Isolation and Structure of Monomethylated GM₃-Type Ganglioside Molecular Species from the Starfish *Luidia maculata*

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Two monomethylated GM₃-Type ganglioside molecular species, 1 and 2, have been obtained from the polar lipid fraction of the chloroform/methanol extract of the starfish *Luidia maculata*. The structures of these gangliosides have been determined on the basis of chemical and spectroscopic evidence as 1-O-[8-O-methyl-(N-acetyl- α -D-neuraminosyl)-(2 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl]-ceramide (1) and 1-O-[8-O-methyl-(Nglycolyl- α -D-neuraminosyl)-(2 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl]-ceramide (2). The ceramide moieties were composed of heterogeneous unsubstituted fatty acid, 2-hydroxy fatty acid, sphingosine and phytosphingosine units. Compound 1, designated as LMG-3, represents new ganglioside molecular species. Compound 2 was a known ganglioside molecular species.

Key words glycosphingolipid; ganglioside; starfish; Luidia maculata

In the course of our continuing research on biologically active glycosphingolipids (GSLs) from echinoderms, a series of studies on the isolation and structure elucidation of the GSLs from starfish have been performed in our laboratory.¹⁻⁶⁾ In the study of the GSLs of the starfish *Luidia maculata* (Yatsudesunahitode in Japanese), we reported on the isolation and structure of a sulfatide and a ganglioside molecular species, LMG-1⁷⁾ and LMG-2.⁸⁾ Continuing the previous studies, further isolation and characterization of gangliosides from *L. maculata* was conducted. In this paper, we report on the isolation and structure of two ganglioside molecular species from the whole bodies of *L. maculata*.

The polar lipid fraction, which was obtained from the chloroform/methanol extract of the whole bodies of *L. maculata*, was subjected to repeated column chromatography to give two ganglioside molecular species, **1** and **2**, each showing a single spot on silica gel thin-layer chromatography (TLC).

In its ¹³C-NMR spectrum (Fig. 1, Table 1), 1 exhibits the characteristic signals of a sphingosine-type ceramide, possessing a 2-hydroxy fatty acid and a sugar moiety at C-1 [δ : 69.6 (C-1), 54.1 (C-2), 72.5 (C-3), 131.7 (C-4), 132.8 (C-5), 175.5 (C-1'), 72.5 (C-2')]. The 13 C-NMR spectrum of 1 also features signals due to three anomeric carbons at δ : 106.0, 105.3 and 99.9, one of which (δ : 99.9) is a quaternary carbon signal, indicating the presence of a sialic acid residue. The negative FAB-MS exhibits a series of quasi-molecular ion peaks $[M-H]^-$ at m/z: 1230—1330. Therefore, 1 is suggested to be a molecular species of a sphingosine-type ganglioside, possessing 2-hydroxy fatty acid groups and three monosaccharide units. It is further presumed to have mainly normal-type fatty acids and normal and ante-iso type longchain bases, since the carbon signals for the terminal methyl groups are observed at δ : 14.3 (normal form) and δ : 11.6 and 19.4 (ante-iso form) in the ¹³C-NMR spectrum (Fig. 1, Table 1).

The structure of the ceramide moiety was examined first. When **1** was methanolyzed with methanolic hydrochloric acid, a mixture of fatty acid methyl esters (FAM) and longchain bases (LCB) was obtained, together with methyl glycosides. The FAM mixture was analyzed by GC-MS, which revealed the presence of three components. These were characterized as methyl octadecanoate, methyl 2-hydroxydo-cosanoate (major) and methyl 2-hydroxytricosanoate. The LCB mixture was found to be composed of 2-amino-1,3-di-hydroxy-4-octadecene, 2-amino-1,3-dihydroxy-4-nonadecene (major), 2-amino-1,3,4-trihydroxy-heptadecane and 2-amino-1,3,4-trihydroxy-nonadecane, based on GC-MS analysis of its TMS derivative (Fig. 1).

The stereochemistry of the ceramide moiety is presumed to be (2S, 3R, 4E, 2'R), since the aforementioned ¹³C-NMR signals assignable to C-1, 2, 3, 4, 5 and 2' of **1** are in good agreement with those of the sphingosine-type glucocerebroside molecular species possessing (2S, 3R, 4E, 2'R) configurations.⁹

The structure of the trisaccharide moiety of 1 was established as follows. The GLC analysis of the TMS derivatives of the methyl glycosides, which was obtained by methanolysis of 1 (vide supra), showed the existence of one mole each of glucose (Glc) and galactose (Gal). A detailed analysis of the ¹³C-NMR spectrum of **1** revealed the characteristic signals [\delta: 173.0 (C-1), 99.9 (C-2), 42.2 (C-3), 54.1 (C-5), 62.3 (C-9), 172.9 (C-10), and 22.9 (C-11)] of an N-acetylneuraminic acid (NeuAc) derivative residue together with the signal due to a methoxy group (δ : 58.7) (Table 1). The existence of the methoxy group in the NeuAc residue is also supported by the ¹H-NMR spectrum of **1** showing a singlet signal at δ : 3.78. In the negative FAB-MS of 1, molecular ion and fragment ion peaks arising from cleavage of the glycosidic linkages are observed at m/z: 1230—1330, 930—1030, 770-850, and 600-650, indicating the presence of the trisaccharide moiety, NeuAc (OMe)→Hexose→Hexose, as shown in Fig. 3.

Methylation of 1, according to the Ciucanu–Kerek method,¹⁰⁾ afforded the permethylated product 3a. Partially methylated alditol acetates prepared from 3a were analyzed by GC-MS and identified as the alditols derived from 3-linked hexopyranose (S-1) and 4-linked hexopyranose (S-2). The structure of the sialic acid moiety was established as follows. Since the sialic acid residue has a methoxy group, per-



Fig. 1. Structure of Compound 1 (LMG-3)

Table 1. ¹³C-HMR Spectral Data (δ Values) of the Gangliosides in C₅D₅N

(2	1	4	2
Ceramide				
1	(t)	69.6	70.0	69.7
2	(d)	54.1	54.5	51.4
3	(d)	72.5	72.6	75.1
4	(d)	131.7	131.8	72.4
5	(d)	132.8	132.8	
1'	(s)	175.5	175.8	175.8
2'	(d)	72.5	72.6	72.4
$CH_3^{(a)}$	(q)	14.3	14.3	14.2
CH ₃ ^{b)}	(q)	11.6	11.6	11.5
CH ₃ ^{c)}	(q)	19.4	19.4	19.3
Glc				
1	(d)	105.3	105.0	104.9
2	(d)	74.6	74.7	74.4
3	(d)	76.3	76.6	76.2
4	(d)	81.8	81.8	82.0
5	(d)	76.4	77.3	76.2
6	(t)	61.7	62.1	61.7
Gal				
1	(d)	106.0	105.7	105.7
2	(d)	70.1	72.4	70.1
3	(d)	78.2	75.2	77.7
4	(d)	69.1	70.1	68.9
5	(d)	76.5	76.5	76.3
6	(t)	61.7	62.0	61.7
NeuAc (GC) (172.0^d) (77.7^d)				
1	(s)	173.04)		173.7
2	(s)	99.9		100.6
3	(t)	42.2		42.4
4	(d)	68.5		68.2
5	(d)	54.1		53.8
6	(d)	/5.6		/4.4
/	(D)	69.2 81.8		09./
8	(a)	81.8		81.8
9	(I) (a)	02.3		01./ 175.5 ^e)
10	(s)	1/2.9		1/3.37
	(\mathbf{q}, \mathbf{r})	22.9 58 7		02.3 58.6
UCH ₃	(Ψ)	30.7		30.0

a-c) Terminal methyl groups in the normal and *ante*-iso type of side chain (see Fig. 1). d, e) Assignments may be interchanged in each vertical column.

trideuteriomethylated product **3b** was prepared. Upon methanolysis followed by acetylation of **3b**, the partially trideuteriomethylated NeuAc (S-3) derived from the terminal 8-*O*-Me-NeuAc was detected by GC-MS analysis. **1** was hydrolyzed with 5% aq. AcOH to give a ceramide dihexoside **4**, which was determined as ceramide lactoside $[1-O-\beta-D-galac$ topyranosyl- $(1\rightarrow 4)-\beta-D-glucopyranosyl$ ceramide] by comparison of the ¹³C-NMR spectral data of sugar moiety with a reference compound.¹¹

On the basis of the above evidence, the trisaccharide moiety of **1** must be 8-*O*-Me-NeuAc- $(2\rightarrow 3)$ - β -galactopyranose- $(1\rightarrow 4)$ - β -glucopyranose. The configuration of NeuAc is believed to be α on the basis of its anomeric carbon signal (δ : 99.9)^{1,12}) in the ¹³C-NMR spectrum of **1**. Consequently, if NeuAc, Gal and Glc are assumed to belong to the most commonly found D-series, respectively, then **1** is the 8-*O*-methyl-(N-acetyl- α -D-neuraminosyl)- $(2\rightarrow 3)$ - β -D-galactopyranosyl- $(1\rightarrow 4)$ - β -D-glucopyranoside of a ceramide, composed of heterogeneous fatty acid and long-chain base units. The major components of the fatty acid and long-chain base moiety of **1** are (2R)-2-hydroxydocosanoic acid and (2S,3R,4E)-C₁₉sphingosine, respectively (Fig. 1).

To the best of our knowledge, compound (1) represents a new GM₃ type ganglioside molecular species found to contain a monomethylated NeuAc moiety, and designated as LMG-3. Compound **2** was identified as the 8-*O*-methyl-(*N*glycolyl- α -D-neuraminosyl)-(2 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-glucopyranoside of a ceramide composed of (2*S*,3*S*,4*R*)-C₁₇-phytosphingosine and (2*R*)-2-hydroxydocosanoic acid as major components (see Fig. 2), on the basis of chemical and spectroscopic evidence like **1**. Compound (**2**) has already been obtained from the starfish *Aphelasterias japonica*¹³⁾ and *Asterias amurensis versicolor*,²⁾ although it differs from them slightly in the ceramide moiety. The biological activity of these gangliosides will be examined in due course.



Fig. 2. Structure of Compound 2





Fig. 3. The Negative Ion FAB-MS Fragmentation of Compounds 1 and 2

Experimental

Melting points were determined on a micro melting point apparatus (Yanako MP-3) without correction. IR spectra were obtained on a Jasco FT/IR-410 infrared spectrophotometer. NMR spectra were recorded on a Varian Unity-500 spectrometer (¹H: 500 MHz, ¹³C: 125 MHz). Negative-ion FAB-MS spectra were acquired with a JEOL SX-102 mass spectrometer (xenon atom beam; matrix, HMPA-TEG). GC-MS was taken with a Shimadzu QP-1000 [EI mode; ionizing potential, 70 eV; separator and ion source temperature 250 °C; column (A), TC-1701 (0.25 mm×30 m, GL Science Inc.); column (B), CBP10-W12-100 (0.53 mm×12 m, Shimadzu); carrier gas, He]. GC was run on a Shimadzu GC-14B [FID mode; column, Fused Silica Capillary Column DB-17 (0.32 mm×30 m, J & W Scientific); carrier, N₂].

Separation of 1 and 2 For the extraction and fractionation of the crude ganglioside fraction from the whole bodies of the starfish *Luidia maculata* (17 kg), the preceding paper should be referred to.⁸⁾ The crude ganglioside fraction, the 100% MeOH eluate, was successively separated by chromatography on silica gel (solvent CHCl₃–MeOH–7% NH₃ aq., 6:4:1) to afford 1 (13 mg) (*Rf*=0.38) and 2 (31 mg) (*Rf*=0.36) [silica gel TLC, solvent CHCl₃–MeOH–H₂O (6:4:0.9)].

Compound **1** (LMG-3): Amorphous powder, mp 154—160 °C. IR (KBr) cm⁻¹: 3387 (OH), 1645, 1542 (amide). Negative-ion FAB-MS m/z: 1230—1330 $[M-H]^-$ series (see Fig. 3). ¹H-NMR (C₅D₅N) δ : 1.94 (3H, s, COCH₃), 3.78 (3H, s, OMe). ¹³C-NMR: See Table 1.

Compound **2**: Amorphous powder, mp 135—140 °C. IR (KBr) cm⁻¹: 3397 (OH), 1647, 1541 (amide). Negative-ion FAB-MS m/z: 1250—1350 $[M-H]^-$ series (see Fig. 3). ¹H-NMR (C₅D₅N) δ : 3.80 (3H, s, OMe). ¹³C-NMR: See Table 1.

Methanolysis of 1 Compound **1** (1 mg) was heated with 5% HCl in MeOH (1 ml) at 70 °C for 12 h. The reaction mixture was then extracted with *n*-hexane, and the extract was concentrated *in vacuo* to yield a mixture of FAM. The MeOH layer was concentrated under a stream of N₂ to give a mixture of LCB and methyl glycosides.

GC-MS Analysis of FAM from 1 A FAM mixture from 1 was subjected to GC-MS [column (A), column temp. 180–250 °C (rate of temp. increase 4 °C/min)]. The results were as follows: methyl octadecanoate, t_R [min]=3.6, m/z: 298 (M⁺), 255 (M-49)⁺; methyl 2-hydroxydocosanoate (major), t_R =8.6, m/z: 370 (M⁺), 311 (M-59)⁺; methyl 2-hydroxytri-

cosanoate, $t_{\rm R} = 9.4$, m/z: 384 (M⁺), 325 (M-59)⁺.

GC-MS Analysis of TMS Ethers of LCB from 1 The mixture of LCB and methyl glycoside from **1** was heated with 1-(trimethylsilyl) imidazole– pyridine (1:1) for 30 min at 70 °C and the reaction mixture (TMS ethers) was analyzed by GC-MS [column (A), column temp. 180—250 °C (rate of temp. increase 4 °C/min)]. The results were as follows: 2-amino-1,3-dihydroxy-4-octadecene, $t_{\rm R}$ [min]=7.9, m/z: 340 (M-103)⁺, 132; 2-amino-1,3,4-trihydroxy-heptadecane, $t_{\rm R}$ =8.8, m/z: 326 (M-193)⁺, 285 (M-234)⁺, 132; 2-amino-1,3,4-trihydroxy-anonadecene (major), $t_{\rm R}$ =9.4, m/z: 354 (M-103)⁺, 132; 2-amino-1,3,4-trihydroxy-nonadecane, $t_{\rm R}$ =11.9, m/z: 354 (M-193)⁺, 313 (M-234)⁺, 132.

GC Analysis of TMS Ethers of Methyl Glycoside from 1 The mixture of TMS ethers of LCB and methyl glycosides was analyzed by GC [column temp.: 100–250 °C (rate of temp. increase 3 °C/min)]: $t_{\rm R}$ [min]=22.0 and 22.7 (methyl galactopyranosides); $t_{\rm R}$ =23.8 and 24.0 (methyl glucopyranosides).

Methylation of 1 (Ciucanu–Kerek Method) NaOH–dimethylsulfoxide (DMSO) solution, which was prepared from powdered NaOH (40 mg) and DMSO (1 ml), and CH_3I (or CD_3I) (0.2 ml) were added to 1 (2 mg), and the mixture was stirred for 30 min. The reaction mixture was then diluted with H_2O (15 ml), extracted with CHCl₃ (10 ml×3), and the CHCl₃ phases were washed with H_2O , and the solvent was evaporated *in vacuo* to give permethylated (or pertrideuteriomethylated) 1, denoted 3a (or 3b).

Preparation and GC-MS Analysis of Partially Methylated Alditol Acetates from 3a Compound 3a (0.7 mg) was heated with 90% HCOOH-10% CF₃COOH (1:1) (1 ml) at 70 °C for 18 h in a small-volume sealed vial, and then the solvents were evaporated in vacuo. The residue was dissolved in $\rm H_2O$ (5 ml), and 28% $\rm NH_3$ (2 drops), and $\rm NaBD_4$ (10 mg) were added. After allowing the mixture to stand at room temp. for 7 h, it was acidified with AcOH to pH=3.5 and concentrated in vacuo. H₃BO₃ present in the residue was removed by distillation with MeOH (three times). The residue was heated with Ac₂O-C₅H₅N (1:1, 0.3 ml) at 70 °C for 2 h. After dilution with H₂O, the mixture was extracted with $CHCl_2$ (0.2 ml×3). The combined CHCl₃ extracts were washed with H₂O, and the solvent was evaporated to give partially methylated alditol acetate. The acetate was subjected to GC-MS [column (A), column temp. 150 °C], with the following results: S-1, $t_{\rm R}$ [min]=14.3, m/z: 45, 118, 161, 234 [1,3,5-tri-O-acetyl-2,4,6-tri-O-methylhexitol (derived from 3-linked hexopyranose)]; S-2, t_R=15.3, m/z: 45, 118, 233 [1,4,5-tri-O-acetyl-2,3,6-tri-O-methylhexitol (derived from 4-linked hexopyranose)].

Preparation and GC-MS Analysis of Partially Trideuteriomethylated NeuAc Derivative from 3b Compound **3b** (0.5 mg) was heated with 5% HCl in MeOH (0.5 ml) at 70 °C for 4 h in a small-volume sealed vial. The reaction mixture was then concentrated *in vacuo*, and the residue (methanolysate) was heated with Ac₂O-C₅H₅N (1:1, 0.2 ml) at 70 °C for 2 h. The resulting mixture was diluted with H₂O and extracted with CHCl₃ (0.2 ml×3), the combined CHCl₃ extracts were washed with H₂O, and the solvent was evaporated *in vacuo*. The residue was subjected to GC-MS [column (B), column temp.: 180-250 °C (rate of temp. increase 4 °C/min)]: S-3, t_{R} =4.8 min, *m/z*: 135, 280, 327, 360, 371, 388 [methyl *N*-acetyl-*N*-trideuteriomethyl-2,8-di-*O*-methyl-4,7,9-tri-*O*-trideuteriomethylneuraminate (derived from terminal 8-*O*-Me-NeuAc)].

Partial Hydrolysis of 1 Compound 1 (5 mg) was heated with 5% aq. AcOH (3 ml) at 90 °C for 4 h. The mixture was then extracted with AcOEt/*n*-BuOH (2 : 1), the organic layer was concentrated *in vacuo*, and the residue was chromatographed on silica gel [CHCl₃-MeOH-H₂O (8 : 2 : 0.1)] to give 4 (2 mg) as an amorphous powder (13 C-NMR: Table 1).

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References

- Higuchi R., Inagaki K., Natori T., Komori T., Kawajiri S., *Liebigs Ann. Chem.*, **1991**, 1–10 (1991).
- Higuchi R., Inukai K., Jhou J.-X., Honda M., Komori T., Tsuji S., Nagai Y., *Liebigs Ann. Chem.*, **1993**, 359–366 (1993).
- Higuchi R., Matsumoto S., Fujita M., Komori T., Sasaki T., *Liebigs* Ann., 1995, 545–550 (1995).
- Miyamoto T., Inagaki M., Isobe R., Tanaka Y., Higuchi R., Iha M., Teruya K., *Liebigs Ann.*, **1997**, 931–936 (1997).
- 5) Inagaki M., Isobe R., Higuchi R., Eur. J. Org. Chem., 1999, 771-774

(1999).

- Miyamoto T., Yamamoto A., Wakabayashi M., Nagaregawa Y., Inagaki M., Higuchi R., Iha M., Teruya K., *Eur. J. Org. Chem.*, 2000, 2295– 2301 (2000).
- Kawatake S., Inagaki M., Isobe R., Miyamoto T., Higuchi R., *Liebigs* Ann., 1997, 1797–1800 (1997).
- Kawatake S., Inagaki M., Miyamoto T., Isobe R., Higuchi R., *Eur. J.* Org. Chem., **1999**, 765–769 (1999).
- Higuchi R., Inagaki M., Togawa K., Miyamoto T., Komori T., *Liebigs* Ann. Chem., 1994, 79–81 (1994).
- 10) Ciucanu I., Kerek F., Carbohydr. Res., 131, 209-217 (1984).
- Sugiyama S., Honda M., Higuchi R., Komori T., *Liebigs Ann. Chem.*, 1991, 349—356 (1991).
- Higuchi R., Mori T., Sugata T., Yamada K., Miyamoto T., *Eur. J. Org. Chem.*, **1999**, 3175–3178 (1999).
- Smirnova G. P., Kochetkov N. K., Sadovskaya V. L., *Biochim. Biophys.* Acta, 920, 47–55 (1987).