

Fern Constituents: Triterpenoids from *Adiantum capillus-veneris*

Takahisa NAKANE,^a Yoshiko MAEDA,^a Hideharu EBIHARA,^a Yoko ARAI,^a Kazuo MASUDA,^a Akihito TAKANO,^a Hiroyuki AGETA,^a Kenji SHIOJIMA,^{*,a} Shao-Qing CAI,^b and Osama Bashir ABDEL-HALIM^c

^aShowa Pharmaceutical University; Machida, Tokyo 194–8543, Japan; ^bDivision of Pharmacognosy, School of Pharmaceutical Sciences, Peking University; 38 Xue-yuan Road, Haidian District, Beijing 100083, China; and ^cMansoura University; 35511 Mansoura, Egypt. Received April 18, 2002; accepted May 31, 2002

Two new migrated hopane triterpenoids, viz. 4 α -hydroxyfilican-3-one and fern-9(11)-en-12 β -ol, and olean-18-en-3-one and olean-12-en-3-one as the first example of oleanane compounds from *Adiantum* ferns were isolated along with many other known triterpenoids from *Adiantum capillus-veneris* of China and Egypt. Their structures were elucidated by spectroscopic analyses.

Key words *Adiantum capillus-veneris*; triterpenoid; Adiantaceae; 4 α -hydroxyfilican-3-one; fern-9(11)-en-12 β -ol

In preceding papers of this series,^{1,2)} we reported the isolation of twenty-two triterpenoids, including nine new compounds from the fresh fronds of *Adiantum capillus-veneris* L. (Adiantaceae) collected in Japan. Further investigation of *A. capillus-veneris* L. collected in China and Egypt has resulted in the isolation of two new triterpenoids; viz. 4 α -hydroxyfilican-3-one (**1**) and fern-9(11)-en-12 β -ol (**2**) of Chinese origin, and two oleanane triterpenoids; olean-12-en-3-one (**3**) and olean-18-en-3-one (**4**) of Egyptian origin (Chart 1). In fact, **3** and **4** are the first examples of oleanane compounds from *Adiantum* ferns. This paper deals with the isolation and structure elucidation of these new compounds.

The constituents of the crude hexane extract of these fronds were purified by various chromatographic techniques (see Experimental) to give nineteen triterpenoids, seventeen of which were known: olean-18-en-3-one (**3**),³⁾ olean-12-en-3-one (**4**),³⁾ fern-9(11)-ene (**5**),⁴⁾ ferna-7,9(11)-diene (**6**),⁴⁾ fern-7-ene (**7**),⁴⁾ hop-22(29)-ene (**8**),⁴⁾ filic-3-ene (**9**),⁴⁾ neo-hop-12-ene (**10**),⁴⁾ adiantoxide (**11**),¹⁾ adiantone (**12**),⁵⁾ fern-9(11)-en-12-one (**13**),⁵⁾ 28-hydroxyfern-9(11)-ene (**14**),¹⁾ isoadiantone (**15**),⁶⁾ isoglaucanone (**16**),⁵⁾ hydroxyhopane (**17**),⁵⁾ isoadiantol (**18**)⁵⁾ and hydroxyadiantone (**19**).⁵⁾

Compound **1** was obtained as colorless needles, and its high resolution HR-MS showed M⁺ at *m/z* 442.3804 suggesting the molecular formula to be C₃₀H₅₀O₂ (Calcd, 442.3810). Its IR spectrum indicated the presence of a hydroxyl and a carbonyl group. The ¹H-NMR spectrum of **1** displayed signals for six tertiary and two secondary methyl groups, and the chemical shifts of methyl protons (H-25–H-30) resembled those of filican-3-one (**20**) (Table 1). The ¹³C chemical shifts of **1** were also very close to those of **20** (Table 2), except for those of C-2, C-4, C-5, C-6 and C-24. The large down field shifts of C-4 by 22.8 ppm suggested that the hydroxyl group of **1** was located at C-4.

The heteronuclear multiple bond correlation (HMBC) spectrum of **1** also fully corroborated the above observation (Fig. 1). MS of **1** showed distinctive peaks at *m/z* 442 (M)⁺ (100), 427 (M–15)⁺ (10), 424 (M–H₂O)⁺ (9), 409 (M–CH₃–H₂O)⁺ (5), 399 (M–C₃H₇)⁺ (30), 381 (M–C₃H₇–H₂O)⁺ (12), 273 (a) (53), 205 (b) (25), and 191 (c) (27) (Chart 2) which also supported the assigned structure. The orientation of the hydroxyl group was deduced from the nuclear Overhauser enhancement spectroscopy (NOESY) spectrum of **1**, which exhibited NOE correlations between the H₃-23 and β -oriented H₃-24, H₃-24 and β -oriented H₃-25, H₃-25 and H₃-26, and H₃-27 and α -oriented H₃-28 (Fig. 2), and therefore the OH group at C-4 must be α -oriented. Thus, the structure of **1** was established as 4 α -hydroxyfilican-3-one.

Compound **2** was obtained as a colorless needles, and its IR spectrum suggested the presence of hydroxyl group in the molecule. Its molecular formula was deduced to be C₃₀H₅₀O by HR-MS (M⁺ at *m/z* 426.3878; Calcd, 426.3861). The molecular ion peak at *m/z* 426 (M)⁺ and the other diagnostic peaks⁶⁾ at *m/z* 411 (M–CH₃)⁺ (35), 393 (M–CH₃–H₂O)⁺ (22), 273 (d) (81), 255 (d–H₂O)⁺ (44) and 134 (e) (79) (Chart 2) in its MS indicated that **2** is a fernene derivative with a hydroxyl group at ring A, B or C of the molecule. The ¹H-NMR spectrum of **2** indicated the presence of six tertiary methyl groups, two secondary methyl groups, one trisubstituted vinylic methine proton and a hydroxyl methine proton (Table 1). The ¹³C-NMR chemical shifts of **2** were similar to those of fern-9(11)-ene (**5**)⁶⁾ except for the signals of C-11, C-12, C-13 and C-18 (Table 2). The large down-field shifts of carbon signals at C-12 by 36.4 ppm, and the up-field shift at C-18 signal by ca. 5.3 ppm suggested that the hydroxyl group of **2** was located at C-12. The HMBC data also fully corroborated the above observation.

Compound **3** was obtained as colorless needles, and its IR spectrum indicated the presence of a carbonyl group. The ¹H-NMR spectrum of **3** displayed signals for six tertiary and two secondary methyl groups, and the chemical shifts of methyl protons (H-25–H-30) resembled those of filican-3-one (**20**) (Table 1). The ¹³C chemical shifts of **3** were also very close to those of **20** (Table 2), except for those of C-2, C-4, C-5, C-6 and C-24. The large down field shifts of C-4 by 22.8 ppm suggested that the hydroxyl group of **3** was located at C-4.

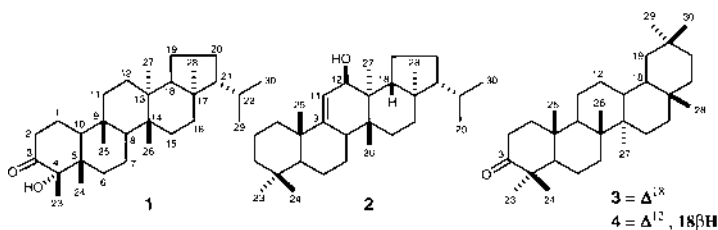


Chart 1

* To whom correspondence should be addressed. e-mail: y-arai@ac.shoyaku.ac.jp

Table 1. $^1\text{H-NMR}$ Spectral Data (500 MHz, CDCl_3 , δ) for **1**–**5** and **20**

^1H	1	20	2	5	3	4
23	1.154	0.900 (d, 6.7)	0.848	0.847	1.081	1.097
24	0.792	0.711	0.899	0.888	1.035	1.057
25	0.907	0.902	1.095	1.053	0.964	1.073
26	0.907	0.913	0.942	0.733	1.107	1.023
27	1.000	0.984	0.812	0.822	0.749	0.842
28	0.796	0.796	0.792	0.759	1.028	0.842
29	0.889 (d, 6.7)	0.890 (d, 6.4)	0.907 (d, 6.4)	0.890 (d, 6.4)	0.943	0.874
30	0.829 (d, 6.7)	0.829 (d, 6.4)	0.842 (d, 6.4)	0.830 (d, 6.4)	0.943	0.874
2 α	2.970 (ddd, 2.1, 4.9, 13.4)					
10 α	2.167 (dd, 3.4, 12.8)					
11			5.511 (dd, 2.4, 4.8)	5.286 (ddd, 2.4, 2.4, 5.1)		
12			3.476 (bs)			5.208 (dd 3.4, 3.7)
19					4.864	

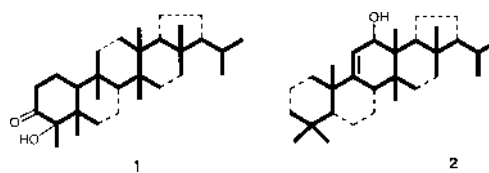
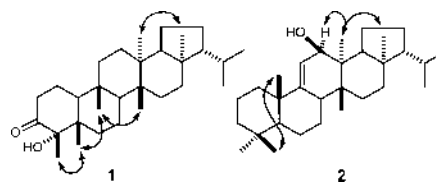
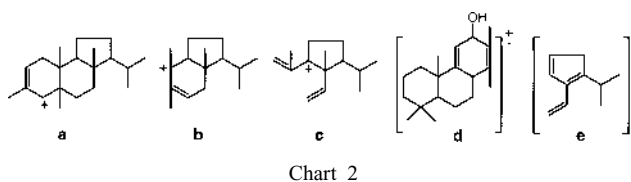
Multiplicity and coupling constant (J , Hz) are shown in parentheses.

Table 2. $^{13}\text{C-NMR}$ Spectral Data^{a)} (125 MHz, CDCl_3 , δ) of **1**–**5** and **20**

	1	20	2	5	3	4
1	21.72	22.32	41.31	41.49	39.86	39.31
2	37.06	41.55	19.43	19.56	34.08	34.22
3	213.10	213.30	42.27	42.43	218.39	217.93
4	81.01	58.22	33.74	33.64	47.27	47.49
5	44.57	42.16	44.53	44.88	54.87	55.32
6	33.18	41.04	19.37	19.53	19.69	19.66
7	17.80	18.25	17.85	17.90	33.81	32.18
8	48.78	49.48	40.23	39.98	40.63	39.80
9	37.71	37.94	154.57	151.68	50.51	46.88
10	59.50	59.69	38.09	38.05	36.93	36.69
11	35.62	35.53	117.68	115.60	21.70	23.66
12	28.35	28.34	73.17	36.78	26.22	121.51
13	39.03	39.03	40.74	36.74	38.52	145.30
14	40.09	40.13	37.10	37.69	43.36	41.87
15	29.14	29.13	31.04	29.28	27.49	26.13
16	35.71	35.60	36.03	36.19	37.36	26.93
17	42.72	42.71	42.23	42.97	34.35	32.53
18	51.71	51.69	46.73	52.02	142.57	47.32
19	19.91	19.91	20.48	20.15	129.88	46.79
20	28.41	28.34	28.27	28.23	32.37	31.10
21	60.10	60.07	59.47	59.68	33.32	34.73
22	30.78	30.78	30.75	30.80	37.64	37.11
23	16.91	16.81	32.69	32.80	26.91	26.48
24	16.39	14.60	21.63	21.68	20.93	21.51
25	20.19	20.14	24.36	25.06	16.59	15.23
26	16.18	16.22	17.81	15.84	15.94	16.74
27	15.74	15.67	16.37	15.43	14.49	25.89
28	16.63	16.39	14.22	14.00	25.28	28.43
29	21.95	21.94	22.16	22.14	31.33	33.33
30	22.90	22.90	23.00	23.02	29.18	23.68

a) Assignments were made on the basis of DEPT, $^1\text{H-}^1\text{H}$ COSY, $^1\text{H-}^{13}\text{C}$ COSY and HMBC spectra.

rated the above observation (Fig. 1). The relative stereochemistry at most of the chiral centers of **2** as well as the orientation of the OH group was deduced from its NOESY spectrum which showed NOE interactions as depicted in Fig. 2. The observed interaction of H-12 (δ 3.476) with H₃-27 indicated that the OH group was β oriented. Finally the structure of **2** was confirmed by its preparation from fern-9(11)-en-12-

Fig. 1. Partial Structures of **1** and **2** Derived from HMBC DataFig. 2. NOE Interactions Observed in the NOESY Spectra of Compounds **1** and **2**

one (**21**) with LiAlH_4 . The reaction yielded two products viz. fern-9(11)-en-12 α -ol (less polar) (**22**) and fern-9(11)-en-12 β -ol (more polar) (**2'**), of which IR and $^1\text{H-NMR}$ spectra proved to be identical to those of **2**. Thus, the structure of **2** was established as fern-9(11)-en-12 β -ol.

It was noteworthy that two oleanane compounds **3** and **4** were obtained from the fronds of *Adiantum capillus-veneris* (Egyptian origin). Similarity of the *Adiantum* ferns between Japanese^{1,2)} and Egyptian origin was suggested by detection of many hopane and migrated hopane triterpenoids from both samples.

Experimental

General mp.s: uncorr.; EI-MS: 30 eV; TLC: on precoated Kiesel gel 60. ^1H - (500 MHz) and ^{13}C -NMR (125 MHz) spectra: in CDCl_3 (TMS as int. standard); HPLC: reverse phase C_{18} column ($8\phi \times 250$ mm), RI detector, $\text{CH}_3\text{CN}-\text{CHCl}_3$ (9:1); CC: silica gel 60 (230–400 mesh, Merck) and 20% AgNO_3 -impregnated silica gel (Mallincrodt); GC: 1.4% SE-30 on Chromosorb G HP, Oven: 260 °C. (cholestane as int. standard).

The fronds of *Adiantum capillus-veneris* were collected in Hunan, China and Mansoura, Egypt. Voucher specimens have been deposited in the herbarium of Showa Pharmaceutical University, Tokyo.

Extraction and Separation (Chinese Material) The dried fronds (50 g) were extracted with hexane three times to give extracts (1.0 g). The extract was refluxed with benzene for 1 h and kept for 1 d at room temperature. The insoluble materials were filtered off (fraction A: fr. A) and the filtrate was evaporated to dryness to afford a gummy residue which was chromatographed over silica gel to give twelve fractions: fr. B (eluted with hexane), fr. C [hexane–benzene (8:2)], fr. D, E, F, G [hexane–benzene (7:3)], fr. H [hexane–benzene (1:1)], fr. I, J (benzene), fr. K [benzene–ether (9:1)], fr. L (ether) and fr. M (methanol). Each eluate was further subjected to silica gel CC and HPLC repeatedly and furnished **1**, **2** and the known compounds, viz. **5** (mp 170–171 °C, 13 mg),⁴⁾ **6** (mp 200–202 °C, 0.5 mg),⁴⁾ **7** (mp 213–214 °C, 3 mg),⁴⁾ **8** (mp 211–212 °C, 1 mg),⁴⁾ **9** (mp 232–234 °C, 0.5 mg),⁴⁾ **10** (mp 210–212 °C, 0.5 mg),⁴⁾ **11** (mp 227–229 °C, 47 mg),¹⁾ **12** (mp 229–231 °C, 90 mg),⁵⁾ **13** (mp 229–231 °C, 1 mg),⁵⁾ **14** (mp 159–161 °C, 4 mg),¹⁾ **15** (mp 236–237 °C, 7 mg),⁵⁾ **16** (mp 243–245 °C, 4 mg),⁵⁾ **17** (mp 253–255 °C, 1 mg),⁴⁾ **18** (mp 213–215 °C, 20 mg)⁵⁾ and **19** (trace).⁵⁾

Extraction and Separation (Egyptian Material) The dried fronds (114 g) were extracted with petroleum ether three times to give the extracts (3.0 g). The extract was refluxed with benzene for 1 h and kept for 1 d. The insoluble materials were filtered off (fraction A: fr. A) and the filtrate was evaporated to dryness to afford a gummy residue which was chromatographed over silica gel to give seven fractions: fr. B (eluted with *n* hexane), fr. C [*n* hexane–benzene (8:2)], fr. D [*n* hexane–benzene (7:3)], fr. E [*n* hexane–benzene (1:1)], fr. F [*n* hexane–benzene (1:1)], fr. G (benzene) and fr. H [benzene– Et_2O (9:1)]. Each eluate was further subjected to silica gel CC and HPLC repeatedly and furnished **3**, **4** and the known compounds, viz. **11** (mp 229–230 °C, 158 mg),¹⁾ **12** (mp 227–230 °C, 145 mg),⁵⁾ **15** (mp 236–237 °C, 19 mg),⁵⁾ **16** (mp 243–245 °C, 8 mg)⁵⁾ and **18** (mp 213–215 °C, 30 mg).⁵⁾

4 α -Hydroxyfilican-3-one (1): Fraction K was chromatographed over silica gel with benzene followed by crystallization from $\text{MeOH}-\text{CHCl}_3$ to give **1** (3 mg). mp 263–265 °C. $[\alpha]_{\text{D}} -4.8^\circ$ ($c=0.2$, CHCl_3). $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} , 3505 (OH) and 1719 (C=O). EI-MS m/z (rel. int. %): 442 (M^+) (100), 427

($\text{M}-\text{CH}_3$)⁺ (10), 424 ($\text{M}-\text{H}_2\text{O}$)⁺ (9), 409 ($\text{M}-\text{H}_2\text{O}-\text{COCH}_3$)⁺ (5), 399 (30), 381 (12). HR-MS; M^+ m/z : 442.3804 (Calcd for $\text{C}_{30}\text{H}_{50}\text{O}_2$: 442.3801). ^1H - and ^{13}C -NMR spectral analyses: Tables 1 and 2.

Fern-9(11)-12 β -ol (2): Fraction F was chromatographed over silica gel with *n* hexane–benzene (7:3) and then each of the fractions was subjected to preparative HPLC followed by crystallization to give **2** (3 mg). mp 191–192 °C. $[\alpha]_{\text{D}} -47.4^\circ$ ($c=0.2$, CHCl_3). $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} , 3450 (OH) and 1018 (C–O). EI-MS m/z (rel. int. %): 426 (M^+) (15), 441 ($\text{M}-\text{CH}_3$)⁺ (35), 393 ($\text{M}-\text{H}_2\text{O}-\text{CH}_3$)⁺ (22), 273 (81), 255 (44), 134 (79). HR-MS; M^+ m/z : 426.3878 (Calcd for $\text{C}_{30}\text{H}_{50}\text{O}_2$: 426.3861). ^1H - and ^{13}C -NMR spectral analyses: Tables 1 and 2.

Olean-18-en-3-one (3): mp 183–185.5 °C. $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} , 1700 (C=O). EI-MS m/z (rel. int. %): 424 (M^+) (46), 409 ($\text{M}-\text{CH}_3$)⁺ (68), 218 (11), 205 (59), 204 (73), 177 (100). ^1H -NMR δ : 1.081 (3H, s, H-23), 1.035 (3H, s, H-24), 0.964 (3H, s, H-25), 1.107 (3H, s, H-26), 0.749 (3H, s, H-27), 1.028 (3H, s, H-28), 0.943 (3H, s, H-29), 0.943 (3H, s, H-30), 4.864 (1H, s, H-19).³⁾

Olean-12-en-3-one (4): mp 166–167 °C. $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} , 1700 (C=O). EI-MS m/z (rel. int. %): 424 (M^+) (26), 409 ($\text{M}-\text{CH}_3$)⁺ (19), 218 (100), 205 (23), 203 (63), 189 (73). ^1H -NMR δ : 1.097 (3H, s, H-23), 1.057 (3H, s, H-24), 1.073 (3H, s, H-25), 1.023 (3H, s, H-26), 1.042 (3H, s, H-27), 0.842 (3H, s, H-28), 0.874 (3H, s, H-29), 0.874 (3H, s, H-30), 5.208 (1H, dd, $J=3.4, 3.7$ Hz, H-12).³⁾

LiAlH₄ Reduction of 21 **21** (20 mg) was reduced with LiAlH_4 in anhyd. ether and treated in the usual manner. The reaction product was purified by silica CC to give **2'** (6 mg) and **22** (4 mg).

Acknowledgements The authors are grateful to Mr. Y. Takase for mass measurements.

References

- 1) Nakane T., Arai Y., Masuda K., Ishizaki Y., Ageta H., Shiojima K., *Chem. Pharm. Bull.*, **47**, 543–547 (1999).
- 2) Shiojima K., Arai Y., Nakane T., Ageta H., Cai S.-Q., *Chem. Pharm. Bull.*, **45**, 1608–1610 (1997).
- 3) Shiojima K., Masuda K., Suzuki H., Lin T., Ooishi Y., Ageta H., *Chem. Pharm. Bull.*, **43**, 1634–1639 (1995).
- 4) Ageta H., Shiojima K., Arai Y., Suzuki H., Kiyotani T., *Chem. Pharm. Bull.*, **42**, 39–44 (1994).
- 5) Shiojima K., Sasaki Y., Ageta H., *Chem. Pharm. Bull.*, **41**, 268–271 (1993).
- 6) Shiojima K., Arai Y., Masuda K., Takase Y., Ageta T., Ageta H., *Chem. Pharm. Bull.*, **40**, 1683–1690 (1992).