

Additional Cytotoxic Diacetylenes from the Stony Coral *Montipora* sp.

Naseer ALAM,^a Jongki HONG,^b Chong-Ok LEE,^c Jae Sue CHOI,^d Kwang Sik IM,^a and Jee H. JUNG^{*a}

^a College of Pharmacy, Pusan National University; Pusan 609–735, Korea; ^b Korea Basic Science Institute; Seoul, Korea;

^c Korea Research Institute of Chemical Technology; Taejon, Korea; and ^d Pukyong National University; Pusan, Korea.

Received November 12, 2001; accepted January 22, 2002

Three new diacetylenes (1, 4, 6) have been isolated as cytotoxic constituents from the methanolic extract of the stony coral *Montipora* sp. The structures have been elucidated on the basis of spectroscopic evidence. The compounds were evaluated for cytotoxicity against a small panel of human tumor cell lines and showed moderate to significant activity.

Key words stony coral; *Montipora* sp.; diacetylene; cytotoxicity

The secondary metabolites of the stony corals so far investigated are few but diverse.¹⁾ Among them, diacetylene derivatives have been isolated from the genera *Montipora* and *Pectinia*.^{1–4)} *Montipora* sp. (Scleractinia, Coelenterata) is a hermaphroditic coral and has been investigated from both ecological and chemical viewpoints.

We previously reported several new diacetylenic compounds (montiporyne A–M) from *Montipora* sp. collected from Korean waters.^{1,2)} Some have shown significant cytotoxic activity. Subsequently, we isolated additional congeners by activity-guided fractionation of the methanolic extract of the same coral. Here, we report the isolation and structure elucidation of three additional diacetylenes (**1**, **4**, **6**) that share the same gross structure with the compounds reported earlier.¹⁾

Compound **1** was isolated as a light yellow oil. ¹H-NMR signals of **1** resembled those of methyl montiporate B (**2**) reported earlier from the same source.¹⁾ Two singlets of two protons each at δ 4.33 and 4.17 were due to H-1 and H-2' protons, respectively, while the methoxy protons resonated at δ 3.74. The signals at δ 5.80 (ddt), 5.00 (dd), and 4.95 (dd) were assigned to the terminal olefinic protons. A triplet at δ 2.30 was attributed to the α -acetylenic methylene protons (H-6). The compound showed an [M+Na]⁺ ion peak at m/z 271 in the FAB-MS analysis that matched well with the molecular formula C₁₅H₂₀O₃ which was assumed from the NMR data. All this evidence suggests that **1** is a lower-mass analogue of methyl montiporate B (**2**, C₁₇H₂₄O₃) and the structure was determined to be methyl 2-*O*-(11-dodecene-2,4-diynyl)-2-hydroxy ethanoate.

Compound **4** showed an isolated oxygenated methylene (δ 4.24, H-1) and an α -acetylenic methylene (δ 2.29, H-6) characteristic of montiporynes. Two triplets at δ 3.59 ($J=5.5$ Hz)

and 3.66 ($J=5.5$ Hz) were similar to those for the H-2' and H-1' protons in montiporyne H (**3**). The FAB-MS showed the [M+Na]⁺ ion at m/z 271, and thus the molecular formula was deduced to be C₁₆H₂₄O₂, which showed that compound **4** was a higher-mass analogue of montiporyne H (**3**, C₁₄H₂₀O₂).¹⁾ Thus the structure was assigned as 2-*O*-(13-tetradecene-2,4-diynyl)-1,2-ethanediol.

Compound **6** was a β -hydroxy ketone derivative similar to montiporyne J (**5**, C₁₅H₂₀O₂).¹⁾ Two doublets of doublets at δ 2.78 ($J=16.5, 5.5$ Hz) and 2.86 ($J=16.5, 8.0$ Hz) were due to H-3 protons, while the H-4 proton resonated as a doublet of doublets at δ 4.78 ($J=8.0, 5.5$ Hz). Signals corresponding to an α -acetylenic methylene and a monosubstituted olefin were also observed. The FAB-MS showed an [M+Na]⁺ ion at m/z 269, which corresponded to the molecular formula C₁₆H₂₂O₂Na. Thus the structure was elucidated to be 4-hydroxy-15-hexadecene-5,7-diyne-2-one. The stereochemistry at C-4 remains to be determined.

The isolated compounds were tested against a small panel of human cancer cell lines and the results are shown in Table 1. The compounds showed a structure–activity relationship (SAR) profile similar to their earlier reported congeners.¹⁾ Methyl montiporate C (**1**) was active only against a skin cancer cell line. Compound **4** was moderately active, while **6**, with β -hydroxy ketone functionality, exhibited more potent cytotoxicity.

Experimental

General NMR spectra were recorded on a Varian INOVA 500 spectrometer. Chemical shifts were reported in reference to the respective residual solvent peaks (δ_{H} 3.3 and δ_{C} 49.0 for CD₃OD). LR-FAB-MS data were obtained using a JEOL JMS-HX110/110A. HPLC was performed on a Gilson 370 pump with a YMC ODS-H80 (250×10 mm i.d., S-4 μm , 80 Å) column using a Shodex RI-71 detector.

Animal Material The animals were collected by hand using scuba gear at a depth of 8 m in November 1996, along the shore of Mundo, Cheju Is-

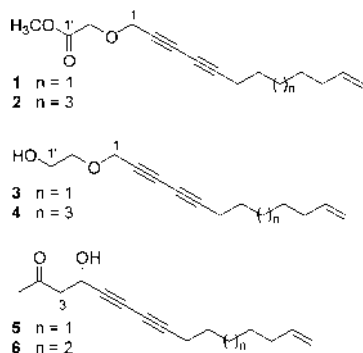


Table 1. Cytotoxicities (ED₅₀, $\mu\text{g/ml}$) of Compounds **1**, **4**, and **6** against Human Solid Tumor Cells^{a)}

Compound	A549	SK-OV-3	SK-MEL-2	XF498	HCT15
1	>30	27.1	5.51	>30	>30
4	11.29	13.80	4.36	12.97	8.43
6	6.20	4.78	3.85	7.24	6.94
Doxorubicin	0.02	0.13	0.03	0.08	0.04
Cisplatin	0.75	1.09	2.18	1.18	0.85

a) A549, human lung cancer; SK-OV-3, human ovarian cancer; SK-MEL-2, human skin cancer; XF498, human CNS cancer; HCT15, human colon cancer.

land, Korea, and were described in a previous report.²⁾ A voucher specimen was deposited in the Natural History Museum, Ewha Womans University (voucher no. EWUA. Ant. 961104).

Extraction and Isolation The frozen coral (2.5 kg, wet wt.) was extracted with MeOH at room temperature. Guided by the brine shrimp lethality assay,⁵⁾ the MeOH extract was partitioned between H₂O and EtOAc. The EtOAc layer was further partitioned between H₂O and CHCl₃ to afford 8.8 g of the CHCl₃ layer (LD₅₀ 30–86 μg/ml) which was subjected to a reverse-phase medium pressure liquid chromatography (MPLC) (YMC gel ODS-A, 60 Å 500/400 mesh) eluted with a step gradient solvent system of 25→0% H₂O/MeOH to obtain 14 fractions (1–14). Fraction 3 (3.1 g) was very active in the brine shrimp test and was further separated into 26 fractions on a reverse-phase MPLC (YMC gel ODS-A, 60 Å, 500/400 mesh), eluted with a step gradient solvent system of 20→0% H₂O/MeOH. Sub fraction 3-18 was repeatedly chromatographed on HPLC (YMC ODS-H80, 250×10 mm i.d., S-4 μm, 80 Å) eluted with 90% MeOH/H₂O to yield compound **1** (1 mg). Compound **4** (0.6 mg) was purified from subfraction 3-21 using 90% MeOH/H₂O on the same column. Compound **6** (0.8 mg) was purified from sub fraction 3-11 using the same column with 80% MeOH/H₂O as the mobile phase.

Methyl Montiporate C (1): Light yellow oil. UV (MeOH) λ_{max} nm (log ε): 286 (3.40), 226 (3.76). IR (film) ν_{max} cm⁻¹: 3376, 2927, 2856, 1716, 1071. ¹H-NMR (CD₃OD) δ: 1.25–1.55 (6H, m, H-7–H-9), 2.07 (quart, J=7.5 Hz, H-10), 2.30 (t, J=7.0 Hz, H-6), 3.74 (s, OCH₃), 4.17 (s, H-2'), 4.33 (s, H-1), 4.95 (dd, J=10.0, 2.5 Hz, H-12), 5.00 (dd, J=17.0, 2.5 Hz, H-12), 5.80 (ddt, J=17.0, 10.0, 7.0 Hz, H-11). ¹³C-NMR (CD₃OD) δ: 172.0 (C-1'), 139.8 (C-11), 115.4 (C-12), 82.4, 72.7, 71.9, 65.2 (C-2–5), 67.2 (C-2'), 59.8 (C-1), 52.3 (OCH₃), 34.4 (C-10), 29.3–29.8 (C-7–9), 19.7 (C-6). FAB-MS m/z: 271 [M+Na]⁺.

Dihomomontiporyne H (4): Light yellow oil. ¹H-NMR (CD₃OD) δ: 1.25–1.55 (10H, m, H-7–H-11), 2.05 (quart, J=7.0 Hz, H-12), 2.29 (t, J=7.0 Hz, H-6), 3.59 (t, J=5.5 Hz, H-2'), 3.66 (t, J=5.5 Hz, H-1'), 4.24 (s, H-1), 4.94

(dd, J=10.0, 2.0 Hz, H-14), 4.99 (dd, J=17.2, 2.0 Hz, H-14), 5.80 (ddt, J=17.2, 10.0, 6.5 Hz, H-13). ¹³C-NMR (CD₃OD) δ: 139.8 (C-13), 114.9 (C-14), 81.9, 72.8, 72.0, 65.3 (C-2–5), 72.4 (C-2'), 61.9 (C-1'), 59.7 (C-1), 34.6 (C-12), 29.2–29.6 (C-7–11), 19.7 (C-6). FABMS m/z: 271 [M+Na]⁺.

Homomontiporyne J (6): Light yellow oil. ¹H-NMR (CD₃OD) δ: 1.28–1.52 (8H, m, H-10–H-13), 2.06 (quart, J=7.0 Hz, H-14), 2.16 (s, H-1), 2.28 (t, J=7.0 Hz, H-9), 2.78 (dd, J=16.5, 5.5 Hz, H-3), 2.86 (dd, J=16.5, 8.0 Hz, H-3), 4.78 (dd, J=8.0, 5.5 Hz, H-4), 4.94 (dd, J=10.2, 2.0 Hz, H-16), 4.99 (dd, J=17.0, 2.0 Hz, H-16), 5.80 (ddt, J=17.0, 10.2, 6.8 Hz, H-15). ¹³C-NMR (CD₃OD) δ: 207.6 (C-2), 139.9 (C-15), 115.0 (C-16), 82.3, 76.8, 70.1, 65.3 (C-5–8), 59.0 (C-4), 51.8 (C-3), 34.6 (C-14), 29.3–29.8 (C-10–13), 30.6 (C-1), 19.7 (C-9). FAB-MS m/z: 269 [M+Na]⁺.

Acknowledgments Thanks are due to Professor Jun-Im Song for the identification of the coral. A grant from Pusan National University is gratefully acknowledged.

References and Notes

- 1) Alam N., Bae B. H., Hong J.-K., Lee C.-O., Im K. S., Jung J. H., *J. Nat. Prod.*, **64**, 1059–1063 (2001) and references cited therein.
- 2) Bae B. H., Im K. S., Choi W. C., Hong J.-K., Lee C.-O., Choi J. S., Son B. W., Song J.-I., Jung J. H., *J. Nat. Prod.*, **63**, 1511–1514 (2000).
- 3) Coll J. C., Bowden B. F., Meehan G. V., Konig G. M., Carroll A. R., Tapiolas D. M., Alino P. M., Heaton A., De Nys R., Leone P. A., Maida M., Aceret T. L., Willis R. H., Babcock R. C., Willis B. L., Florian Z., Clayton M. N., Miller R. L., *Mar. Biol.*, **118**, 177–182 (1994).
- 4) Higa T., Tanaka J., Kohagura T., Wauke T., *Chem. Lett.*, **1990**, 145–148 (1990).
- 5) Meyer B. N., Ferrigni N. R., Putnam J. E., Jacobsen L. B., Nichols D. E., McLaughlin J. L., *Planta Medica*, **45**, 31–34 (1982).