# **Biomimetic Oxidation of 2-Methylimidazole Derivative with a Chemical Model System for Cytochrome P-450**

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**A chemical model system for cytochrome P-450, consisting of tetraphenylporphyrin manganese chloride (TPPMnCl) and iodosylbenzene, efficiently oxidized 2-methylimidazole to 2-methylimidazolone. This system was next applied to 4-(2-methyl-1-imidazolyl)-2,2-diphenylbutyramide, a muscarinic acetylcholine receptor antagonist under clinical trial, affording the previously unisolated imidazole ring 5-mono-oxidized derivative that is considered to be the precursor of the main metabolites. This system, which is superior to the copper-ascorbate system, should be applicable to** *in vitro* **studies of various drugs containing the 2-methylimidazole moiety.**

**Key words** 2-methylimidazole derivative; 2-methylimidazolone derivative; metabolic intermediate; drug metabolism; P-450 chemical model; metalloporphyrin

Cytochrome P-450 (designated as P-450) is a family of heme-containing metallo-enzymes named after the absorption band at 450 nm of their carbon monoxide form. P-450 metabolizes various kinds of xenobiotics, including drugs, oxidatively or reductively, playing an important role in drug metabolism.<sup>1)</sup> Many chemical models for P-450 have been developed to elucidate and/or mimic the function of the enzymes. Several models function as efficient oxidation systems for simple substrates, but there have been only a few reports on the application of these models to the *in vitro* metabolic study of heterocycles, which are often used as an important structural backbone for drugs and candidate drugs. For example, in the case of the oxidation of 3-isobutyryl-2 isopropylpyrazolo[1,5-a]pyridine (Ibudilast; IBPP), which has been developed as an antiasthma drug and cerebral vasodilator, ring hydroxylation of the pyrazolo<sup>[1,5-a]pyridine</sup> nucleus occurred smoothly to give the monoepoxide derivative along with the diepoxide derivative.<sup>2)</sup> Those compounds are unstable precursors of the final metabolites of IBPP.

2-Methylimidazole is an important pharmacophore, *e.g.*, in thromboxane  $A_2$  synthase inhibitors, aromatase inhibitors, 5-hydroxytryptamine (5HT) inhibitors, muscarinic acetylcholine receptor antagonists, and so on.<sup>3)</sup> In general, the imidazole ring is rather metabolically labile, affording ring polyoxygenated and related metabolites.<sup>4)</sup> However, there has been no report on the isolation and structural determination of imidazole ring mono-oxygenated metabolites (designated as imidazolones), which are thought to be precursors of the final metabolites of various drugs and candidate drugs.

It has been reported for several imidazole-containing drugs that trace amounts of drug equivalents are retained in connective tissue after administration to laboratory animals.<sup>5—7)</sup> Although the mechanism(s) of the retention in connective tissue is not yet known, it may involve the reaction of a reactive intermediate (such as imidazolone), formed by metabolism of the imidazole moiety, with tissue macromolecules.<sup>8)</sup> Therefore, it is important to develop an efficient method for the preparation of imidazolone derivatives under biomimetic reaction conditions. Direct transformation of 2 methylimidazole to 2-methylimidazolone using a cuproascorbate system (cupric chloride and L-ascorbic acid) was reported, but the yield was very poor.<sup>9)</sup> We retested this system, but found that reproducibility was poor (data not shown).

In this paper, we present a novel mono-oxidation of 2 methylimidazole by using metalloporphyrin–iodosylbenzene as a chemical model system for cytochrome P-450.<sup>10—12)</sup> Application of this system to a 2-methylimidazole-containing candidate drug is also described.

#### **Experimental**

**General** Melting points were determined on a Yanagimoto micro melting point apparatus and are uncorrected. <sup>1</sup>H-NMR spectra were measured in  $CDCl<sub>3</sub>$  with tetramethylsilane (TMS) and the solvent peak as internal standards, on a JEOL JMN-A400 spectrometer. Mass spectra (MS) were obtained on a JEOL JMS-HX110 spectrometer. Column chromatography was carried out on Merck silica gel 60. Analytical thin-layer chromatography (TLC) was performed on Merck precoated silica gel  $60F_{254}$  plates, and the compounds were visualized by UV illumination (254 nm) or by heating after spraying with phosphomolybdic acid in ethanol. HPLC analysis was performed on a TOSOH ODS-80T<sub>M</sub> column (4.6×250 mm). Chromatographic conditions were as follows: samples were eluted at a flow-rate of 0.5 ml/min with 0.05 mol/l ammonium acetate–methanol solution  $(25:1 \text{ v/v})$ . The samples were monitored by measuring the absorbance at 220 nm. The areas of the chromatographic peaks were calculated by use of a Hitachi chromato-integrator (D-2000). Elemental analysis was performed in the microanalytical laboratory of Kyorin Pharmaceutical Co., Ltd.

**Chemicals** 2-Methylimidazole, tetraphenylporphyrin manganese chloride (TPPMnCl), tetraphenylporphyrin iron chloride (TPPFeCl), manganese chloride (MnCl<sub>2</sub>), iodosylbenzene (PhIO), dichloromethane, and acetonitrile were of reagent grade and were used without further purification. 4-(2- Methyl-1-imidazolyl)-2,2-diphenylbutyramide<sup>13—16)</sup> (KRP-197) and its tetradeuterio derivative  $(D_4 - KRP - 197)$  were synthesized by the Discovery Research Laboratory of Kyorin Pharmaceutical Co. Ltd.



Chart 1. Reaction of 2-Methylimidazole with the TPPMnCl–PhIO System



Chart 2. Reaction of 2-Methylimidazolone with Benzaldehyde

**Reaction with the Chemical Model** In accordance with the method reported by Cook et al.,<sup>17)</sup> 1 equivalent of substrate, 0.01—0.05 molar equivalent(s) of metalloporphyrin and  $1-3$  molar equivalent(s) of oxidant were dissolved in 10 ml of mixed solvent of dichloromethane and acetonitrile  $(1:1 \text{ v/v})$ , and the mixture was stirred at room temperature (see Chart 1). The reaction was monitored by HPLC and/or TLC.

### **Results and Discussion**

Figure 1 shows HPLC chromatograms indicating the formation of 2-methylimidazolone from 2-methylimidazole in the TPPMnCl–PhIO system. When an equimolar amount of PhIO was used as the oxidant, the reaction was incomplete, and a considerable amount of the starting material remained, even after 5 h. However, the reaction went to completion when 2 molar-equivalents of PhIO was used, and 2 methylimidazolone was obtained almost quantitatively. Without metalloporphyrin manganese chloride, little reaction took place. Furthermore, little reaction took place when  $MnCl<sub>2</sub>$ was used instead of TPPMnCl. These results clearly indicated that 2-methylimidazole was oxidized not by PhIO itself, but by an oxidant formed by the reaction between TPPMnCl and PhIO, presumably a manganese-oxenoid species.<sup>18)</sup> The effect of the central metal is small, because similar efficiency was obtained by the use of TPPFeCl as a catalyst. The structural determination of 2-methylimidazolone was done by  ${}^{1}$ H-NMR, and high-resolution mass spectrometry.<sup>19)</sup> Furthermore, the structure was confirmed by derivatization of 2-methylimidazolone to the aldol adduct with benzaldehyde.

2-Methylimidazolone reacted smoothly with benzaldehyde at room temperature in the absence of base to afford the (*Z*)-  $\alpha, \beta$ -unsaturated ketone derivative<sup>20)</sup> in good yield, indicating that the acidity of the methylene moiety of 2-methylimidazolone is high.

Based on these observations, we applied this biomimetic oxidation reaction system to 4-(2-methyl-1-imidazolyl)-2,2 diphenyl butyramide<sup>13—16)</sup> (KRP-197; Figure 2), which was developed as a dual antagonist of muscarinic acetylcholine receptors  $M_1$  and  $M_3$ . KRP-197 is a candidate drug for the treatment of urinary incontinence associated with bladder muscle instability, and is currently under clinical study. The imidazole ring mono-oxidation product of KRP-197 (designated as KRP-197-imidazolone) is thought to be an unstable precursor of the final metabolites of KRP-197, but has not yet been isolated.<sup>21)</sup> To make structural analysis easier, a mixture of KRP-197 and its tetradeuterio analog (designated as  $D_4$ -KRP-197 (Chart 3)) was used as a substrate (designated as  $(H_4, D_4$ -KRP-197)).

When a mixture of  $(H_4, D_4)$ -KRP-197, 0.01 molar-equivalent of TPPMnCl, and 1.5 molar-equivalent of PhIO in a mixed solvent of dichloromethane–acetonitrile  $(1:1 \text{ v/v}; 10)$ ml) was stirred for 6 h at room temperature, a product which showed molecular-ion peaks at 336  $(M+H)^+$  and 340  $(M+H)^+$  was isolated in about 30% yield, together with considerable recovery of the starting material (Chart 4).<sup>22)</sup> In the mass spectrum (see Fig. 3),  $(M+H)^+=336$  was 16 *m/z* larger than KRP-197, and  $(M+H)^+=340$  was 16  $m/z$  larger than  $D_4$ -KRP-197. These mass unit gains correspond to one oxygen atom. These results indicated that the product is a monooxygenated derivative of  $(H_4, D_4)$ -KRP-197. The mass fragments of 238  $(M+H)^+$  and 242  $(M+H)^+$  corresponded to the  $(C_6H_5)$ ,  $C(CONH_2)CX$ ,  $CX=H$ , or D) moiety, and no



Fig. 1. HPLC Chromatograms of the Reaction Products of 2-Methylimidazole with the TPPMnCl-PhIO System

(A) 2-methylimidazole. (B) Synthetic 2-methylimidazolone.<sup>29)</sup> (C) After 5 h reaction (1 eq of PhIO was used). (D) After 4 h reaction (2 eq of PhIO was used).  $\Box$  and  $\blacksquare$  indicate 2-methylimidazole and 2-methylimidazolone respectively.



Fig. 2. Chemical Structure of KRP-197



a) BrCD<sub>2</sub>CD<sub>2</sub>Br, DB-18-crown-6, 50% NaOH; b) 2-methylimidazole, TEA, DMF; c) KOH, 2-PrOH.

Chart 3. Synthesis of  $D_4$ -KRP-197



Chart 4. Oxydation of  $(H_4, D_4)$ -KRP-197 with the P-450 Chemical Model System

fragment with a mass 16 units higher was seen in the spectrum. These results indicated that mono-oxygenation had occurred not on the phenyl ring, but on the imidazole ring of KRP-197. The oxygenated position of the imidazole nucleus was determined by heteronuclear multiple bond connectivity (HMBC) analysis to be the 5-position (see Chart 4). Furthermore, the 5-imidazolone structure was confirmed by comparison of the NMR spectra of the isolated product and an authentic sample, which was prepared from 2,2-diphenylacetonitrile.<sup>23)</sup> These compounds exhibited identical  $H$ - and



Fig. 3. Mass Spectrum of the Product of the Reaction of  $(H_4, D_4)$ -KRP-197 with the TPPMnCl-PhIO System

 ${}^{13}$ C-NMR spectra.

In this paper, we have shown that 2-methylimidazole and a derivative of it were oxidized effectively with a chemical model system for cytochrome P-450, TPPMnCl–PhIO, to afford the 2-methylimidazolone mono-oxidation product. The reaction system is superior to the previously described cuproascorbate system in respect of reproducibility and yield of the product. The formation of 2-methylimidazolone from 2 methylimidazole was reported to occur through a cytochrome P450-independent pathway,<sup>24)</sup> but this suggestion was based on very limited findings. Considering our present results and a report showing that simple imidazole was oxidized through a P-450-dependent pathway to afford hydantoin and hydantoic acid, $24$ ) we consider that the metabolic pathway to 2methylimidazolone from 2-methylimidazole is at least partly P-450-dependent. Further study is under way.

Interestingly, various imidazoles, such as imidazole, 2 methylimidazole, *N*-methylimidazole and so on, have been used as ligands for metalloporphyrins in order to enhance the reactivity of the catalyst and/or to increase the yield of the product. $25-27$  However, there is little information about the fate of the added imidazoles during the oxidation reaction, although they were used in excess molar amounts as compared to the metalloporphyrin catalyst.<sup>28)</sup> Our present results indicate that in certain cases, not only the substrate itself, but also the added imidazole might be oxidized, affording both the desired product and the oxidized imidazole.

## **Conclusion**

We showed that a chemical model system for cytochrome P-450 could oxidize 2-methylimidazole to 2-methylimidazolone. A 2-methylimidazole-containing drug was similarly mono-oxidized to afford a putative precursor of the main metabolites. The efficiency of the reaction was superior to that of a reported method using a copper-ascorbate system. Further application of this chemical model system for *in vitro* metabolic studies of various drugs and/or drug candidates containing a heterocyclic structure is in progress.

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- 19) <sup>1</sup> <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.99 (3H, s), 3.90 (2H, s), 10.76 (1H, br s). FAB-MS  $m/z$ : 98.0505 (Calcd for C<sub>4</sub>H<sub>6</sub>N<sub>2</sub>O: 98.0480).
- 20) (*Z*)-5-Benzylidene-2-methyl-3,5-dihydro-imidazol-4-one; mp. 171.0— 171.5 °C. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  2.39 (3H, s), 7.08 (1H, s), 7.37—7.46 (3H, m), 8.13 (2H, dd, J=8.3, 1.7 Hz), 8.82 (1H, br s). FAB-MS  $m/z$ : 186.0772 (Calcd for C<sub>11</sub>H<sub>10</sub>N<sub>2</sub>O: 186.0793). Anal. Calcd for  $C_{11}H_{10}N_2O$ : C, 70.95; H, 5.41; N, 15.04. Found: C, 70.89; H, 5.38; N, 14.99.
- 21) The metabolic study of KRP-197 will be published elsewhere.
- 22) <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  2.14 (3H, t, J=2.0 Hz), 2.61—2.65 (2H, m), 3.44—3.47 (2H, m), 3.99 (2H, d,  $J=2.0$  Hz), 5.51 (1H, br s), 5.57 (1H, br s), 7.30—7.40 (10H, m). <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 15.71, 37.77, 38.41, 58.23, 58.92, 128.44—128.80, 142.26, 163.57, 176.24, 181.42. FAB-MS  $m/z$ : 336.1728 (Calcd for C<sub>20</sub>H<sub>22</sub>N<sub>3</sub>O<sub>2</sub>: 336.1712), 340.1977 (Calcd for C<sub>20</sub>H<sub>18</sub>D<sub>4</sub>N<sub>3</sub>O<sub>2</sub>: 340.1963).
- 23) Synthesis of authentic KRP-197-5-imidazolone will be published elsewhere.
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