{O}_2 - \text{C}_3\text{H}_7$). $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 0.86 (3H, d, $J=6.8$ Hz, H_3-13), 1.00 (3H, d, $J=7.1$ Hz, H_3-12), 1.07 (3H, t, $J=7.3$ Hz, H_3-5'), 1.90 (3H, dd, $J=7.8$, 2.0 Hz, H_3-15), 2.05 (1H, ddd, $J=14.4$, 4.4 , 4.4 Hz, H-6 β), 2.16 (3H, d, $J=1.2$ Hz, H_3-6'), 2.42 (1H, dd, $J=17.3$, 6.1 Hz, H-1 β), 2.63 (1H, m, H-9), 4.84 (1H, d, $J=1.5$ Hz, H-10a), 5.16 (1H, dd, $J=1.0$, 1.0 Hz, H-10b), 5.57 (1H, dd, $J=4.4$, 3.7 Hz, H-7), 5.67 (1H, q, $J=1.2$ Hz, H-2'), 6.53 (1H, qd, $J=7.8$, 2.0 Hz, H-14). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3): see Table 1.

1 α -Angeloyloxy-7 β -(4-methylseneciolyloxy)oplopa-3(14)Z,8(10)-dien-2-one (4) Colorless oil, $[\alpha]_{\text{D}}^{28} -166.7^\circ$ ($c=0.04$, CHCl_3). IR $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$: 1730 , 1708 , 1644 . UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 224 (4.5). HR-MS m/z : 428.2555 (M^+ , Calcd for $\text{C}_{26}\text{H}_{36}\text{O}_5$; 428.2563). EI-MS m/z : 428 (M^+), 328 ($\text{M}^+ - \text{C}_5\text{H}_8\text{O}_2$), 314 ($\text{M}^+ - \text{C}_6\text{H}_{10}\text{O}_2$), 214 ($\text{M}^+ - \text{C}_6\text{H}_{10}\text{O}_2 - \text{C}_5\text{H}_8\text{O}_2$), 173 ($\text{M}^+ - \text{C}_6\text{H}_{10}\text{O}_2 - \text{C}_5\text{H}_8\text{O}_2 - \text{C}_3\text{H}_7$). $^1\text{H-NMR}$ (600 MHz, CDCl_3) δ : 0.87 (3H, d, $J=7.0$ Hz, H_3-13), 0.98 (3H, d, $J=7.0$ Hz, H_3-12), 1.07 (3H, t, $J=7.3$ Hz, H_3-5'), 1.86 (3H, dq, $J=1.5$, 1.5 Hz, H_3-5''), 1.96 (3H, dq, $J=7.3$, 1.5 Hz, H_3-4''), 2.15 (3H, d, $J=1.1$ Hz, H_3-6'), 2.16 (3H, d, $J=7.3$ Hz, H_3-15), 2.32 (1H, m, H-11), 2.70 (1H, br s, H-9), 4.83 (1H, s, H-10a), 5.18 (1H, s, H-10b), 5.52 (1H, dd, $J=3.7$ Hz, H-7), 5.58 (1H, d, $J=3.3$ Hz, H-1), 5.64 (1H, q, $J=1.1$ Hz, H-2'), 6.10 (1H, qq, $J=7.3$, 1.5 Hz, H-3''), 6.41 (1H, q, $J=7.3$ Hz, H-

14). $^{13}\text{C-NMR}$ (150 MHz, CDCl_3): see Table 1.

1 α ,7 β -Di(4-methylseneciolyloxy)oplopa-3(14)Z,8(10)-dien-2-one (5) Colorless oil, $[\alpha]_{\text{D}}^{29} -173.9^\circ$ ($c=0.02$, CHCl_3). IR $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$: 1731 , 1710 , 1646 . UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 222 (4.6). HR-MS m/z : 442.2694 (M^+ , Calcd for $\text{C}_{27}\text{H}_{38}\text{O}_5$; 442.2719). EI-MS m/z : 442 (M^+), 328 ($\text{M}^+ - \text{C}_6\text{H}_{10}\text{O}_2$), 214 ($\text{M}^+ - \text{C}_6\text{H}_{10}\text{O}_2 \times 2$), 173 ($\text{M}^+ - \text{C}_6\text{H}_{10}\text{O}_2 \times 2 - \text{C}_3\text{H}_7$). $^1\text{H-NMR}$ (600 MHz, CDCl_3) δ : 0.89 (3H, d, $J=7.0$ Hz, H_3-13), 0.97 (3H, d, $J=7.0$ Hz, H_3-12), 1.06 (3H, t, $J=7.3$ Hz, H_3-5''), 1.07 (3H, t, $J=7.3$ Hz, H_3-5'), 2.15 (3H, d, $J=1.1$ Hz, H_3-6'), 2.16 (3H, d, $J=7.3$ Hz, H_3-15), 2.17 (3H, d, $J=1.1$ Hz, H_3-6''), 4.84 (1H, dd, $J=1.8$, 1.1 Hz, H-10a), 5.17 (1H, s, H-10b), 5.53 (2H, m, H-1, H-7), 5.64 (2H, q, $J=1.1$ Hz, H-2'), 6.40 (1H, q, $J=7.3$ Hz, H-14). $^{13}\text{C-NMR}$ (150 MHz, CDCl_3): see Table 1.

Acknowledgments We are grateful to Mrs. S. Sato and T. Matsuki of this university for measurement of the mass and NMR spectra.

References and Notes

- 1) Part V in a series of studies on the constituents of the flower buds of *Tussilago farfara* L. Part IV: Yaoita Y., Kikuchi M., *Natural Medicines*, **52**, 273—275 (1998).
- 2) Shi W., Han G., *J. Chin. Pharm. Sci.*, **5**, 63—67 (1996).
- 3) Suzuki N., Kikuchi M., *Yakugaku Zasshi*, **112**, 571—576 (1992).
- 4) Kikuchi M., Suzuki N., *Chem. Pharm. Bull.*, **40**, 2753—2755 (1992).
- 5) Kikuchi M., Mori M., *Tohoku Yakka Daigaku Kenkyu Nempo*, **39**, 69—73 (1992).
- 6) Joseph-Nathan P., Villagomez J. R., Roman L. U., Hernandez J. D., *Phytochemistry*, **28**, 1207—1209 (1989).
- 7) Aal A. M., Bohlmann F., Sarg T., El-Domiati M., Nordenstam B., *Phytochemistry*, **27**, 2599—2602 (1988).