

## A New Pyrroloquinazoline Alkaloid from *Linaria vulgaris*

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A new alkaloid, 1,2,3,9-tetrahydropyrrolo(2,1-*b*)quinazolin-1-carboxylic acid (**1**), together with eight known compounds, 7-hydroxy vasicine (**2**), benzyl alcohol  $\beta$ -D-(2'-*O*- $\beta$ -xylopyranosyl)glucopyranoside (**3**), benzyl alcohol *O*- $\beta$ -D-glucopyranoside (**4**), benzyl alcohol *O*- $\beta$ -D-primveroside (**5**), 3,5-dimethyl-4-hydroxy benzaldehyde (**6**), gluco-syringic acid (**7**), syringin (**8**), and liriiodendrin (**9**), were isolated from the plants of *Linaria vulgaris*. Their structures were established by spectroscopic methods.

**Key words** *Linaria vulgaris*; Scrophulariaceae; pyrroloquinazoline alkaloid; 1,2,3,9-tetrahydropyrrolo(2,1-*b*)quinazolin-1-carboxylic acid

*Linaria vulgaris* MILL. is a grassy plant that occurs in northeast China. The plant is used in traditional folk medicine for the treatment of coughs and asthma and as an expectorant.<sup>1)</sup> Previous phytochemical investigations of the plant have revealed the alkaloids, flavonoids, triterpenoids, steroids, and iridoid glucosides.<sup>2–9)</sup> From the ethanol extract of the plants of *L. vulgaris*, we isolated a new compound **1** and eight known compounds. By comparison of physical and spectroscopic properties (mp, IR, MS, <sup>1</sup>H- and <sup>13</sup>C-NMR spectra), the known compounds were identified as 7-hydroxy vasicine (**2**),<sup>10)</sup> benzyl alcohol  $\beta$ -D-(2'-*O*- $\beta$ -xylopyranosyl)glucopyranoside (**3**),<sup>11,12)</sup> benzyl alcohol *O*- $\beta$ -D-glucopyranoside (**4**),<sup>13)</sup> benzyl alcohol *O*- $\beta$ -D-primveroside (**5**),<sup>14)</sup> 3,5-dimethyl-4-hydroxy benzaldehyde (**6**), gluco-syringic acid (**7**),<sup>15,16)</sup> syringin (**8**),<sup>17)</sup> and liriiodendrin (**9**).<sup>18)</sup> In this article, we present the isolation and structural determination of the new compound and give the <sup>1</sup>H- and <sup>13</sup>C-NMR data of **2**, which have not been reported previously.

### Results and Discussion

Compound **1**, called linarinic acid, was obtained as colorless needles, and the molecular formula was determined to be C<sub>12</sub>H<sub>12</sub>O<sub>2</sub>N<sub>2</sub> by high resolution electron impact (HR-EI)-MS. The IR spectrum indicated the presence of a carboxyl group (1666 cm<sup>-1</sup>) and aromatic ring (1614, 1586, 1500 cm<sup>-1</sup>). The <sup>1</sup>H-NMR spectrum indicated the presence of an *ortho*-disubstituted benzene [ $\delta$  7.25 (ddd, *J*=7.8, 7.5, 1.9 Hz), 7.16 (ddd, *J*=7.8, 7.5, 1.0 Hz), 7.13 (dd, *J*=7.8, 1.9 Hz), 6.97 (dd, *J*=7.8, 1.0 Hz)] and AB system of CH<sub>2</sub> [4.92, 4.68 (d, *J*=15.2 Hz)]. It gave signals at  $\delta$  2.91 (m, 2H), 2.21 (m, 1H), 2.53 (m, 1H), and 4.21 (dd, *J*=9.0, 4.5 Hz), which were assigned to the structural fragment –CH<sub>2</sub>–CH<sub>2</sub>–CH– by <sup>1</sup>H–<sup>1</sup>H correlation spectroscopy (COSY). The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra suggested **1** to be 1,2,3,9-tetrahydropyrrolo[2,1-*b*]quinazoline alkaloid.<sup>19)</sup> The EI-MS peak at *m/z* 171 (M<sup>+</sup>–COOH) and <sup>13</sup>C-NMR signal at 175.63 showed the presence of a carboxyl group. In the heteronuclear multiple bond connectivity (HMBC) spectrum, the long-range coupling between the H-1 proton at  $\delta$  4.21 and the carbon signals at  $\delta$  175.63 (COOH), 165.03 (C-3a), 46.44 (C-9), 30.41 (C-3), and 25.57 (C-2), suggested that the carboxyl group was linked with C-1. Thus compound **1** was assigned to be 1,2,3,9-tetrahydropyrrolo(2,1-*b*)quinazolin-1-carboxylic acid, which was confirmed by single-crystal X-ray diffraction.

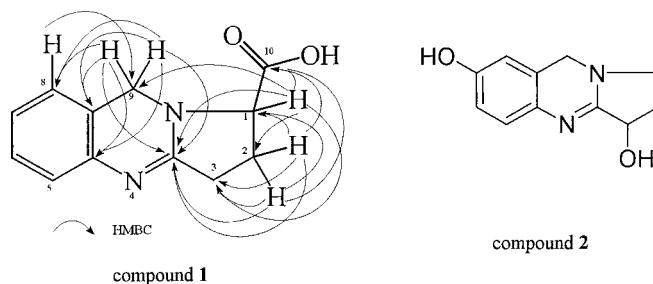


Fig. 1. HMBC Correlations Observed for Compound **1** and the Structure of Compound **2**

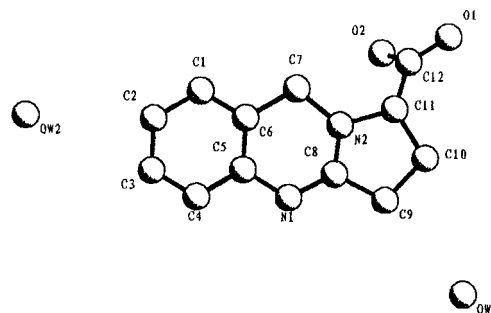


Fig. 2. PLUTO Drawing of Compound **1**

### Experimental

**General Experimental Procedures** Melting points were uncorrected. Optical rotations were measured on a Perkin-Elmer 241MC polarimeter. The UV spectra were obtained with a Shimadzu UV-260 spectrophotometer. The IR spectra were recorded on a Bruker IR S-55 instrument. <sup>1</sup>H- and <sup>13</sup>C-NMR spectra were measured on a Bruker AC(E)-300 instrument. EI-MS and HR-MS were obtained on a VG-70SE mass spectrometer. Chromatography was performed with D 101 macroporous resin and silica gel (200–300 mesh).

**Plant Material** The plants of *L. vulgaris* (MILL.) were collected in Heilongjiang Province, People's Republic of China, in August 1995 and were identified by Professor Shiwen Su, Department of Chinese Traditional Medicine, Shenyang Pharmaceutical University. A voucher specimen (LV 960801) has been deposited at the Department of Natural Products Chemistry, Shenyang Pharmaceutical University.

**Extraction and Isolation** The air-dried aerial parts of plant (10 kg) were extracted with 95% ethanol under reflux (80×31) for 2 h each time. The alcohol extract was concentrated and successively extracted with petroleum ether, CHCl<sub>3</sub>, EtOAc, and *n*-BuOH. The CHCl<sub>3</sub>-soluble fraction (76 g) was chromatographed over a silica gel (700 g) column eluted with CHCl<sub>3</sub> and CHCl<sub>3</sub>–MeOH (9:1) to yield **6** (8 mg). The *n*-BuOH-soluble fraction (200 g) was chromatographed over a D 101 macroporous resin column with 0%, 20%, 40%, 60% and 95% EtOH in H<sub>2</sub>O as eluants (5000 ml of each eluant). The fraction (129 g) eluted with H<sub>2</sub>O was subjected to silica gel (1000 g) column chromatography eluted with a solvent system of CHCl<sub>3</sub>–MeOH (9:1) to afford **7** (6 mg). The combined fractions (26 g) eluted with

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CHCl<sub>3</sub>-MeOH (8:2) were isolated on a silica gel (250 g) column eluted with an EtOAc-MeOH (92:8, 6:4) gradient to afford **2** (14 mg) and **1** (25 mg). The fraction (47 g) eluted with 20% EtOH was separated on column chromatography over 500 g silica gel with a gradient (5000 ml of each eluant) of CHCl<sub>3</sub>-MeOH (9:1, 88:12, 85:15) to give **4** (100 mg), **5** (25 mg), **3** (34 mg), **8** (14 mg) and **9** (20 mg).

**Linarinic Acid (1):** Colorless needles (MeOH). mp 218 °C (dec.);  $[\alpha]_D^{18}$  -217° ( $c=0.01$ ,  $l=0.2$ , MeOH). UV  $\lambda_{max}$  (MeOH) nm: 281, 211. IR  $\nu_{max}$  (KBr) cm<sup>-1</sup>: 3331, 2915, 2863, 2759, 1666, 1614, 1586, 1500, 1460, 1385, 1291, 764. <sup>1</sup>H-NMR (CD<sub>3</sub>OD, 300 MHz)  $\delta$ : 2.21 (1H, m, H-2), 2.53 (1H, m, H-2), 2.91 (2H, m, H-3), 4.21 (1H, dd,  $J=9.0, 4.5$  Hz, H-1), 4.68, 4.92 (1H each, d,  $J=15.0$  Hz, H-9), 6.97 (1H, dd,  $J=7.8, 1.0$  Hz, H-5), 7.13 (1H, dd,  $J=7.8, 1.9$  Hz, H-8), 7.16 (1H, ddd,  $J=7.8, 7.5, 1.0$  Hz, H-7), 7.26 (1H, ddd,  $J=7.8, 7.5, 1.9$  Hz, H-6); <sup>13</sup>C-NMR (CD<sub>3</sub>OD, 75 MHz)  $\delta$ : 25.57 (t, C-2), 30.41 (t, C-3), 46.44 (t, C-9), 69.71 (d, C-1), 118.99 (d, C-5), 127.43 (d, C-7), 128.05 (d, C-8), 130.01 (d, C-6 and s, C-8a), 134.65 (s, C-4a), 165.03 (s, C-3a), 175.63 (s, C-10); EI-MS  $m/z$  (rel. int.) 216 [M]<sup>+</sup> (94), 215 (100), 214 (37), 171 (97), 144 (96.5); HR-EI-MS  $m/z$  216.0900 (Calcd for C<sub>12</sub>H<sub>12</sub>O<sub>2</sub>N<sub>2</sub>, 216.0899).

**7-Hydroxy Vasicine (2):** Colorless needles (MeOH). mp 260 °C (dec.).  $[\alpha]_D^{18}$  -35.5° ( $c=0.013$ ,  $l=0.2$ , MeOH). UV  $\lambda_{max}$  (MeOH) nm: 292, 204. <sup>1</sup>H-NMR (CD<sub>3</sub>OD, 300 MHz)  $\delta$ : 2.08 (1H, m, H-2), 2.62 (1H, m, H-2), 3.67 (2H, m, H-1), 4.72, 4.80 (1H each, d,  $J=15.6$  Hz, H-9), 5.08 (1H, t,  $J=8.0$  Hz, H-3), 6.60 (1H, d,  $J=2.4$  Hz, H-8), 6.72 (1H, dd,  $J=8.7, 2.4$  Hz, H-6), 6.98 (1H, d,  $J=8.7$  Hz, H-5). <sup>13</sup>C-NMR (CD<sub>3</sub>OD, 75 MHz)  $\delta$ : 30.94 (t, C-2), 47.51 (t, C-9), 51.63 (t, C-1), 72.32 (d, C-3), 114.44 (d, C-8), 116.89 (d, C-6), 119.57 (s, C-8a), 119.64 (d, C-5), 124.06 (s, C-4a), 158.22 (s, C-7), 165.51 (s, C-3a). EI-MS  $m/z$  (rel. int.): 204 (M<sup>+</sup>, 68), 203 (100), 175 (12), 147 (14), 38 (6.5), 36 (19.6).

**Single-Crystal X-Ray Structure Determination** Crystal data for **1**: C<sub>12</sub>H<sub>12</sub>O<sub>2</sub>N<sub>2</sub>·2H<sub>2</sub>O, MW=252.11, orthorhombic, space group *P*2<sub>1</sub>2<sub>1</sub>2<sub>1</sub>,  $a=7.240(1)$  Å,  $b=8.051(2)$  Å,  $c=21.127(5)$  Å,  $V=1231.5(5)$  Å<sup>3</sup>,  $Z=4$ ,  $D_c=1.355$  g/cm<sup>3</sup>. The crystal of **1** with approximate dimensions 0.15×0.2×0.4 mm was selected for X-ray crystallographic analysis. The X-ray intensity data were measured on a MAC DIP2030K X-ray diffractometer with MoK $\alpha$  radiation. The detector-to-crystal distance was 100 mm. A total of 1137 unique reflections was recorded, of which 895 reflections were considered observed on the basis  $|F|^2 > 8.0\sigma|F|^2$ . The structure was solved by direct methods with the use of the SHELX-86 program. All the hydrogen atoms were located from a difference Fourier map and hydrogen parameters were refined. Nonhydrogen atoms were given anisotropic thermal parameters. Final *R*-factors were  $R=0.075$  and  $R_w=0.073$ .

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## References

- 1) Jiangsu College of New Medicine, "A Dictionary of the Traditional Chinese Medicines," People's Hygiene Publisher, Beijing, 1977.
- 2) Men Shikov G. P., Ban'kovskii A. I., Frolova V. I., *Zhur Obshchei Khim.*, **29**, 3846 (1959) [*Chem. Abstr.*, **54**, 19744i (1960)].
- 3) Morita N., Shimizu M., Arisawa M., Kobayashi K., *Yakugaku Zasshi*, **94**, 913-916 (1974).
- 4) Hua H.-M., Sun J., Li X., *Chin. Trad. Herbal Drugs*, **30**, 332-334 (1999).
- 5) Sticher O., *Phytochemistry*, **10**, 1974-1975 (1971).
- 6) Ilieva E. I., Handjieva N. V., Popov S. S., *Phytochemistry*, **31**, 1040-1041 (1992).
- 7) Ilieva E. I., Handjieva N. V., Spassov S., Popov S. S., *Phytochemistry*, **32**, 1068-1070 (1993).
- 8) Hua H.-M., Hou B.-L., Li W., Li X., Zhang Y., *Chin. Trad. Herbal Drugs*, **31**, 409-412 (2000).
- 9) Hua H.-M., Li X., Zhang H.-Q., *J. Shenyang Pharm. Univ.*, **17**, 40-42, 48 (2000).
- 10) Ghosal S., Chauhan R. B. P. S., Mehta R., *Phytochemistry*, **14**, 830-832 (1975).
- 11) Sudo H., Ide T., Otsuka H., Hirata E., Takushi A., Shinzato T., Takeda Y., *Chem. Pharm. Bull.*, **48**, 542-546 (2000).
- 12) Kamel M., Mohamed K. M., Hassanean H. A., Ohtani K., Kasai R., Yamasaki K., *Phytochemistry*, **55**, 353-357 (2000).
- 13) Miyase T., Ueno A., Takizawa N., *Chem. Pharm. Bull.*, **35**, 1109-1117 (1987).
- 14) Kijima H., Ide T., Otsuka H., *Phytochemistry*, **44**, 1551-1557 (1997).
- 15) Xu L., Yang X., Li B., *Chin. J. Trad. Chin. Med.*, **19**, 675-676 (1994).
- 16) Sano K., Sanada S., Ida Y., *Chem. Pharm. Bull.*, **39**, 865-870 (1991).
- 17) Karasawa H., *Yakugaku Zasshi*, **106**, 721-724 (1986).
- 18) Jolad S. D., Hoffmann J. J., Cole J. R., *J. Org. Chem.*, **45**, 1327-1329 (1980).
- 19) Joshi B., Bai Y., Puar M. S., Dubose K. K., Pelletier S. W., *J. Nat. Prod.*, **57**, 953-962 (1994).