

Triterpenoid Saponins from Berries of *Hedera colchica*

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Fifteen triterpenoid saponins were isolated from the berries of *Hedera colchica* and their structures established on the basis of chemical and spectroscopic evidence. Among them, two are new compounds: colchiside A (3) and colchiside B (15) and four are described for the first time in the berries of *Hedera colchica* (compounds 1, 8, 9 and 11).

Key words *Hedera colchica*; Araliaceae; triterpenoid saponin; NMR; MS; colchiside

Hedera colchica K. KOCH. (Araliaceae), a plant mainly growing in West Georgia, was used in traditional medicine as a bronchospasmolytic, secretolytic and antiinflammatory remedy.^{1,2} The occurrence of triterpenoid saponins in the leaves of this plant has been previously reported.^{3a,b,4} In this paper, we describe the isolation and structure elucidation of fifteen saponins from the berries of *Hedera colchica* (Fig. 1). Saponins 3 and 15, respectively named colchisides A and B were new compounds, while saponins 1, 8, 9 and 11 were isolated for the first time from the berries of this plant. The *n*-butanolic extract of the berries containing crude saponins was subjected to repeated chromatography affording saponins 1–15 (see experimental part).

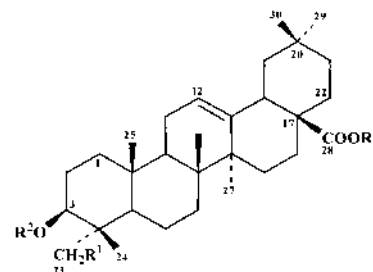
Compounds 8 and 11, which were obtained for the first time in the genus *Hedera*, were identified as staunoside A⁵) and scheffleraside II⁶) respectively, while triterpenoid saponins 1 and 9, identified as saponin I⁷) and saponin I,⁸) were isolated for the first time from the berries of *Hedera colchica*. The other saponins 2, 4, 5, 6, 7, 10, 12, 13 and 14 were identified as saponin B,⁸) saponin 2,⁷) α -hederin,⁹) saponin 4,⁷) heteroside E2,¹⁰) heteroside I,¹¹) hederasaponin D,¹²) hederasaponin B^{12,13}) and hederasaponin C,^{13,14}) respectively.

Colchiside A (3) was assigned the molecular formula C₃₅H₅₆O₈ (FAB-MS (*m/z*): 603 [M–H][–]). Acid hydrolysis of 3 yielded xylose as the sugar and hederagenin as the aglycone. The ¹³C-NMR spectrum exhibited one anomeric carbon at δ 106. The resonances of C-3 at δ 83 and C-28 at δ 182 reflected the fact that the sugar chain was linked to C-3. Moreover, the ¹³C chemical shifts of the xylose moiety were indicative of a β -D-xylopyranosyl.^{15,16} Thus 3 was a monodesmoside and its structure was elucidated as 3-*O*-(β -D-xylopyranosyl)-hederagenin.

The FAB-MS of colchiside B (15) gave the molecular ion at *m/z* 1247 [M–H][–] in agreement with a molecular formula of C₆₀H₉₆O₂₇. TLC analysis of the acid hydrolysis of 15 yielded the same sugars (glucose, rhamnose, glucuronic acid) and aglycone (oleanolic acid) as 11. The ¹³C-NMR spectrum of 15 exhibited five anomeric carbons (Table 1) located at δ 95.76, 102.72, 102.94, 104.27 and 106.83 ppm. Moreover, the ¹³C-NMR signal due to the C-28 genin moiety indicated esterification of the carboxyl group with a sugar, while the deshielded position of C-3 reflected a substitution by a sugar

chain. Saponin 15 was consequently a bidesmoside. On alkaline hydrolysis 15 gave a prosapogenin. Acid hydrolysis of this prosapogenin yielded glucuronic acid and rhamnose as the sugars. Further analysis of the ¹³C-NMR data showed that the sugar chain linked at C-28 was identical for saponins 11 and 15. Moreover, the downfield shift (83.6 ppm) of C-3 in the glucuronic acid moiety was indicative of a substitution at C-3 by a rhamnose unit. Indeed, these ¹³C-NMR results were in perfect agreement with the literature data for the sugar chain linked at C-28¹³) and for the sugar arrangement at C-3¹⁴) in which C-3 of glucuronic acid was found at δ 83.

Moreover, the values for the carboxylic group of the glucuronic part were found for compounds 11 and 15 at δ 176. In previously reported data,¹⁴) the carboxylic signals res-



	R ¹	R ²	R ³
1	H	Ara	H
2	OH	Ara	H
3	OH	Xyl	H
4	OH	Glc	H
5	OH	Ara ² — ¹ Rha	H
6	OH	Glc ² — ¹ Glc	H
7	H	Glc ² — ¹ Glc	H
8	OH	Glc	Glc ⁶ — ¹ Glc
9	H	Glc A	H
10	OH	Glc ² — ¹ Glc	Glc ⁶ — ¹ Glc
11	H	Glc A	Glc ⁶ — ¹ Glc ¹ — ¹ Rha
12	OH	Ara	Glc ⁶ — ¹ Glc ¹ — ¹ Rha
13	H	Ara ² — ¹ Rha	Glc ⁶ — ¹ Glc ¹ — ¹ Rha
14	OH	Ara ² — ¹ Rha	Glc ⁶ — ¹ Glc ¹ — ¹ Rha
15	H	Glc A ² — ¹ Rha	Glc ⁶ — ¹ Glc ¹ — ¹ Rha

Ara : α -L-arabinopyranosyl
 Rha : α -L-rhamnopyranosyl
 Xyl : β -D-xylopyranosyl
 Glc : β -D-glucopyranosyl
 Glc A : β -D-glucuronopyranosyl

Fig. 1. List of Triterpenoid Glycosides Isolated from the Berries of *Hedera colchica*

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Table 1. ^{13}C -NMR Data for Sugar Moieties of Saponins **1**, **3**, **8**, **9**, **11** and **15** (CD_3OD)

1		3		8		9		11		15	
Sugar on C-3 ^{a)}											
ara 1	107.08	xyl 1	106.30	glc 1	105.72	glc A1	106.97	glc A1	106.96	glc A1	106.83
2	72.84	2	75.55	2	75.62	2	76.52	2	76.55	2	75.37
3	74.34	3	78.14	3	78.33	3	77.70	3	77.72	3	83.59
4	69.46	4	71.26	4	71.53	4	73.22	4	73.26	4	71.85
5	66.29	5	66.84	5	77.72	5	75.32	5	75.32	5	76.71
				6	62.72	6	172.91	6	175.97	6	176.06
										rham 1	102.72
										2	72.26
										3	72.36
										4	74.04
										5	69.94
										6	17.87
Sugar on C-28 ^{a)}											
				glc 1	95.75			glc 1	95.77	glc 1	95.76
				2	73.83			2	73.86	2	73.84
				3	79.79			3	79.59	3	79.52
				4	70.94			4	70.99	4	70.95
				5	78.16 ^{b)}			5	78.06	5	78.06
				6	69.49			6	69.42	6	69.41
				glc 1	104.63			glc 1	104.28	glc 1	104.27
				2	75.12			2	75.32	2	75.31
				3	77.99 ^{b)}			3	76.84	3	76.84
				4	71.53			4	78.21	4	78.18
				5	77.81			5	76.74	5	76.71
				6	62.72			6	61.88	6	61.87
								rham 1	102.96	rham 1	102.94
								2	72.24	2	72.23
								3	72.44	3	72.43
								4	73.78	4	73.76
								5	70.67	5	70.65
								6	17.85	6	17.87

a) ara = α -L-arabinopyranosyl; xyl = β -D-xylopyranosyl; glc = β -D-glucopyranosyl; glc A = β -D-glucuronopyranosyl; rham = α -L-rhamnopyranosyl. b) Assignments may be reversed.

onated between 172 and 176 ppm depending on the pH of the saponin solutions. It can be concluded that **15** was 3-O-[- α -L-rhamnopyranosyl-(1 \rightarrow 3)- β -D-glucuronopyranosyl]-28-O-[- α -L-rhamnopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl]-oleanolate.

Experimental

FAB-MS were obtained from a Nermag R-10-10H mass spectrometer in the negative ion mode. ^1H - and ^{13}C -NMR spectra were recorded on a Bruker AMX-400 spectrometer and chemical shifts are given on a δ (ppm) scale with tetramethylsilane (TMS) as the internal standard. Melting points were determined on an Electrothermal IA 9300 apparatus. Optical rotations $[\alpha]_D^{25}$ were measured on a Perkin-Elmer model 341 Orot polarimeter. TLC analyses of saponins and sugars were performed on precoated silica gel plates (Kieselgel 60F254, Merck) using the following solvent systems: CHCl_3 -MeOH- H_2O (26:14:3) [system 1]; *n*-BuOH-HOAc- H_2O (4:1:5) [system 2]; CHCl_3 -MeOH (20:1) [system 3]; CH_2Cl_2 -MeOH- H_2O (50:25:5) [system 4]. Spots were detected by spraying the plates with phosphoric acid naphthoresorcinol for sugars and H_2SO_4 for saponins and genins followed by heating at 110 °C.

Extraction and Separation Plant material was collected in the Bagdathi region of Georgia (January 1996) and dried in the shade. A voucher specimen is kept in the Department of Pharmacobotanic, Institute of Pharmacology, Tbilisi, Georgia (berries No. 80197). Crushed berries (720 g) were extracted with MeOH- H_2O (80:20). After concentration, the aqueous layer was treated with CHCl_3 , then with BuOH to obtain a crude extract of saponins. The BuOH extract (146 g) was subjected to column chromatography (CC) on silica gel (0.04–0.063 mm, Merck) eluting with CHCl_3 -MeOH- H_2O (26:14:3) to afford 3 fractions. Fraction 1 subjected to repeated CC, eluting with CHCl_3 -MeOH (from 9:1 to 4:1) yielded **1** (11 mg), **2** (80 mg), **3** (51 mg), **4** (600 mg), **5** (1.0 g) and a mixture of **6** and **7** with other unidentified minor compounds. Saponins **6** and **7** were purified by

LPC on RP18 (15–25 μm , Merck), eluting with 80% MeOH to give **6** (45 mg) and **7** (1.0 g). Fraction 2 submitted to silica gel CC afforded saponin **14** (600 mg) and a mixture which treated by repeated CC on polyamide (SC6 0.07, Macherey-Najel) eluting with a gradient of MeOH in H_2O (from 10 to 65%) yielded **8** (75 mg), **9** (22 mg), **10** (57 mg), **12** (300 mg) and **13** (110 mg). Fraction 3 was chromatographed on a silica gel column eluting with CHCl_3 -MeOH- H_2O (55:40:10), then subjected to repeated purification on a polyamide column (MeOH from 30 to 50%) to give **11** (35 mg) and colchiside B (**15**) (100 mg).

Acid Hydrolysis of 3 and 15 The saponin (3 mg) was heated with aqueous 10% HCl (3 ml) in a sealed tube at 100 °C for 4 h. The sapogenin was extracted with Et_2O ; then the aqueous layer was neutralized with *N,N*-diethylmethylamine (10% in CHCl_3) and dried. The sapogenin and sugars were identified by TLC analysis with authentic samples in systems 3 and 4, respectively.

Alkaline Hydrolysis of 3 and 15 The saponin (5 mg) in 5% aqueous KOH (5 ml) was heated at 100 °C in a sealed tube for 90 min. After neutralization with 10% HCl (pH=5) the prosapogenin was extracted with BuOH. TLC analysis was performed using systems 1 and 2.

Colchiside A (**3**): White powder; R_f =0.87 (in system 1). $[\alpha]_D^{25}$ =+12.6° (MeOH). Neg. FAB-MS (m/z): 603 $[\text{M}-\text{H}]^-$ (Calcd for $\text{C}_{35}\text{H}_{56}\text{O}_8$). ^{13}C -NMR data for aglycone part (CD_3OD): 39.52 (C-1), 26.31 (C-2), 83.45 (C-3), 43.90 (C-4), 49.03 (C-5), 18.95 (C-6), 33.80 (C-7), 40.60 (C-8), 48.24 (C-9), 37.73 (C-10), 24.15 (C-11), 123.70 (C-12), 145.28 (C-13), 43.00 (C-14), 28.85 (C-15), 24.67 (C-16), 47.70 (C-17), 42.80 (C-18), 47.35 (C-19), 31.60 (C-20), 34.98 (C-21), 33.50 (C-22), 65.02 (C-23), 13.51 (C-24), 16.49 (C-25), 17.87 (C-26), 26.57 (C-27), 182.15 (C-28), 33.63 (C-29), 24.07 (C-30). For sugars see Table 1.

Colchiside B (**15**): White powder; R_f =0.12 (in system 1); mp 180 °C; $[\alpha]_D^{25}$ =+15° (MeOH). Neg. FAB-MS (m/z): 1247 $[\text{M}-\text{H}]^-$ (Calcd for $\text{C}_{60}\text{H}_{96}\text{O}_{27}$). 1101 $[(\text{M}-\text{H})-146]^-$; 777 $[(\text{M}-\text{H})-146-2\times 162]^-$; ^{13}C -NMR data for aglycone part (CD_3OD): 39.8 (C-1), 26.9 (C-2), 91.2 (C-3), 40.73 (C-4), 57.02 (C-5), 17.9 (C-6), 33.28 (C-7), 40.19 (C-8), 48.81 (C-9),

37.92 (C-10), 24.59 (C-11), 123.85 (C-12), 144.85 (C-13), 42.94 (C-14), 28.95 (C-15), 24.04 (C-16), 48.07 (C-17), 42.55 (C-18), 47.25 (C-19), 31.57 (C-20), 34.91 (C-21), 33.95 (C-22), 28.54 (C-23), 17.05 (C-24), 16.13 (C-25), 17.87 (C-26), 26.34 (C-27), 178.10 (C-28), 33.52 (C-29), 24.15 (C-30). For sugars see Table 1.

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References

- 1) Sakartvelos Flora, *Metsniereba*, Tbilisi, 1984, IX, 124.
- 2) Rastitelnie, Resursi SSSR, Nauka, Leningrad *Fam. Rutaceae-Elaeagnaceae*, 1988, 4, 62.
- 3) a) Dekanosidze G. E., Pkheidze R. A., Gorovits T., Kemertelidze E. P., *Khim. Prir. Soedi.*, **4**, 484—485 (1970); b) Dekanosidze G. E., Djikia O. D., Vulgalter M. M., Kemertelidze E. P., *ibid.*, **6**, 747—749 (1990).
- 4) Mshvildadze V. D., Dekanosidze G. E., Kemertelidze E. P., Shashkof A. S., *Bioorganicheskaia Khimia*, **62**, 1001—1007 (1993).
- 5) Wang H. B., Mayer R., Ruecker G., *Phytochemistry*, **33**, 1469 (1993).
- 6) Jiang Q. P., Xiao Z. Y., *West China J. Pharm. Sci.*, **5**, 133 (1990).
- 7) Hostettmann K., *Helv. Chim. Acta*, **63**, 606—609 (1980).
- 8) Kizu H., Kitayama S., Nakatani F., Tomimori T., Namba T., *Chem. Pharm. Bull.*, **33**, 3324—3329 (1985).
- 9) Haar V. D., *Ber. Deutsch. Chem.*, **54**, 3142—3149 (1921).
- 10) Loloiko A. A., Grishkovets V. I., Shashkov A. S., Chirva V. I., *Khim. Prir. Soedi.*, **5**, 721—726 (1988).
- 11) Grishkovets V. I., Loloiko A. A., Shashkov A. S., Chirva V. I., *Khim. Prir. Soedi.*, **6**, 779—783 (1984).
- 12) Tschesche R., Schmidt W., Wulff Gr., *Zhur. Naturforsch.*, **20**, 708—709 (1965).
- 13) Elias R., Diaz-Lanza A. M., Vidal-Ollivier E., Balansard G., Faure R., Babadjamian A., *J. Nat. Prod.*, **54**, 98—103 (1991).
- 14) Crespin F., Ollivier E., Lavaud C., Babadjamian Faure R., Debrauwer L., Balansard G., *Phytochemistry*, **33**, 657—661 (1993).
- 15) Breitmaier E., Voelter W., “¹³C-NMR Spectroscopy,” 1978 Edition, Verlag, New York.
- 16) Agrawal P. K., “Carbon-13-NMR of Flavonoids in studies in Organic Chemistry, 39,” 1989 Edition, Elsevier, New York.