Synthesis and Pharmacological Analysis of High Affinity Melatonin Receptor Ligands

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We report the synthesis and radioligand binding analysis of a series of naphthalenic melatonin receptor ligands, *N***-[2-(7-alkoxy-2-methoxy-1-naphthyl)ethyl]propionamide. This series of ligands exhibits subpicomolar** binding affinity to both MT₁ and MT₂ melatonin receptors expressed in chinese hamster ovary (CHO) cells.

Key words melatonin; MT_1 , MT_2 receptors/receptor binding; naphthalenic melatonin receptor ligands

Melatonin is a hormone that is involved in a variety of physiological and pathological responses such as circadian rhythms, retinal physiology, seasonal breeding, cardiovascular regulation, anti-oxidative activity and oncogenesis. $1 - 7$) Except anti-oxidative activity, melatonin may mediate its effect *in vivo* through melatonin receptors. Presently, two human melatonin receptor subtypes exist and are now defined as either the MT_1^{8} (formerly known as the Mel_{1a} receptor)⁹ and the MT₂⁸ (formerly known as the Mel_{1b} receptor).¹⁰⁾ The ability of melatonin or melatonin-like compounds to bind to and elicit physiological responses is dependent upon specific interactions between the ligand and receptor. The elucidation of the components of melatonergic ligands that are involved in high-affinity binding is becoming clearer. Currently, what we know is that the indole ring of melatonin is not crucial for melatonin receptor recognition. For instance, the naphthalene ring^{11—20)} can serve as a bioisostere of the indole nucleus of melatonin and analog **2** was reported to have equivalent affinity to melatonin for melatonin receptors in ovine pars tuberalis. $11)$ In addition, other groups such as amidotetralin,²¹⁾ methoxychroman,²²⁾ amido indane,²³⁾ benzofuran,^{18,24)} benzothiophene^{18,24)} and quinoline²⁵⁾ can serve as a bioisostere of the indole nucleus of melatonin. Langlois *et al.* reported that the addition of a 2-methoxy group (OMe) to compound **2** to form **3** results in an order of magnitude increase in receptor affinity over compound 2 (Fig. 1).¹²⁾ It was postulated that the 2-OMe group binds to the accessory binding site of the receptor. Recently, we reported the synthesis

and receptor binding studies of several analogs of **3** (general structure **4**) with the substituents either hydrophilic or hydrophobic in nature with different sizes.¹⁶⁾ Preliminary results show that analogs with smaller substituents in R, irrespective of their hydrophilic or hydrophobic nature, exhibit higher affinity for melatonin receptors when compared to those with larger substituents. We postulate that the close proximity of the amidoethyl side chain may be interfering with the conformation of the substituent on 2-position in **4** (Fig. 1). Thus, to determine the validity of this hypothesis, the amidoethyl group was transferred from C1 to C8 to eliminate the steric interference between the amidoethyl group and the substituent on 2-position in compound **4**. The transfer should not affect the affinity of the ligands for the melatonin receptor as shown by Langlois *et al.* in which compound **5** ($K_i = 0.67$ nm) has the same affinity for melatonin receptors in chicken brain when compared to compound **2** $(K_i=0.54 \text{ nm})$ (Fig. 1).¹²⁾ After eliminating the steric interference of the amidoethyl side chain and the substituents on 2 position, we then synthesized compounds **6**—**10** with the substituents on 7-position of varying sizes.

Results and Conclusions

Chemistry The synthesis of compounds **6**—**10** is summarized in Fig. 2. Monobenzylation of 2,7-dihydroxynaphthalene **11** by reaction with 1 eq of benzyl bromide to form **12** (39%) followed by selective formylation at the 1-position (Reimer–Tiemann reaction)²⁶⁾ yielded the desired phenolic

Fig. 1. Structures of Compounds **1**—**10**

Fig. 2. Synthesis of Compounds **6**—**10**

Reagents and Conditions: a. PhCH₂Br, K₂CO₃, acetone, reflux, 4 h, 39%; b. i) NaOH, CHCl₃, H₂O, reflux, 2 h; ii) H₃O⁺, 14%; c. CH₃I, K₂CO₃, acetone, reflux, 20 h, 95%; d. CH₃NO₂, NH₄OAc, reflux, 4 h, 100%; e. i) LiAlH₄, THF, 40 °C, 18 h; ii) C₂H₃COCl, Et₃N, CH₂C₁, r.t., 1 h, 63%; f. 10% Pd–C, MeOH, H₂, r.t., 18 h, 100%; g. RI (R=C₂H₅, C₃H₇, C_4H_9), \bar{K}_2CO_3 , acetone, reflux, 18 h, 76—94%.

aldehyde **13** (14%). The structure of **13** was confirmed unambiguously by ¹H-NMR spectra in which four protons have a large coupling constant of 9.0 Hz and the proton of OH moved downfield to 13.15 ppm. Methylation of **13** with methyl iodide formed **14** which was condensed with nitromethane in the presence of NH4OAc to furnish nitroalkene **15** (95% yield in 2 steps). Reduction of 15 with LiAlH₄ followed by acylation with propionyl chloride afforded amide **10** (63% yield based on **15**). The benzyl group in **10** was removed in Pd–H₂ to give phenol **6** (100%). Treatment of **6** with various alkyl iodides yielded the corresponding alkylated products **7**—**9** (76—94% yield).

Melatonin Receptors Ligands' Binding Affinity The affinities of compounds **6**—**10** for melatonin receptors were evaluated *in vitro* and in duplicate by competition binding analysis using the radioligand $2-[^{125}]]$ -iodomelatonin (80— 100 pM; DuPont, Boston, MA, U.S.A.) as described previously.²⁷⁾ The competition binding assays were performed $3-$ 5 times for each compound. The affinity of $2-[125]$ -iodomelatonin for the MT_1 and MT_2 melatonin receptors was 80 and 150 pM, respectively. To ensure that the affinities of compounds **6**—**10** for the melatonin receptors were not artifact, these compounds, like melatonin, were dissolved in ethanol, the pipette tips were changed in between serial dilutions, and melatonin competition curves were run in parallel with every compound tested. As shown in Table 1, compounds **6**—**10** displayed higher affinity than melatonin and exhibited subpicomolar binding affinities to both MT_1 and MT_2 receptors. The affinity of these compounds for the melatonin receptors increased from 0.85 pM to ≤ 0.01 pM as the R' substituent increased from H to *n*-Bu. A further increase in size of the R' substituent (benzyl) resulted in a decrease in binding affinity. However, this is in complete contrast to the affinities of compound 4 for MT_1 and MT_2 melatonin receptors in regard to varying the size of the R' substituent. In compound 4, the compound with the highest affinity to both MT_1 and MT_2 receptors are $R' = CH_3$ and C_2H_5 . Any increase in size in the R' substituent results in a decrease in binding affinity.¹⁶⁾ It has been stated that the 5-methoxy group in melatonin is involved in critical hydrogen bonding to the receptor recognition site.28) If it is assumed that the methoxy group in compound **4** mimics the 5-methoxy group of melatonin, then

Table 1. Competition of Melatonin and Substituted Naphthalene for 2- [125 I]-iodomelatonin Binding to Human MT₁ or MT₂ Melatonin Receptors Stably Expressed in CHO Cells

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ŌК CH ₃ O ₂

transferring the amidoethyl group from C1 to C8 eliminated the steric interference between the amidoethyl group and the 2 substituent in compound **4** and resulted in an increase in binding affinity to both MT_1 and MT_2 receptors.

In conclusion, A series of *N*-[2-(7-alkoxy-2-methoxy-1 naphthyl)ethyl]propionamides were prepared and their binding affinities on both MT_1 and MT_2 melatonin receptors were evaluated. The analogs exhibit high binding affinities to the receptors and the affinity of these compounds for the melatonin receptors decreased from 0.85 pM to ≤ 0.01 pM as the R['] substituent increased from H to *n*-Bu. A further increase in size of the R['] substituent (benzyl) resulted in a decrease in binding affinity.

Experimental

The Preparation of 7-(Benzyloxy)-2-naphthol (12) To a solution of 20.8 g (130 mmol) of 2,7-dihydroxynaphthalene **11** was added 15.2 g (110 mmol) of benzyl bromide. The reaction mixture was refluxed for 4 h and cooled to room temperature (r.t.). The reaction mixture was filtered and the filtrate was concentrated *in vacuo.* The residue was purified by column chromatography on silica gel using CH_2Cl_2 –petroleum ether (4 : 1) as eluent, giving 10.8 g (39%) of monobenzylated product 12; mp $151.5 - 152$ °C; ¹H-NMR (300 MHz, DMSO-*d*₆) δ 5.17 (s, 2H, PhCH₂), 6.89–6.99 (m, 2H, ArH), 7.03 (d, 1H, *J*=2.1 Hz, ArH), 7.20 (d, 1H, *J*=2.1 Hz, ArH), 7.31— 7.68 (m, 7H, ArH), 9.66 (s, 1H, OH). *Anal*. Calcd for C₁₇H₁₄O₂: C, 81.58; H, 5.64. Found: C, 81.62; H, 5.47.

The Preparation of 7-(Benzyloxy)-2-hydroxy-1-naphthylaldehyde (13) 7-Benzyloxy-2-naphthol **12** (5.0 g, 20 mmol) was added to a solution of NaOH (6.4 g, 0.16 mol) in 40 ml of water and the mixture was heated to 65—70 °C. Chloroform (3.6 ml, 44.8 mmol) was added to the mixture in three portions at intervals of 15 min . After the addition of CHCl₂, the reaction mixture was stirred at 100 °C for 1 h. After cooling, the reaction mixture was acidified with diluted sulfuric acid and then extracted with CH₂Cl₂ $(3\times60 \text{ ml})$. The organic extracts were dried (Na_2SO_4) and evaporated. The residue was purified by silica gel chromatography using $CH₂Cl₂$ –petroleum ether $(3:1)$ as eluent, yielding 0.75 g $(14%)$ of pure aldehyde 13. mp 125– 126.5 °C; ¹H-NMR (300 MHz, CDCl₃) δ 5.20 (s, 2H, PhCH₂), 6.95 (d, 1H, *J*=9.0 Hz, ArH), 7.13 (dd, 1H, *J*=2.1, 9.0 Hz, ArH), 7.43 (m, 5H, ArH), 7.69 (d, 1H, *J*=9.0 Hz, ArH) 7.72 (d, 1H, *J*=2.1 Hz, ArH), 7.86 (d, 1H, *J*=9.0 Hz, ArH), 10.65 (s, 1H, CHO), 13.15 (s, 1H, OH, D₂O exchangeable). *Anal.* Calcd for C₁₈H₁₄O₃: C, 77.68; H, 5.07. Found: C, 77.46; H, 5.38.

The Preparation of 7-(Benzyloxy)-2-methoxy-1-naphthylaldehyde (14) To a solution of the aldehyde **13** (1.6 g, 5.76 mmol) in dry acetone (60 ml) was added methyl iodide (CH₂I, 1.2 ml, 19.3 mmol) and K_2CO_3 (1.2 g, 8.7 mmol). The reaction mixture was refluxed for 20 h and then filtered to remove the inorganic salt. Concentration of the filtrate gave the crude product (as a solid) which was washed with petroleum ether to remove excess methyl iodide furnishing 1.88 g (95%) of pure **14** after silica gel column chromatography (petroleum ether/CH₂Cl₂, 1:2). mp 111.5—112.5 °C; ¹H-NMR (300 MHz, CDCl₃) δ 4.01 (s, 3H, OCH₃), 5.22 (s, 2H, CH₂Ph), 7.07—7.53 (m, 7H, ArH), 7.65 (d, 1H, *J*=9.0 Hz, ArH), 7.95 (d, 1H, *J*=9.0 Hz, ArH), 8.95 (d, 1H, $J=2.3$ Hz, ArH), 10.87 (s, 1H, CHO). *Anal*. Calcd for C₁₉H₁₆O₃: C, 78.06; H, 5.52. Found: C, 78.03; H, 5.62.

The Preparation of 7-(Benzyloxy)-2-methoxy-1-[(E)-2-nitroethenyl] naphthalene (15) A solution of the aldehyde **14** (1.72 g, 5.9 mmol) and ammonium acetate (0.3 g, 3.9 mmol) in nitromethane (18.4 ml) was refluxed for 4 h. After evaporation of the solvent *in vacuo*, the residue was dissolved in CH₂Cl₂ (30 ml) and H₂O (25 ml) was added. The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2×25 ml). The combined organic extracts were washed with $H₂O$ (30 ml) and then dried (Na₂SO₄). Evaporation of the solvent under reduced pressure gave 1.97 g $(\sim 100\%)$ of pure product 15; mp 163—164.5 °C; ¹H-NMR (300 MHz, CDCl₃) δ 4.05 (s, 3H, OCH₃), 5.28 (s, 2H, CH₂Ph), 7.07 (d, 1H, *J*=9.0 Hz, ArH), 7.14 (dd, 1H, *J*=2.4, 9.0 Hz, ArH), 7.33—7.53 (m, 6H, ArH), 7.70 (d, 1H, *J*59.0 Hz, ArH), 7.85 (d, 1H, *J*59.0 Hz, ArH), 8.08 (d, 1H, *J*513.2 Hz, CHNO₂), 8.68 (d, 1H, $J=13.2$ Hz, ArH). *Anal.* Calcd for $C_{20}H_{17}NO₄$: C, 71.63; H, 5.11; N, 4.18. Found: C, 71.82; H, 5.03; N, 4.26.

The Preparation of *N***-[2-(7-(Benzyloxy)-2-methoxy-1-naphthyl)ethyl] propionamide (10)** To a stirred suspension of $LiAlH₄$ (1.0 g, 26.3 mmol) in anhydrous tetrahydrofuran (THF) (60 ml) at 0° C was added dropwise a solution of compound **15** (1.5 g, 4.48 mmol) in THF (20 ml). The reaction mixture was stirred at 40 °C for 18 h and then cooled to 0 °C. Solutions were then added to the mixture in the following order: $H₂O$ (1.0 ml), 15% NaOH solution (1.0 ml) , EtOAc (50 ml) and $H₂O$ (30 ml) . The mixture was filtered, and the filtrate was dried (Na₂SO₄). The solvent was evaporated *in vacuo* and the crude product thus obtained was dried overnight *in vacuo* and used for the next step without further purification.

To a solution of the above crude product in anhydrous CH_2Cl_2 (25 ml) at 0° C were added triethylamine (1.2 ml, 8.6 mmol) in one portion and then propionyl chloride (0.5 ml, 5.72 mmol) dropwise. The reaction mixture was stirred at r.t. for 1 h and was diluted with CH_2Cl_2 (30 ml), washed with saturated NaHCO₃ solution (20 ml) and H₂O (20 ml). The organic layer was dried (Na_2SO_4) and concentrated. The residue was purified by silica gel chromatography using petroleum ether : EtOAc (1 : 1) as eluent, yielding 880 mg (54% based on compound **15**) of amide **10**; mp 121.5—122.5 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.06 (t, 3H, *J*=7.5 Hz, CH₃), 2.10 (q, 2H, *J*=7.5 Hz, CH₂CO), 3.23 (t, 2H, *J*=6.9 Hz, ArCH₂), 3.50 (m, 2H, CH₂N), 3.93 (s, 3H, OCH3), 5.22 (s, 2H, CH2Ph), 5.76 (br s, 1H, NH), 7.08 (dd, 1H, *J*52.4, 9.0 Hz, ArH), 7.10 (d, 1H, *J*59.0 Hz, ArH), 7.32—7.50 (m, 6H, ArH), 7.67 (d, 1H, *J*59.0 Hz, ArH), 7.68 (d, 1H, *J*59.0 Hz, ArH). *Anal.* Calcd for $C_{23}H_{25}NO_3$: C, 76.01; H, 6.93; N, 3.85. Found: C, 76.15; H, 6.82; N, 3.96.

The Preparation of *N***-[2-(7-Hydroxy-2-methoxy-1-naphthyl)ethyl]propionamide (6)** Amide **10** (1.2 g, 3.3 mmol) in methanol (38 ml) was hydrogenated over 10% Pd on activated carbon at r.t. for 18 h. The reaction mixture was filtered through celite. The filtrate was concentrated and purified by silica gel chromatography using petroleum ether–EtOAc (1 : 2) as eluent, yielding $0.9 g$ (\sim 100%) of compound 6; mp 118—120 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.02 (t, 3H, *J*=7.5 Hz, CH₃), 2.07 (q, 2H, *J*=7.5 Hz, CH₂CO), 3.03 (t, 2H, *J*=6.6 Hz, ArCH₂) 3.18 (m, 2H, CH₂N), 3.87 (s, 3H, OCH₃), 6.92 (dd, 1H, *J*=1.8, 9.0 Hz, ArH), 7.16 (d, 1H, *J*=9.0 Hz, ArH), 7.23 (d, 1H, $J=1.8$ Hz, ArH), 7.68 (d, 2H, $J=9.0$ Hz, ArH), 7.92 (brt, 1H, *J*=5.1 Hz, ArH), 9.64 (s, 1H, OH). *Anal.* Calcd for C₁₆H₁₉NO₃: C, 70.31; H, 7.01; N, 5.12. Found: C, 70.42; H, 6.87; N, 4.98.

The Preparation of *N***-[2-(7-Ethoxy-2-methoxy-1-naphthyl)ethyl]propionamide (7)** To a solution of phenol **6** (100 mg, 0.366 mmol) in acetone (6 ml) was added K_2CO_3 (76 mg, 0.55 mmol) followed by ethyl iodide (0.12 ml, 1.5 mmol). The reaction mixture was refluxed for 18 h and then cooled, filtered. The filtrate was concentrated *in vacuo* and the residue was purified by flash chromatography using ethyl acetate–methylene chloride $(1:1)$ as the eluent, yielding pure 7 (95 mg, 86%); mp 119—120.5 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.06 (t, 3H, J=7.5 Hz, CH₃), 1.47 (t, 3H, *J*=6.9 Hz, CH₃), 2.10 (q, 2H, *J*=7.5 Hz, CH₂CO), 3.25 (t, 2H, *J*=6.7 Hz, ArCH₂), 3.53 (m, 2H, CH₂N), 3.93 (s, 3H, OCH₃), 4.17 (q, 2H, *J*=6.9 Hz, CH₂O), 5.79 (br s, 1H, NH), 6.99 (dd, 1H, J=2.1, 9.0 Hz, ArH), 7.09 (d, 1H, *J*59.0 Hz, ArH), 7.26 (d, 1H, *J*52.1 Hz, ArH), 7.66 (d, 2H, *J*59.0 Hz, ArH). *Anal.* Calcd for C₁₈H₂₃NO₃: C, 71.73; H, 7.69; N, 4.65. Found: C, 71.52; H, 7.73 ; N, 4.53.

The Preparation of *N***-[2-(2-Methoxy-7-propoxy-1-naphthyl)ethyl]propionamide (8)** The synthesis of compound **8** is similar to the procedure for the synthesis of **7** using *n*-propyl iodide instead of iodoethane (94% yield); mp 111—112.5 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.07 (m, 6H, 2×CH₃), 1.86 (m, 2H, CH₂), 2.09 (q, 2H, *J*=7.5 Hz, CH₂CO), 3.26 (t, 2H, *J*=6.6 Hz, ArCH₂), 3.53 (m, 2H, CH₂N), 3.93 (s, 3H, OCH₃), 4.05 (t, 2H, *J*=6.3 Hz, CH₂O), 5.79 (br s, 1H, NH), 6.99 (dd, 1H, *J*=2.1, 9.0 Hz, ArH), 7.09 (d, 1H, $J=9.0$ Hz, ArH), 7.25 (d, 1H, $J=2.1$ Hz, ArH), 7.66 (d, 2H, *J*=9.0 Hz, ArH). *Anal.* Calcd for C₁₉H₂₅NO₃: C, 72.35; H, 7.99; N, 4.44. Found: C, 72.63; H, 7.87; N, 4.51.

The Preparation of *N***-[2-(7-Butoxy-2-methoxy-1-naphthyl)ethyl]propionamide (9)** The synthesis of compound **9** is similar to the procedure for the synthesis of **7** using *n*-butyl iodide instead of iodoethane (76% yield); mp 92—93 °C; ¹H-NMR (300 MHz, CDCl₃) δ 0.99 (t, 3H, *J*=7.5 Hz, CH₃), 1.53 (m, 2H, CH₂), 1.84 (m, 2H, CH₂), 2.09 (q, 2H, $J=7.5$ Hz, CH₂CO), 3.26 (t, 2H, *J*=6.6 Hz, ArCH₂), 3.54 (m, 2H, CH₂N), 3.93 (s, 3H, OCH₃), 4.09 (t, 2H, *J*=6.3 Hz, CH₂O), 5.80 (br s, 1H, NH), 6.99 (dd, 1H, *J*=2.1, 9.0 Hz, ArH), 7.09 (d, 1H, *J*=9.0 Hz, ArH), 7.25 (d, 1H, *J*=2.1 Hz, ArH), 7.66 (d, 2H, *J*=9.0 Hz, ArH). *Anal*. Calcd for C₂₀H₂₇NO₃: C, 72.92; H, 8.26; N, 4.25. Found: C, 72.88; H, 8.19; N, 4.31.

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References

- 1) Morgan P. J., Barrett P., Howell H. E., Helliwell R., *Neurochem. Int*., **24**, 101—146 (1994).
- 2) Dubocovich M. L., *Trends Pharmacol. Sci*., **16**, 50—56 (1995).
- 3) Krause D. N., Barrios V. E., Duckles S. P., *Eur. J. Pharmacol*., **276**, 207—213 (1995).
- 4) Brzezinski A., *N. Engl. J. Med*., **336**, 186—195 (1997).
- 5) Ting K. N., Dunn W. R., Davies D. J., Sugden D., Delagrange P., Guardiola-Lemaitre B., Scalbert E., Wilson V. G., *Br. J. Pharmacology*, **122**, 1299—1306 (1997).
- 6) Gozzo A., Lesieur D., Duriez P., Fruchart J. C., Teissier E., *Free Radic. Biol. Med*., **26**, 1538—1543 (1999).
- Bucher B., Gauer F., Pevet P., Masson-Pevet M., *J. Cardiovasc. Pharmacol*., **33**, 316—322 (1999).
- 8) Dubocovich M. L., Cardinali D. P., Guardiola-Lemaitre R. M., Hagan R. M., Krause D. N., Sugden D., Vanhoutte P. M., Yocca F. D., "The IUPHAR Compendium of Receptor Characterization and Classification IUPHAR Media," London, 1998, pp. 187—193.
- 9) Reppert S. M., Weaver D. R., Ebisawa T., *Neuron*, **13**, 1177—1185 (1994).
- 10) Reppert S. M., Godson C., Mahle C. D., Weaver D. R., Slaugenhaupt S. A., Gusella, J. F., *Proc. Natl. Acad. Sci*., **92**, 8734—8738 (1995).
- 11) Yous S., Andrieux J., Howell H. E., Morgan P. J., Renard P., Pfeiffer B., Lesieur D., Guardiola-Lemaitre B., *J. Med. Chem*., **35**, 1484—1486 (1992).
- 12) Langlois M., Bremont B., Shen S., Poncet A., Andrieux J., Sicsic S., Serraz I., Mathe-Allainmat M., Renard P., Delagrange P., *J. Med. Chem*., **38**, 2050—2060 (1995).
- 13) Leclerc V., Depreux P., Lesieur D., Caignard D. H., Renard P., Delagrange P., Buardiola-Lemaitre B., Morgan, P., *Bioorg. & Med. Chem. Lett*., **6**, 1071—1076 (1996).
- 14) Mathe-Allainmat M., Andrieux J., Langlois M., *Exp. Opin. Ther.*

Patents, **7**, 1447—1458 (1997).

- 15) Pang C. S., Tang P. L., Song Y., Pang S. F., Ng K. W., Guardiola-Lemaitre B., Delagrange P., Brown G. M., *J. Pineal Res*., **23**, 148— 155 (1997).
- 16) Li P. K., Chu G. H., Gillen M. L., Parekh T., Witt-Enderby P. A., *Bioorg. & Med. Chem. Lett*., **7**, 2409—2414 (1997).
- 17) Witt-Enderby P. A., Chu G. H., Gillen M. L., Li P. K., *J. Med. Chem*., **40**, 4195—4198 (1997).
- 18) Leclerc V., Fourmaintraux E., Dupreux P., Lesieur D., Morgan M., Howell H. E., Renard P., Caignard D. H., Pfeiffer B., Delagrange P., Guardiola-Lemaitre B., Andrieux J., *Bioorg. & Med. Chem*., **6**, 1875— 1887 (1998).
- 19) Masson-Pevet M., Recio J., Guerrero H. Y., Mocaer E., Delagrange P., Guardiola-Lemaitre B., Pevet P., *J. Pineal Res*., **25**, 172—176 (1998).
- 20) Andries V., Depreux P., *Synth. Commun*., **27**, 4207—4214 (1997).
- 21) Copinga S., Tepper P. G., Grol C. J., Horn A. S., Dubocovich M. L., *J.*

Med. Chem., **36**, 2891—2898 (1993).

- 22) Sugden D., *Eur. J. Pharmacol*., **254**, 271—275 (1994).
- 23) Jansen J. M., Copinga S., Gruppen G., Molinar E. J., Dubocovich M. L., Grol C. J., *Bioorg. & Med. Chem*., **4**, 1321—1332 (1996).
- 24) Depreux P., Lesieur D., Mansour H. A., Morgan P., Howell H. E., Renard P., Caignard D. H., Pfeiffer B., Delagrange P., Guardiola B., Yous S., Demarque A., Adam G., Andrieux J., *J. Med. Chem*., **37**, 3231— 3239 (1994).
- 25) Li P. K., Chu G. H., Gillen M. L., Witt-Enderby P. A., *Bioorg. & Med. Chem. Lett*., **7**, 2177—2180 (1997).
- 26) Wynberg H., *Chem. Rev*., **60**, 169—184 (1960).
- 27) Witt-Enderby P. A., Dubocovich M. L., *Mol. Pharmacol*., **50**, 166— 174 (1996).
- 28) Sugden D., Chong N. W. S., Lewis D. F. V., *Br. J. Pharmacol*., **114**, 618—623 (1995).