

Synthesis of Water-Soluble C₆₀-Porphyrin Hybrid Compounds

Kensuke OKUDA,^a Chiho ABETA,^a Takashi HIROTA,^a Masataka MOCHIZUKI,^b and Tadahiko MASHINO^{*,b}

^a Faculty of Pharmaceutical Sciences, Okayama University; 1-1-1 Tsushima-naka, Okayama 700-8530, Japan; and

^b Kyoritsu College of Pharmacy; 1-5-30 Shibakoen, Minato-ku, Tokyo 105-8512, Japan.

Received February 8, 2002; accepted March 14, 2002

Water-soluble C₆₀-porphyrin hybrid molecules were first synthesized toward their pharmaceutical applications.

Key words fullerene; porphyrin; antioxidant; superoxide; water-solubility

Much interest has been generated in fullerene and porphyrin because of their unique properties, especially their photochemical activity; thus, many researchers are studying fullerene and porphyrin interactions in order to develop useful applications, *e.g.*, a photosynthetic system.¹⁾ These fullerene and porphyrin hybrid compounds are all hydrophobic, and little is known about their water-soluble derivatives and their application.

Here we describe the synthesis of water-soluble C₆₀-porphyrin hybrid compounds in which one *meso*-position is occupied by the *p*-phenylene C₆₀ derivative, while the remaining positions contain the *N*-methylpyridinium-4-yl substituents. We also show a preliminary example of their pharmaceutical application.

Our synthetic pathway is shown in Chart 1. 5-(4-Methoxycarbonylphenyl)-10,15,20-tris(4-pyridyl)porphine **1**²⁾ was obtained according to standard Adler's procedure with a 3% yield. The ester group of **1** was converted to aldehyde **3** in two steps. Then **3** was introduced to the C₆₀ core by 1,3-dipolar cycloaddition with sarcosine to give **4**. The C₆₀-porphyrin hybrid compound **4** was methylated with excess methyl tosylate (30 eq) in chloroform to afford the tosylate salt of **5**, which was converted to its chloride salt by being passed through an anion-exchange resin (Amberlyte IRA 400). Finally, **6**, a Mn complex of **5**, was obtained from the tosylate salt of **5**. Mn was inserted into **5** using excess Mn(OAc)₂ (10 eq), purified as perchlorate salt, and then converted again to the chloride salt of **6** by an anion-exchange resin. The structures of these compounds were confirmed by standard instrumental analyses. **5** and **6** had sufficient water-solubility, as we had anticipated.

Next, we tested the pharmaceutical evaluation of **5** and **6**. We have previously studied fullerene derivatives against active oxygen toxicity and have already shown that anionic water-soluble C₆₀ derivatives have moderate superoxide-quenching activity.³⁾ They actually decreased the active oxygen toxicity in *E. coli*.⁴⁾ Meanwhile, cationic water-soluble C₆₀ derivatives had toxicity in *E. coli*⁵⁾ in spite of their potent activity on superoxide quenching.⁶⁾ Therefore, cationic C₆₀ derivatives that have no toxicity against biological systems are expected for the application of fullerene as an antioxidant. On the other hand, manganese *meso*-tetrakis(*N*-methylpyridinium-4-yl)porphine (Mn-TMPyP) is well known for its potent superoxide-quenching activity,⁷⁾ which is why we became interested in the activity of the hybrid molecules.

We evaluated **5** and **6** for superoxide-quenching activity by the standard cytochrome *c* method.⁸⁾ Their IC₅₀ was 23 μM

for **5** and 0.76 μM for **6**, while their porphyrin control compounds TMPyP and Mn-TMPyP showed similar values (7.2 μM for TMPyP and 0.70 μM for Mn-TMPyP⁷⁾). As we had anticipated, the activity of **5** and **6** was much higher than that of a water-soluble C₆₀ molecule with a negative charge (IC₅₀ *ca.* 100 μM).⁶⁾ Obviously, the difference in C₆₀-porphyrin hybrid compounds and porphyrin control on activity was not regarded as significant.

We expect that these derivatives reduce superoxide toxicity in a more hydrophobic environment, and we are now investigating their effect in *E. coli* systems.

Experimental

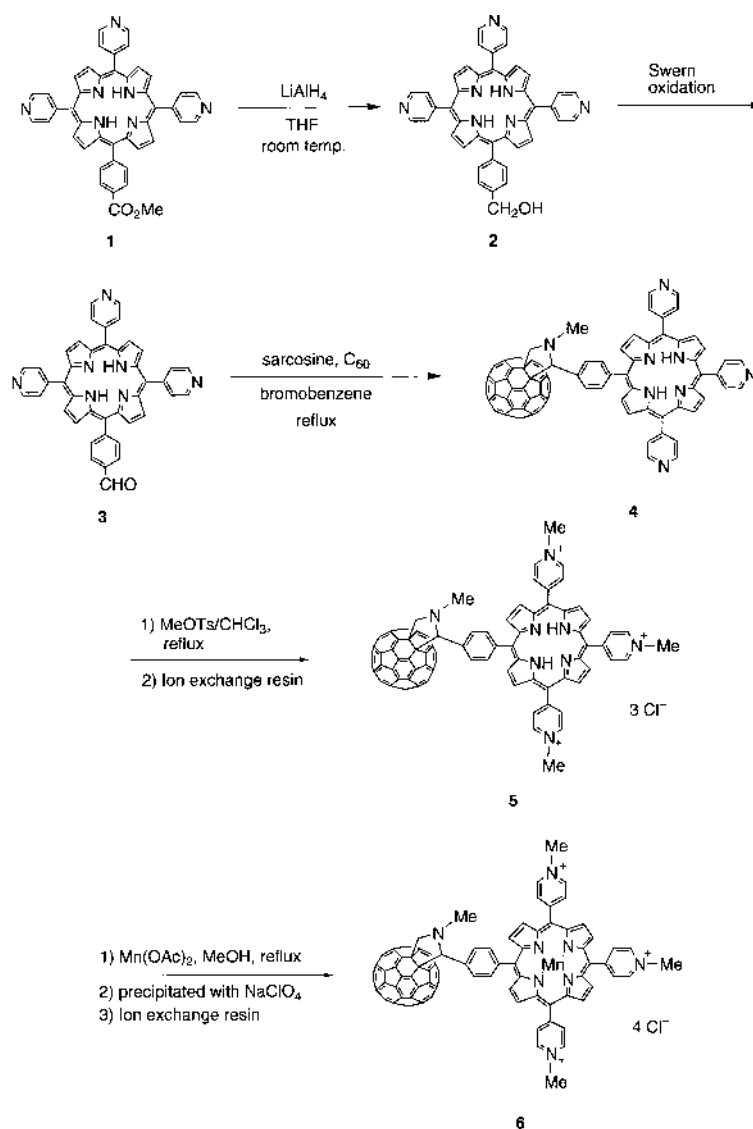
General Methods The ¹H-NMR spectra were recorded on a Varian VXR-200 or 500 spectrometer with tetramethylsilane as an internal standard. IR absorption spectra were recorded on a JASCO FT/IR-200 spectrometer. FAB-MS spectra were measured on a VG-70 spectrometer using *m*-nitrobenzyl alcohol as a matrix. Elemental analyses were performed on a Yanagimoto MT-5 CHN Corder elemental analyzer. Silica gel TLC was performed with Merck Kieselgel F₂₅₄ precoated plates, and the silica gel used for column chromatography was Daisogel.

5-(4-Hydroxyphenyl)-10,15,20-tris(4-pyridyl)porphine (2) To a solution of 5-(4-methoxycarbonylphenyl)-10,15,20-tris(4-pyridyl)porphine (**1**, 710 mg, 1.05 mmol) in anhydrous THF (100 ml) was added LiAlH₄ (319 mg, 8.40 mmol) at room temperature. After being stirred for 30 min, AcOEt (10 ml) was added and then poured into H₂O (100 ml). The aqueous suspension was extracted with CHCl₃ (300 ml), washed with brine, dried over MgSO₄, and evaporated under reduced pressure to yield **2** (360 mg, 53%). **2** was used for the next reaction without further purification. An aliquot was purified by silica gel column chromatography with CHCl₃-EtOH (97:3) for analysis. ¹H-NMR (CDCl₃) δ: -2.88 (2H, s, internal pyrrole), 5.10 (2H, s, -CH₂OH), 7.80 (2H, d, 3,5-phenyl, *J*=7.6 Hz), 8.19 (8H, m, 2,6-phenyl and 3,5-pyridyl), 8.86 (8H, m, pyrrole β), 9.06 (6H, d, 2,6-pyridyl, *J*=4.4 Hz). FAB-MS *m/z*: 648 ([M+H]⁺).

5-(4-Formylphenyl)-10,15,20-tris(4-pyridyl)porphine (3) To a solution of (COCl)₂ (0.5 ml, 5.85 mmol) in anhydrous CH₂Cl₂ (5 ml) was dropped DMSO (0.5 ml, 7.05 mmol) in CH₂Cl₂ for 2 min at -60°. After being stirred for 15 min at the same temperature, **2** (99.4 mg, 0.153 mmol) in CH₂Cl₂ (50 ml) was dropped into the above mixture for 10 min. After being further stirred for 20 min at the same temperature, the reaction was completed by the addition of triethylamine (2.5 ml, 17.9 mmol) and was stirred for 30 min longer at the same temperature. After being warmed to room temperature, the organic layer was washed with water, 0.5 M NaHCO₃, and then brine. After being dried over MgSO₄ and evaporated under reduced pressure, the crude **3** was purified by silica gel column chromatography with CHCl₃-EtOH (97:3) to afford **3** (65.3 mg, 66%). ¹H-NMR (CDCl₃) δ: -2.89 (2H, s, internal pyrrole), 8.18 (6H, d, 3,5-pyridyl, *J*=4.8 Hz), 8.32 (2H, d, 3,5-phenyl, *J*=8.2 Hz), 8.41 (2H, d, 2,6-phenyl, *J*=8.2 Hz), 8.87 (8H, d, pyrrole β, *J*=2.6 Hz), 9.07 (6H, br, 2,6-pyridyl), 10.41 (1H, s, -CHO). FAB-MS *m/z*: 646 ([M+H]⁺). IR (KBr) cm⁻¹: 1700 (C=O).

Trispyridylporphyrin-C₆₀ Hybrid (4) To a solution of C₆₀ (159 mg, 221 μmol) in bromobenzene (120 ml) was added **3** (95.0 mg, 147 μmol) and sarcosine (19.7 mg, 221 μmol). The reaction mixture was refluxed in an Ar atmosphere for 1 h and then evaporated under reduced pressure. The crude

* To whom correspondence should be addressed. e-mail: mashino-td@kyoritsu-ph.ac.jp

Chart 1. Preparation of **5** and **6**

mixture was subjected to silica gel column chromatography. After unreacted C₆₀ was eluted with toluene, the elution with CHCl₃-EtOH (95 : 5) afforded **4** (135 mg, 63%). ¹H-NMR (CDCl₃) δ: -2.93 (2H, s, internal pyrrole), 2.97 (3H, s, N-Me), 4.00 (1H, d, *J*=9.2 Hz), 4.80 (1H, d, *J*=9.2 Hz), 4.81 (1H, s), 8.12 (10H, m, phenyl and 3,5-pyridyl), 8.83 (8H, m, pyrrole β), 9.04 (6H, br, 2,6-pyridyl). FAB-MS *m/z*: 1392 (M⁺), 721 ([C₆₀+H]⁺). UV λ_{max} (CH₂Cl₂) nm (ε): 417 (2.6×10⁵), 513 (1.9×10⁴), 547 (1.1×10⁴), 588 (1.0×10⁴), 645 (7.4×10³).

Tris(4-methylpyridyl)porphyrin-C₆₀ Hybrid (5) To a solution of **4** (124 mg, 89.1 μmol) in bromobenzene (25 ml) was added methyl tosylate (0.4 ml, 2.65 mmol). The reaction mixture was refluxed in an Ar atmosphere for 1 h and then evaporated under reduced pressure. The crude tosylate salt of **5** was passed through an anion-exchange resin (Amberlyte IRA 400) repeatedly to yield **5** as a chloride salt (63.0 mg, 39%). ¹H-NMR (DMSO): -3.06 (2H, s, internal pyrrole), 3.11 (3H, s, N-Me), 4.51 (1H, d, *J*=9.2 Hz), 4.72 (9H, s, quaternary ammonium methyl), 5.24 (1H, d, *J*=8.6 Hz), 5.53 (1H, s), 8.32 (8H, m, pyrrole β), 8.96 (6H, d, 3,5-pyridyl, *J*=6.7 Hz), 9.12 (4H, m, phenyl), 9.49 (6H, d, 2,6-pyridyl, *J*=6.7 Hz). UV λ_{max} (H₂O) nm (ε): 426 (1.3×10⁵), 524 (1.2×10⁴), 560 (shoulder), 590 (shoulder), 652 (2.4×10³). *Anal.* Calcd for C₁₀₇H₄₁N₈Cl₃·16H₂O: C, 70.11; H, 4.01; N, 6.11. Found: C, 70.25; H, 3.85; N, 6.32.

Mn-Tris(4-methylpyridyl)porphyrin-C₆₀ Hybrid (6) To a solution of **5** (tosylate salt, 36.7 mg, 188 μmol) in methanol (20 ml) was added manganese (II) acetate tetrahydrate (46.2 mg, 0.189 mmol). The completion of the reaction was determined by a shift in the Soret band. After being refluxed for

1 h, the reaction mixture was evaporated under reduced pressure. The crude product was dissolved in distilled water (10 ml), and then the precipitate was obtained by the addition of sodium perchlorate (42.6 mg, 0.303 mmol). The precipitate was collected by centrifugation and then washed 5 times with 1.2% perchloric acid and 1 time with distilled water to yield **6** as a perchlorate salt. This material was converted by being passed through an anion-exchange resin (Amberlyte IRA 400) to yield **6** as a chloride salt (31.4 mg, 84%). UV λ_{max} (H₂O) nm (ε): 464 (1.1×10⁵), 562 (1.2×10⁴). *Anal.* Calcd for C₁₀₇H₃₉N₈Cl₄Mn·19H₂O: C, 65.05; H, 3.93; N, 5.67. Found: C, 65.32; H, 3.64; N, 5.70.

Measurement of Superoxide-Quenching Activity This assay was performed according to the xanthine-xanthine oxidase/cytochrome c method.⁸⁾ Conditions were as follows: 0.05 M potassium phosphate buffer, 0.10 mM EDTA, pH 7.8, 50 μM xanthine, 10 μM cytochrome c, 700 units/ml catalase. The reproducibility of IC₅₀ was within 10%. We checked that neither sample inhibited xanthine oxidase activity.

Acknowledgments This research was financially supported by Grants-in-Aid for the Encouragement of Young Scientists (No. 12771435 to K.O.) from the Japan Society for the Promotion of Science, the Okayama Foundation for Science and Technology (to K.O.), and the Hoh-ansha Foundation (to T.M.). We are grateful to The SC-NMR Laboratory of Okayama University for NMR experiments.

References and Notes

- 1) For a review, see Imahori H., Sakata Y., *Eur. J. Org. Chem.*, **1999**, 2445—2457 (1999).
- 2) Ishikawa Y., Yamashita A., Uno T., *Chem. Pharm. Bull.*, **49**, 287—293 (2001).
- 3) Okuda K., Mashino T., Hirobe M., *Bioorg. Med. Chem. Lett.*, **6**, 539—542 (1996).
- 4) Okuda K., Hirobe M., Mochizuki M., Mashino T., *Fullerenes: Recent Advances in the Chemistry and Physics of Fullerenes and Related Materials*, **5**, 337—344 (1997).
- 5) Mashino T., Okuda K., Hirota T., Hirobe M., Nagano T., Mochizuki M., *Bioorg. Med. Chem. Lett.*, **9**, 2959—2962 (1999).
- 6) Okuda K., Hirota T., Hirobe M., Nagano T., Mochizuki M., Mashino T., *Fullerene Sci. Technol.*, **8**, 89—104 (2000).
- 7) Faulkner K. M., Liochev S. I., Fridovich I., *J. Biol. Chem.*, **269**, 23471—23476 (1994).
- 8) McCord J. M., Fridovich I., *J. Biol. Chem.*, **244**, 6049—6055 (1969).