

Water-Soluble Constituents of Cumin: Monoterpenoid Glucosides

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From the water-soluble portion of the methanol extract of cumin (fruit of *Cuminum cyminum* L.), which has been used as a spice and medicine since antiquity, sixteen monoterpenoid glucosides, including twelve new compounds, were isolated. Their structures were clarified by spectral investigation.

Key words cumin; *Cuminum cyminum* fruit; *p*-menthane glucoside; hydroxycuminyl glucoside

Cumin [*Cuminum cyminum* L.; Umbelliferae] is known to have been cultivated since antiquity, and is now mainly cultivated in North Africa, Iran, India, Indonesia and China. Its fruit has been used as a popular aromatic herb and spice, such as a main constituent of curry powder. The fruit has also been used for medicinal purposes as a remedy for diarrhea, flatulence and indigestion.¹⁾ It contains an essential oil (3–4%) rich in cuminaldehyde (main; 35–60%), and α - and β -pinene, δ -3-carene, 1,8-cineole, α - and β -phellandrene, *p*-cymene, limonene, α - and γ -terpinene, α -terpineol, terpinene-4-ol cuminyl alcohol, *trans*-dihydrocarvone (menthane type monoterpenoids), myrcene, linalool (acyclic monoterpenoids), β -caryophyllene, β -farnesene, β -elemene (sesquiterpenoids) *et al.* were reported as other constituents.^{2,3)} However, only two flavonoid glycosides (7-*O*- β -D-glucopyranosides of apigenin and luteolin) have been published as constituents of the water-soluble portion of this fruit.⁴⁾ In continuation of our studies on the water-soluble constituents of spices,⁵⁾ and to show the relationship between the essential oil and the water-soluble constituents, we undertook a detailed investigation of this fruit. In this paper, we discuss the isolation and structural elucidation of monoterpenoid glucosides.

Commercial cumin was extracted with 70% methanol, and the methanolic extract was suspended in water and successively extracted with ether and ethyl acetate. The aqueous layer was chromatographed on Amberlite XAD-II to give water and methanol eluate fractions. The methanol eluate fraction (glycoside fraction) was chromatographed on Sephadex LH-20, then subjected to a combination of silica gel, Lobar RP-8 column chromatography and HPLC to isolate sixteen monoterpenoid glucosides (**1**–**16**). Among them, twelve glucosides (**1**–**3**, **5**, **6**, **9**, **11**–**16**) are new compounds. All new glucosides described in this paper were β -D-glucopyranosides as shown by their ¹H- and ¹³C-NMR data (Tables 1 and 2). This was confirmed by hydrolysis to yield D-glucose or by comparison of the $[\alpha]_D$ or $[M]_D$ values with those of their aglycones except **13**.^{6,7)} Their molecular formulae were suggested from the accurate mass number of the $[M+H]^+$ or $[M+Na]^+$ or $[M+K]^+$ ion peak in the high-resolution positive FAB-MS.

Glucoside **1** (C₁₆H₂₄O₇, an amorphous powder, $[\alpha]_D^{24}$ –44°) was obtained as one of the main components of the glycoside fraction, and showed $[M+H]^+$ and $[M-C_6H_{12}O_6+H]^+$ ion peaks at *m/z* 329 and 149 in the positive FAB-MS. The ¹H- and ¹³C-NMR spectral data of **1** (Tables 1, 2) revealed the presence of one 1,4-disubstituted ben-

zene ring, two hydroxymethyls, one *sec*-methyl and one methine, in addition to the β -glucopyranosyl moiety. From analysis of the heteronuclear multiplet-bond correlation (HMBC) spectral data (correlations: H-2/C-4, C-6, C-7; H-3/C-1, C-2, C-4, C-5, C-8; H-5/C-1, C-3, C-6, C-8; H-6/C-2, C-4, C-7; H₂-7/C-1, C-2, C-6, Glc C-1; H-8/C-3, C-4, C-5, C-9, C-10; H₂-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-7), the aglycone was clarified to be 9-hydroxycuminyl alcohol (*p*-mentha-1,3,5-triene-7,9-diol), and the location of the glucosyl group was indicated to be C-7. Enzymatic hydrolysis of **1** gave an aglycone (**1a**; C₁₀H₁₄O₂, an amorphous powder, $[\alpha]_D^{21}$ +13°), which showed a plus $[\alpha]_D$ value, and D-glucose. Since (8*S*)-*p*-mentha-1,3,5-trien-3-ol showed a minus optical rotation ($[\alpha]_D$ –14.7°),⁸⁾ the configuration at C-8 of **1** should be *R*. Then, **1** was characterized as (8*R*)-9-hydroxycuminyl β -D-glucopyranoside.

Glucoside **2** (C₁₆H₂₄O₈, an amorphous powder, $[\alpha]_D^{22}$ –31°) showed $[M+H]^+$ and $[M-C_6H_{10}O_5+H]^+$ ion peaks at *m/z* 345 and 183 in the positive FAB-MS. Enzymatic hydrolysis with β -glucosidase gave an aglycone (**2a**; C₁₀H₁₄O₃, an amorphous powder, $[\alpha]_D^{22}$ +12°) and D-glucose. Glucoside **2** was considered to be an 8-hydroxylated derivative of **1** by comparison of its ¹H- and ¹³C-NMR spectral data and glycosylation shift values with those of **1**. The result of the HMBC experiment (see experimental) also supported this conclusion. As the optical rotation of (8*S*)-8,9-dihydroxycuminyl alcohol was reported to be plus,⁹⁾ the configuration at C-8 of **2** should be *S*. Therefore, **2** was characterized as (8*S*)-8,9-dihydroxycuminyl β -D-glucopyranoside.

Glucoside **3** (C₁₆H₂₄O₇, an amorphous powder, $[\alpha]_D^{24}$ –40°) revealed $[M+Na]^+$ and $[M-C_6H_{12}O_6+H]^+$ ion peaks at *m/z* 351 and 149 in the positive FAB-MS, and the ¹H- and ¹³C-NMR spectral data (Tables 1, 2) showed the presence of one 1,4-disubstituted benzene ring, two *tert*-methyls, one hydroxymethyl and one hydroxylated quaternary carbon, in addition to the β -glucopyranosyl moiety. From the result of the HMBC experiment (see Experimental) of **3**, the aglycone was indicated to be 8-hydroxycuminyl alcohol, and the glucosyl group was located at C-7. Since the glucose was suggested to be D-form from its minus optical rotation,^{6,7)} **3** was characterized as 8-hydroxycuminyl β -D-glucopyranoside.

Glucosides **4** (C₁₆H₂₈O₇, an amorphous powder, $[\alpha]_D^{23}$ –20°), **5** (C₁₆H₂₈O₇, mp 215–217°C, $[\alpha]_D^{23}$ +89°) and **6** (C₁₆H₂₈O₇, an amorphous powder, $[\alpha]_D^{23}$ +126°) have the same molecular formulae, and their ¹H- and ¹³C-NMR spectral data (Tables 1, 2) showed the presence of one β -glucopyranosyl group, and one tri-substituted double bond, one *tert*-

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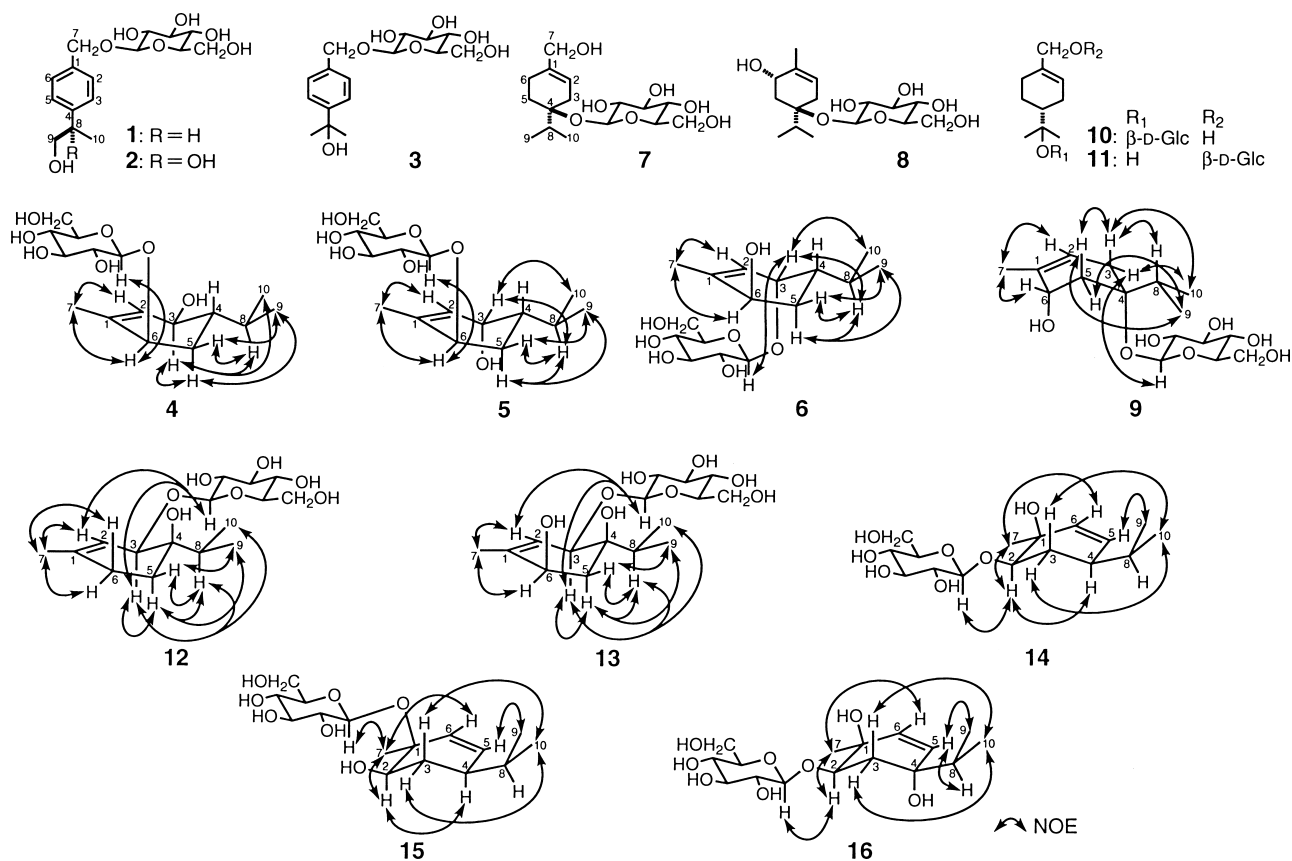


Fig. 1. Structures of 1–16, and NOE Interactions Observed in the NOESY Spectra of 4–6, 9 and 12–16

methyl, two *sec*-methyls, one methylene, two methines and two hydroxylated methines in the aglycone moiety, respectively. From analysis of their HMBC correlations (see Experimental), the aglycones were confirmed to be *p*-menth-1-ene-3,6-diol, and the location of the glucosyl groups was indicated to be C-6 for 4 and 5, and C-3 for 6. The observed nuclear Overhauser effect (NOE) interaction between H₃-9/H-5_{ax}, H-5_{eq} in their NOE spectroscopy (NOESY) spectra suggested that the configuration of their C-4 isopropyl was equatorial. The configuration of C-3 and C-6 hydroxyls of 4 was indicated to be equatorial and axial, respectively, by the NOE interaction between H-3/H-5_{ax} in its NOESY spectrum (Fig. 1), and by the coupling constant of H-3 and H-6 signals (H-3; brd, $J=10.0$ Hz, H-6; dd, $J=3.5, 3.5$ Hz) in the ¹H-NMR spectrum. Since the enzymatic hydrolysis of 4 gave an aglycone (4a; C₁₀H₁₈O₂, an amorphous powder, $[\alpha]_D^{23} +8^\circ$) and D-glucose, 4 was revealed to be 4_{ax}-*H-p*-menth-1-ene-3_{eq}6_{ax}-diol 6-*O*- β -D-glucopyranoside. The absolute configuration at C-6 of 4 was indicated to be *R* by the values of the ¹³C glycosylation shift $[\Delta\delta(\delta \text{ glycoside} - \delta \text{ aglycone})]$ of the α - and β -carbon and the chemical shift of the glucosyl anomeric carbon [C-6 (α -carbon): $\Delta\delta+6.6$, *R*-alcohols, about $\Delta\delta+4$ to $+7$, *S*-alcohols, about $\Delta\delta+9$ to $+11$; C-5 (β -carbon): $\Delta\delta-4.8$, β -pro-*S*-side carbon of *R*-alcohols, about $\Delta\delta-4$ to -5 , β -pro-*S*-side carbon of *S*-alcohols, about $\Delta\delta 0$ to -2 ; glucosyl C-1: $\delta 102.85$, *R*-alcohols, about $\delta 102$, *S*-alcohols, about $\delta 106$; (Table 2)].^{10–16} Thus, 4 was characterized as (3*S*,4*S*,6*R*)-*p*-menth-1-ene-3,6-diol 6-*O*- β -D-glucopyranoside. On the other hand, the configuration of the C-3 and C-6 hydroxyls of 5 was concluded to be axial by the narrow

H-3 and H-6 signals found as a double doublet (H-3; $J=3.0, 5.5$ Hz, H-6; $J=3.0, 3.0$ Hz) in their ¹H-NMR spectra. Enzymatic hydrolysis of 5 gave an aglycone (5a; C₁₀H₁₈O₂, an amorphous powder, $[\alpha]_D^{23} +240^\circ$) and D-glucose, and the absolute configuration at C-6 was indicated to be *R* in the same way described for 4 (Table 2). So, 5 was characterized as (3*R*,4*S*,6*R*)-*p*-menth-1-ene-3,6-diol 6-*O*- β -D-glucopyranoside. Furthermore, 6 was hydrolyzed with β -glucosidase and, from the hydrolyzed mixtures, 5a and D-glucose were obtained. As the position of the glucosyl unit was revealed to be C-3 from the HMBC experiment (see Experimental), 6 was characterized as (3*R*,4*S*,6*R*)-*p*-menth-1-ene-3,6-diol 3-*O*- β -D-glucopyranoside. Although 4 was reported as a constituent of *Eupatorium tinifolium*,¹⁷ the absolute structure was defined in this paper.

Glucosides 7 (C₁₆H₂₈O₇, an amorphous powder, $[\alpha]_D^{24} -16^\circ$) and 8 (C₁₆H₂₈O₇, an amorphous powder, $[\alpha]_D^{24} -29^\circ$) were identified as (4*S*)-*p*-menth-1-ene-4,7-diol 4-*O*- β -D-glucopyranoside and (4*R*,6*S*)-*p*-menth-1-ene-4,6-diol 4-*O*- β -D-glucopyranoside, which were isolated from the fruit of ajowan.¹⁸

Glucoside 9 (C₁₆H₂₈O₇, mp 78–81 °C, $[\alpha]_D^{21} -2^\circ$) showed $[M+Na]^+$ and $[M-C_6H_{12}O_6+H]^+$ ion peaks at m/z 355 and 153 in the positive FAB-MS, and enzymatic hydrolysis of 9 gave an aglycone (9a; C₁₀H₁₈O₂, an amorphous powder, $[\alpha]_D^{21} +40^\circ$) and D-glucose. It showed ¹H- and ¹³C-NMR spectral features similar to those of 8 (Tables 1, 2). From the analysis of HMBC spectral data (correlations: H-2/C-3, C-4, C-6, C-7; H-3_{ax}/C-1, C-2, C-5, C-8; H-3_{eq}/C-1, C-2, C-4, C-5, C-8; H₂-5/C-1, C-3, C-4, C-6, C-8; H-6_{eq}/C-1, C-2, C-4; H₃-7/C-1,

Table 1. ¹H-NMR Chemical Shifts of **1**—**16**, **1a**, **2a**, **4a**, **5a**, **9a**, **11a**, **12a**, **14a** and **16a** (in Pyridine-*d*₅, 500 MHz)

	1	1a	2	2a	3
H-2,6	7.50 d (8.0)	7.40 d (8.0)	7.57 d (8.0)	7.67 d (8.0)	7.57 d (8.0)
H-3,5	7.31 d (8.0)	7.59 d (8.0)	7.86 d (8.0)	7.94 d (8.0)	7.78 d (8.0)
H ₂ -7	4.84 d (11.5)	4.97 s (2H)	4.86 d (12.0)	5.00 s (2H)	4.87 d (12.0)
	5.18 d (11.5)	—	5.20 d (12.0)	—	5.21 d (12.0)
H-8	3.12 sextet (7.0)	3.15 sextet (7.0)	—	—	—
H ₂ -9	3.89 dd (7.0, 10.5)	3.92 dd (7.0, 10.0)	4.08 d (10.5)	4.10 d (11.0)	—
	4.00 dd (7.0, 10.5)	4.03 dd (7.0, 10.0)	4.13 d (10.5)	4.15 d (11.0)	—
H ₃ -9	—	—	—	—	1.73 s
H ₃ -10	1.39 d (7.0)	1.43 d (7.0)	1.86 s	1.89 s	1.73 s
Glc H-1	5.00 d (7.5)	—	5.02 d (7.5)	—	5.02 d (8.0)
	4	4a	5	5a	
H-2	5.95 br s	5.93 br s	6.04 d (5.5)	6.02 d (5.5)	
H-3 _{ax}	4.22 br d (10.0)	4.27 br d (9.5)	—	—	
H-3 _{eq}	—	—	4.31 dd (3.0, 5.5)	4.29 dd (3.0, 5.5)	
H-4 _{ax}	2.28 dddd (3.0, 3.0, 10.0, 13.5)	2.26 dddd (3.0, 3.0, 9.5, 13.0)	1.79 dddd (3.0, 3.0, 8.0, 13.5)	1.86 dddd (3.0, 3.0, 9.0, 13.0)	
H-5 _{ax}	1.31 ddd (3.5, 13.5, 13.5)	1.55 ddd (3.0, 13.0, 13.0)	1.89 ddd (3.0, 13.5, 13.5)	2.12 ddd (4.5, 13.0, 13.0)	
H-5 _{eq}	2.26 ddd (3.0, 3.5, 13.5)	2.05 ddd (3.0, 3.0, 13.0)	2.40 ddd (3.0, 3.0, 13.5)	2.19 ddd (3.0, 3.0, 13.0)	
H-6 _{eq}	4.37 dd (3.5, 3.5)	4.28 dd (3.5, 3.5)	4.40 dd (3.0, 3.0)	4.43 dd (3.0, 4.5)	
H ₃ -7	1.95 s	1.99 s	1.97 s	2.01 s	
H-8	2.50 dq (3.0, 7.0)	2.54 dq (3.0, 7.0)	2.05 dq (6.5, 8.0)	2.09 dq (6.5, 9.0)	
H ₃ -9	1.11 d (7.0)	1.03 d (7.0)	1.15 d (6.5)	1.19 d (6.5)	
H ₃ -10	0.91 d (7.0)	0.91 d (7.0)	1.09 d (6.5)	1.08 d (6.5)	
Glc H-1	4.94 d (7.5)	—	4.96 d (8.0)	—	
	6	7	8	9	
H-2	6.32 d (5.5)	5.88 br s	5.51 br s	5.49 br s	
H-3 _{ax}	—	2.36 br d (18.0)	2.30 br d (18.0)	2.32 dd (2.0, 18.0)	
H-3 _{eq}	4.38 dd (3.0, 5.5)	2.61 br d (18.0)	2.43 br d (18.0)	2.60 dd (2.0, 18.0)	
H-4 _{ax}	1.89 dddd (3.0, 3.0, 7.0, 13.0)	—	—	—	
H-5 _{ax}	2.10 ddd (3.0, 13.0, 13.0)	1.75 ddd (5.5, 8.5, 13.5)	2.01 dd (9.0, 13.0)	1.82 dd (5.0, 14.0)	
H-5 _{eq}	2.16 ddd (3.0, 3.0, 13.0)	2.08 ddd (5.5, 5.5, 13.5)	2.68 dd (5.0, 13.0)	2.45 ddd (2.0, 2.0, 14.0)	
H-6 _{ax}	—	2.85 ddd (5.5, 8.5, 18.0)	5.30 dd (5.0, 9.0)	—	
H-6 _{eq}	4.26 dd (3.0, 3.0)	2.21 ddd (5.5, 5.5, 18.0)	—	4.17 dd (2.0, 5.0)	
H ₂ -7	—	4.30 br s	—	—	
H ₃ -7	1.87 s	—	2.02 s	1.99 s	
H-8	2.28 dq (7.0, 7.0)	2.14 septet (7.0)	2.24 septet (6.5)	2.22 septet (7.0)	
H ₃ -9	1.00 d (7.0)	1.07 d (7.0)	1.00 d (6.5)	1.15 d (7.0)	
H ₃ -10	1.16 d (7.0)	1.05 d (7.0)	1.08 d (6.5)	0.89 d (7.0)	
Glc H-1	4.99 d (7.5)	5.03 d (7.5)	5.06 d (7.5)	4.94 d (7.5)	
	9a	10	11	11a	
H-2	5.50 br s	5.90 br d (3.0)	5.91 br d (3.0)	5.97 br d (3.0)	
H-3 _{ax}	2.19 dd (2.0, 17.5)	2.23 m	2.04 br dd (13.0, 17.0)	2.09 ddd (3.0, 13.0, 17.0)	
H-3 _{eq}	2.31 dd (2.0, 17.5)	2.23 m	2.27 ddd (3.0, 3.0, 17.0)	2.38 br d (17.0)	
H-4 _{ax}	—	1.88 dddd (2.0, 2.0, 13.0, 13.0)	1.69 dddd (3.0, 3.0, 13.0, 13.0)	1.77 dddd (3.0, 3.0, 13.0, 13.0)	
H-5 _{ax}	1.83 dd (4.5, 14.0)	1.32 br ddd (5.0, 13.0, 13.0)	1.40 br ddd (4.0, 13.0, 13.0)	1.44 br ddd (5.0, 13.0, 13.0)	
H-5 _{eq}	2.25 ddd (2.0, 3.0, 14.0)	2.27 br d (13.0)	2.12 br d (13.0)	2.21 br d (13.0)	
H-6 _{ax}	—	1.95 br dd (13.0, 17.0)	2.11 br dd (13.0, 16.0)	2.23 br dd (13.0, 17.0)	
H-6 _{eq}	4.27 br s	2.31 ddd (2.0, 5.0, 17.0)	2.40 br dd (4.0, 16.0)	2.34 ddd (3.0, 3.0, 17.0)	
H ₂ -7	—	4.26 d (8.5)	4.28 d (12.0)	4.31 d (13.5)	
	—	4.28 d (8.5)	4.50 d (12.0)	4.34 d (13.5)	
H ₃ -7	1.99 s	—	—	—	
H-8	1.77 septet (7.0)	—	—	—	
H ₃ -9	1.07 d (7.0)	1.37 ^{a)} s	1.30 ^{a)} s	1.33 ^{a)} s	
H ₃ -10	1.02 d (7.0)	1.39 ^{a)} s	1.31 ^{a)} s	1.35 ^{a)} s	
Glc H-1	—	5.03 d (8.0)	4.92 d (8.0)	—	

Table 1. (Continued)

	12	12a	13
H-2	5.84 brs	5.66 brs	5.89 brs
H-3 _{ax}	4.68 brs	4.45 brs	4.68 brs
H-5 _{ax}	1.57 ddd (5.5, 8.0, 13.0)	1.62 ddd (5.5, 8.0, 13.5)	1.90 dd (4.5, 14.0)
H-5 _{eq}	1.93 ddd (5.5, 5.5, 13.0)	1.98 ddd (5.5, 5.5, 13.5)	2.31 dd (4.5, 14.0)
H-6 _{ax}	2.26 ddd (5.5, 8.0, 17.5)	2.27 ddd (5.5, 8.0, 17.0)	—
H-6 _{eq}	1.83 ddd (5.5, 5.5, 17.5)	1.86 ddd (5.5, 5.5, 17.0)	4.15 dd (4.5, 4.5)
H ₃ -7	1.62 s	1.66 s	1.94 s
H-8	2.31 septet (7.0)	2.22 septet (7.0)	2.33 septet (7.0)
H ₃ -9	1.09 d (7.0)	1.15 d (7.0)	1.09 d (7.0)
H ₃ -10	1.06 d (7.0)	1.05 d (7.0)	1.07 d (7.0)
Glc H-1	5.17 d (8.0)	—	5.16 d (8.0)

	14	14a	15
H-2 _{ax}	4.45 dd (3.0, 7.5)	4.37 dd (3.0, 7.5)	4.41 dd (3.5, 8.0)
H-3 _{ax}	2.08 ddd (7.0, 7.5, 13.5)	2.03 ddd (6.5, 7.5, 13.5)	1.92 ddd (7.0, 8.0, 13.5)
H-3 _{eq}	2.16 ddd (3.0, 5.0, 13.5)	2.25 ddd (3.0, 7.0, 13.5)	2.25 ddd (3.5, 5.0, 13.5)
H-4 _{ax}	2.40 dddd (3.0, 5.0, 7.0, 7.0)	2.42 dddd (2.5, 6.5, 7.0, 7.0)	2.30 dddd (3.0, 5.0, 7.0, 7.0)
H-5	5.66 dd (3.0, 10.0)	5.78 dd (2.5, 10.0)	5.86 dd (3.0, 10.0)
H-6	5.92 d (10.0)	6.00 d (10.0)	6.15 d (10.0)
H ₃ -7	1.67 s	1.75 s	1.75 s
H-8	1.58 octet (7.0)	1.64 octet (7.0)	1.64 octet (7.0)
H ₃ -9	0.86 d (7.0)	0.89 d (7.0)	0.88 d (7.0)
H ₃ -10	0.85 d (7.0)	0.88 d (7.0)	0.88 d (7.0)
Glc H-1	5.04 d (7.5)	—	5.17 d (8.0)

	16	16a
H-2 _{ax}	4.57 dd (2.5, 6.0)	4.45 dd (3.0, 7.0)
H-3 _{ax}	2.30 dd (6.0, 14.5)	2.20 dd (7.0, 13.5)
H-3 _{eq}	2.69 dd (2.5, 14.5)	2.72 dd (3.0, 13.5)
H-5	5.94 d (10.0)	5.99 d (10.0)
H-6	5.96 d (10.0)	6.02 d (10.0)
H ₃ -7	1.71 s	1.75 s
H-8	1.95 septet (7.0)	1.96 septet (7.0)
H ₃ -9	1.09 d (7.0)	1.07 d (7.0)
H ₃ -10	1.16 d (7.0)	1.15 d (7.0)
Glc H-1	5.16 d (8.0)	—

δ in ppm from TMS [coupling constants (*J*) in Hz are given in parentheses]. a) Assignments may be interchanged in each column.

C-2, C-6; H-8/C-3, C-4, C-5, C-9, C-10; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-4), **9** was clarified to be a β -D-glucopyranoside of *p*-menth-1-ene-4,6-diol, and the position of the glucosyl unit was C-4. The observed NOE interaction between H₃-9/H-5_{ax}, H-5_{eq} and between H₃-10/H-3_{ax}, H-3_{eq} in the NOESY spectrum suggested that the configuration of the C-4 isopropyl was equatorial, and the H-6 signal which was found to be a narrow double doublet (*J*=2.0, 5.5 Hz) in the ¹H-NMR spectrum suggested that the configuration of the C-6 hydroxyl was axial. The absolute structure of the aglycone moiety of **9** was examined by comparison of its glycosylation shift values (C-3, C-4, C-5 and C-8) with those of **7** and **8**.¹⁸ As the glycosylation shift values of **9** showed an obvious difference between the values of **7** and **8**, which have a 4*R* configuration [C-3 (**7** and **8**; -4.0 and -4.5, **9**; -5.4), C-4 (**7** and **8**; +8.4 and +8.6, **9**; +7.6), C-5 (**7** and **8**; -2.4 and -1.8, **9**; -2.8), C-8 (**7** and **8**; -3.4 and -3.9, **9**; -1.1)], the absolute configuration at C-4 of **9** was concluded to be *S*. So, **9** was characterized as (4*S*,6*S*)-*p*-menth-1-ene-4,6-diol 4-*O*- β -D-glucopyranoside.

Glucoside **10** (C₁₆H₂₈O₇, an amorphous powder, [α]_D²²

+8°) was identified as (4*R*)-*p*-menth-1-ene-7,8-diol 8-*O*- β -D-glucopyranoside, which was isolated from fennel.¹⁹⁾

Glucoside **11** (C₁₆H₂₈O₇, an amorphous powder, [α]_D²⁴ -6°) revealed [M+H]⁺ and [M-C₆H₁₂O₆+H]⁺ ion peaks at *m/z* 333 and 153 in the positive FAB-MS, and showed similar ¹H- and ¹³C-NMR spectral data with those of **10** (Tables 1, 2). Enzymatic hydrolysis of **11** gave an aglycone (**11a**; C₁₀H₁₈O₂, an amorphous powder, [α]_D²⁴ +66°), which was identical with the aglycone of **10**, as well as D-glucose. So, **11** was indicated to be a β -D-glucopyranoside of (4*R*)-*p*-menth-1-ene-7,8-diol. Since the position of the glucosyl unit was proved to be C-7 from the cross-peak between the glucosyl H-1/C-7 in the HMBC spectrum (see Experimental), **11** was characterized as (4*R*)-*p*-menth-1-ene-7,8-diol 7-*O*- β -D-glucopyranoside.

Glucoside **12** (C₁₆H₂₈O₇, an amorphous powder, [α]_D²¹ -86°) showed [M+H]⁺ and [M-C₆H₁₂O₆+H]⁺ ion peaks at *m/z* 333 and 153 in the positive FAB-MS, and was hydrolyzed with β -glucosidase to an aglycone (**12a**; C₁₀H₁₈O₂, an amorphous powder, [α]_D²¹ -29°) and D-glucose. The ¹H- and ¹³C-NMR spectral data of **12** (Tables 1, 2) showed the

Table 2. ^{13}C -NMR Chemical Shifts of **1**–**16**, **1a**, **2a**, **4a**, **5a**, **9a**, **11a**, **12a**, **14a** and **16a** (in Pyridine- d_5 , 125 MHz)

	1	1a	2	2a	3	4	4a	5	6
C-1	136.64 (−4.7)	141.38	136.80 (−4.8)	141.63	136.54	134.15 (−2.7)	136.83	135.97 (−2.5)	138.61
C-2	128.58	127.98	128.04	126.83	128.16	133.16	131.20	129.96	127.43 (−0.6)
C-3	128.02	127.26	126.20	126.24	125.20	68.88	69.05	64.30	74.94 (+10.5)
C-4	145.10	144.29	147.77	147.04	150.92	42.33	42.74	40.56	41.04 (+0.0)
C-5	128.02	127.26	126.20	126.24	125.20	26.36 (−4.8)	31.13	26.45 (−5.0)	31.87
C-6	128.58	127.98	128.04	126.83	128.16	74.10 (+6.6)	67.50	74.71 (+6.5)	67.99
C-7	71.02 (+6.7)	64.28	70.93 (+6.7)	64.28	70.89	20.79	21.06	21.24	21.35
C-8	43.17	43.17	74.80	74.83	71.40	26.52	26.56	28.58	27.52
C-9	68.50	68.54	71.74	72.07	32.61	21.31	21.37	21.19	21.24
C-10	18.51	18.47	26.88	26.93	32.61	17.14	17.27	21.26	21.98
Glc-1	104.03		104.08		104.03	102.85		102.49	106.14
Glc-2	75.32		75.31		75.31	75.03		75.09	75.71
Glc-3	78.60		78.60		78.60	78.73		78.60	78.71
Glc-4	71.79		71.74		71.74	71.99		72.02	71.94
Glc-5	78.63		78.65		78.65	78.46		78.37	78.28
Glc-6	62.89		62.85		62.84	63.09		63.07	63.04
	5a	7	8	9	9a	10	11	11a	12
C-1	138.50	139.32	139.14	137.66	135.86	139.26	135.14 (−4.0)	139.15	138.57
C-2	128.06	119.17	120.18	120.12	121.65	121.33	125.13	121.59	121.09 (−4.1)
C-3	64.46	31.30 (−4.0)	31.02	30.70 (−5.4)	36.14	27.14	27.24	27.24	76.04 (+8.1)
C-4	41.01	79.75 (+8.4)	82.44	80.21 (+7.6)	72.64	44.59 (−1.3)	45.85	46.17	72.94 (−0.1)
C-5	31.40	29.41 (−2.4)	40.70	36.64 (−2.8)	39.40	23.96	24.16	24.34	26.89
C-6	68.18	23.91	67.95	68.04	68.72	27.12	27.33	27.31	28.21
C-7	21.35	66.23	19.90	21.14	21.19	66.55	73.15 (+6.6)	66.59	23.21
C-8	28.60	34.04 (−3.4)	35.04	36.50 (−1.1)	37.64	79.30 (+8.1)	71.14	71.21	32.14
C-9	21.20	17.72	18.15	18.57	17.14	23.71 ^{a)} (−3.3)	27.03 ^{a)}	27.01 ^{a)}	16.84
C-10	21.48	17.32	17.28	17.50	17.27	24.34 ^{a)} (−3.7)	27.90 ^{a)}	28.01 ^{a)}	18.20
Glc-1		99.12	98.81	99.23		98.63	103.54		102.35
Glc-2		75.59	75.59	75.28		75.43	75.30		75.26
Glc-3		78.83	78.75	78.95		78.96	78.67		78.74
Glc-4		72.09	71.97	72.23		71.94	71.80		71.86
Glc-5		78.12	77.94	78.23		78.15	78.54		78.49
Glc-6		63.17	63.06	63.12		63.05	62.89		62.91
	12a	13	14	15	14a	16	16a		
C-1	136.46	140.29	70.05 (−0.5)	79.28 (+8.7)	70.57	69.21 (−1.4)	70.63		
C-2	125.21	122.30	81.30 (+7.8)	70.99 (−2.6)	73.53	80.98 (+6.5)	74.52		
C-3	67.97	76.63	27.15 (−3.2)	29.71	30.35	32.87 (−3.1)	35.92		
C-4	72.99	75.09	39.18	39.64	38.92	71.69	72.78		
C-5	26.91	34.82	130.45	133.94	130.62	134.61	133.77		
C-6	28.09	68.32	134.16	131.12 (−3.5)	134.61	133.07	134.20		
C-7	23.18	20.47	25.85	23.25 (−2.1)	25.36	26.42	25.63		
C-8	32.58	32.37	32.22	32.55	32.22	38.00	37.82		
C-9	17.11	16.74	20.14	20.28	20.04	17.92	17.83		
C-10	18.35	18.24	20.18	20.28	20.14	17.13	17.11		
Glc-1		102.78	103.35	99.30		102.28			
Glc-2		75.19	75.00	75.54		75.05			
Glc-3		78.71	78.59	78.82		78.69			
Glc-4		71.84	72.05	72.08		72.12			
Glc-5		78.55	78.49	78.41		78.69			
Glc-6		62.91	62.97	63.03		62.98			

δ in ppm from TMS. $\Delta\delta(\delta \text{ glucoside} - \delta \text{ aglycone})$ are given in parentheses. *a*) Assignments may be interchanged in each column.

presence of one *tert*-methyl, two *sec*-methyls, two methyl- enes, one methine, one hydroxylated methine, one hydroxyl- ated quaternary carbon and one trisubstituted double bond, in addition to the β -glucopyranosyl moiety. From analysis of HMBC spectral data (correlations: H-2/C-4, C-6, C-7; H- 3/C-1, C-2, Glc C-1; H-5_{ax}/C-1, C-3, C-4, C-6; H-5_{eq}/C-1, C- 3, C-4, C-6, C-8; H-6_{ax}/C-1, C-2, C-4, C-5; H-6_{eq}/C-1, C-2, C-4, C-5, C-7; H₂-7/C-1, C-2, C-6; H-8/C-3, C-4, C-5, C-9,

C-10; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C- 3), the aglycone was suggested to be *p*-menth-1-ene-3,4-diol, and the location of the glucosyl group was indicated to be C- 3. Since the NOE interactions between H₃-9/H-5_{ax}, H-5_{eq}, be- tween H₃-10/H-3_{ax}, and between H-3_{ax}/H-5_{ax} were observed in the NOESY spectrum of **12**, the configurations of C-3 and C-4 hydroxyls were concluded to be equatorial and axial, re- spectively. So, **12** was confirmed to be a 3-*O*- β -D-glucopyra-

noside of *p*-menth-1-ene-3_{eq},4_{ax}-diol. As the glycosylation shift values of the α - and β -carbon [C-2 (β -pro-*S*); -4.1, C-3 (α); +8.1, C-4 (β -pro-*R*); -0.1] and the chemical shift of the anomeric carbon (δ 102.35) were in agreement with the *R* configuration,^{10–16} the absolute configuration at C-3 was indicated to be *R*. Then, **12** was characterized as (3*R*,4*R*)-*p*-menth-1-ene-3,4-diol 3-*O*- β -D-glucopyranoside.

Glucoside **13** (C₁₆H₂₈O₈, an amorphous powder, [α]_D²² -97°) had one *tert*-methyl, two *sec*-methyls, one methylene, one methine, two hydroxylated methines, one hydroxylated quaternary carbon and one trisubstituted double bond, in addition to the β -glucopyranosyl moiety. By comparison of its ¹H- and ¹³C-NMR spectral data with those of **12** (Tables 1, 2), and from the analysis of HMBC correlations (see Experimental), **13** was clarified to be β -glucopyranoside of *p*-menth-1-ene-3,4,6-triol, and the location of the glucosyl group was indicated to be C-3. The NOE interaction which was observed between H₃-9/H-5_{ax}, H-5_{eq}, and between H-3_{ax}/H-5_{ax} in the NOESY spectrum of **13** suggested that the configurations of C-3 and C-4 hydroxyls were equatorial and axial, respectively. The configuration of C-6 hydroxyl was concluded to be axial by an H-6 narrow double doublet signal (*J*=4.5, 4.5 Hz) in its ¹H-NMR spectrum. So, **13** was confirmed to be *p*-menth-1-ene-3_{eq},4_{ax},6_{ax}-triol 3-*O*- β -D-glucopyranoside. Since this glucose was considered to be the *D*-form, the same as the other glucoses, and the chemical shifts of C-3 and Glc C-1 revealed almost identical values to those of **12** [C-3 (**12**; δ 76.04, **13**; δ 76.63), glc C-1 (**12**; δ 102.35, **13**; δ 102.78)], the absolute configuration at C-3 was deduced to be *R*, the same as **12**. Consequently, **13** was concluded to be (3*R*,4*R*,6*R*)-*p*-menth-1-ene-3,4,6-triol 3-*O*- β -D-glucopyranoside.

Glucosides **14** (C₁₆H₂₈O₇, an amorphous powder, [α]_D²³ +8°) and **15** (C₁₆H₂₈O₇, an amorphous powder, [α]_D²¹ -23°) showed the presence of one disubstituted double bond, one *tert*-methyl, two *sec*-methyls, one methylene, two methines, one hydroxylated methine, one hydroxylated quaternary carbon and a β -glucopyranosyl group in their ¹H- and ¹³C-NMR spectra (Tables 1, 2). From analysis of their HMBC correlations (see Experimental), the aglycone of these glucosides was indicated to be *p*-menth-5-ene-1,2-diol, and the location of the glucosyl group was concluded to be C-2 for **14**, and C-1 for **15**. The observed NOE interaction between H₃-9/H-5_{ax}, H-5_{eq} in their NOESY spectra suggested that the configuration of their C-4 isopropyl was equatorial. The relative configurations of C-1 and C-2 hydroxyls of **14** and **15** were indicated to be axial and equatorial by the NOE interaction between H-2/H-4 and H-2/H₃-7 in their NOESY spectra (Fig. 1). Enzymatic hydrolysis of **14** and **15** gave the same aglycone (**14a**; C₁₀H₁₈O₂, an amorphous powder, [α]_D²² +24°) and *D*-glucose, thus they were suggested to be β -D-glucopyranosides of *p*-menth-5-ene-1_{ax},2_{eq}-diol. The absolute configuration at the C-2 of **14** was indicated to be *R* by the values of the glycosylation shift of the α - and β -carbon and the chemical shift of the glucosyl anomeric carbon, as shown in Table 2. Thus, **14** and **15** were characterized as (1*S*,2*R*,4*R*)-*p*-menth-5-ene-1,2-diol 2-*O*- β -D-glucopyranoside and (1*S*,2*R*,4*R*)-*p*-menth-5-ene-1,2-diol 1-*O*- β -D-glucopyranoside, respectively.

Glucoside **16** (C₁₆H₂₈O₈, an amorphous powder, [α]_D²³ -60°) showed [M+H]⁺ and [M-C₆H₁₂O₆+H]⁺ ion peaks at

m/z 349 and 169 in the positive FAB-MS, and was hydrolyzed with β -glucosidase to an aglycone (**16a**; C₁₀H₁₈O₃, an amorphous powder, [α]_D²³ -50°) and *D*-glucose. By comparison of its ¹H- and ¹³C-NMR spectral data with those of **14** and **15** (Tables 1, 2), **16** was suggested to be a mono-hydroxyl derivative of *p*-menth-5-ene-1,2-diol β -D-glucopyranoside. The locations of the additional hydroxyl group and the glucosyl group were indicated to be C-4 and C-2 from analysis of the HMBC spectral data (see Experimental). The NOE interaction observed between H₃-10/H-3_{ax}, H-3_{eq} in the NOESY spectrum suggested that the configuration of the C-4 isopropyl was equatorial. From the coupling constant of H-2 of **16a** [**16a**; H-2: δ 4.45 (*J*=3.0, 7.0 Hz), **14a**; H-2: δ 4.37 (*J*=3.0, 7.5 Hz)] and the observed NOE interaction between H-2/H₃-7, **14** and **16** were estimated to have the same relative configurations at C-1 and C-2. The absolute configuration at C-2 of **16** was indicated to be *R* by the values of the glycosylation shift of the α - and β -carbon and the chemical shift of the glucosyl anomeric carbon (Table 2). Consequently, **16** was considered to be (1*S*,2*R*,4*S*)-*p*-menth-5-ene-1,2,4-triol 2-*O*- β -D-glucopyranoside.

The ingredient relationship between the essential oil and the water-soluble constituent was confirmed by the isolation of these monoterpenoid glucosides (**1** to **3**), which showed a biosynthetic relation to cuminaldehyde.

Experimental

Melting points were determined on a Yanagimoto micromelting point apparatus and are uncorrected. Optical rotations were measured on a JASCO DIP-370 digital polarimeter. FAB-MS were recorded with a JEOL HX-110 spectrometer using glycerol as a matrix. ¹H- and ¹³C-NMR spectra were taken on a JEOL A-500 spectrometer with tetramethylsilane as an internal standard, and chemical shifts were recorded in δ values. ¹H-¹³C COSY, HMBC, NOESY and 1-D NOESY spectra were obtained with the usual pulse sequence, and data processing was performed with standard JEOL software. Column chromatography (C. C.) was carried out under TLC monitoring using Kieselgel 60 (70–230 mesh, Merck), Sephadex LH-20 (25–100 μ m, Pharmacia), Lobar RP-8 column (Merck) and Amberlite XAD-II (Organo). TLC was performed on silica gel (Merck 5721) and spots were detected with *p*-anisaldehyde-H₂SO₄ reagent. HPLC separation was carried out with Symmetryprep C₁₈ 7 μ m [Waters; column size, 7.8×300 mm; ODS], and carbohydrate analysis [Waters; column size, 3.9×300 mm; CHA] columns. Acetylation was done in the usual way using Ac₂O and pyridine. No acetoxy group had been detected by NMR spectral analysis of the materials prior to acetylation.

Extraction and Separation Commercial cumin (the fruit of *Cuminum Cuminum* L.; purchased from Asaoka Spices Ltd., Lot. No. 99012001; 2.0 kg) was extracted with 70% methanol (101×4), and the extract (281.1 g) was partitioned into ether-water and ethyl acetate-water, respectively. The aqueous portion (152.3 g) was chromatographed over Amberlite XAD-II (H₂O→MeOH) to give water eluate (87.1 g) and methanol eluate (65.2 g) fractions.

The methanol fraction was subjected to Sephadex LH-20 [MeOH-H₂O (9:1)] to give four fractions (frs. A–D). Fraction B (17.95 g) was chromatographed over silica gel [CHCl₃-MeOH-H₂O (17:3:0.2→4:1:0.1→7:3:0.5)→MeOH] to give thirteen fractions (frs. B₁–B₁₃). Fraction B₃ (0.50 g) was passed through a Lobar RP-8 column [MeCN-H₂O (3:17–3:7)] to give fifteen fractions (frs. B_{3,1}–B_{3,15}), and fr. B_{3,10} was subjected to Sephadex LH-20 (MeOH) and HPLC [ODS, MeOH-H₂O (1:1)] to give **9** (17 mg). Fraction B₄ (2.00 g) was passed through a Lobar RP-8 column [MeCN-H₂O (3:17)] to give eleven fractions (frs. B_{4,1}–B_{4,11}), and fr. B_{4,3} was subjected to HPLC [ODS, MeCN-H₂O (3:37) and CHA (MeCN-H₂O (19:1)] to give **5** (15 mg) and **6** (7 mg). Fraction B_{4,4} was passed through Sephadex LH-20 (MeOH) and HPLC [ODS, MeCN-H₂O (1:9) and CHA, MeCN-H₂O (24:1)] to give **8** (13 mg). Fraction B_{4,7} was subjected to HPLC [ODS, MeCN-H₂O (3:17)] to give **4** (20 mg). Fraction B_{4,10} was subjected to HPLC [ODS, MeCN-H₂O (3:17)] to give **7** (151 mg) and **14** (58 mg), and fr. B_{4,10} was subjected to HPLC [ODS, MeCN-H₂O (1:9) and CHA, MeCN-H₂O (24:1)] to give **12** (14 mg). Fraction B₅ (2.00 g) was passed

through a Lobar RP-8 column [MeCN–H₂O (3 : 17)] to give twelve fractions (frs. B₅₋₁–B₅₋₁₂), and fr. B₅₋₆ was subjected to HPLC [CHA, MeCN–H₂O (19 : 1)] to give **3** (14 mg). Fraction B₅₋₇ was passed through Sephadex LH-20 (MeOH) to give fr. B_{5-7a}, B_{5-7b} and **10** (15 mg). Fraction B_{5-7a} was subjected to silica gel column chromatography [CHCl₃–MeOH–H₂O (15 : 5 : 0.4→7 : 3 : 0.5)] to give **1** (293 mg). Fraction B_{5-7b} was acetylated with Ac₂O and pyridine, and the acetylated fraction was subjected to HPLC [ODS, MeCN–H₂O (7 : 3)] to give three fractions. The first fraction was deacetylated by heating in a water bath with 5% NH₄OH–MeOH for 2 h, then passed through a Sephadex LH-20 (MeOH) to give **11** (18 mg). Fraction B₅₋₈ was subjected to HPLC [ODS, MeCN–H₂O (1 : 9) and CHA, MeCN–H₂O (19 : 1)] to give **13** (3 mg). Fraction B₅₋₉ was subjected to HPLC [ODS, MeCN–H₂O (3 : 17) and CHA, MeCN–H₂O (19 : 1)] to give **15** (118 mg). Fraction B₇ (0.82 g) was passed through a Lobar RP-8 column [MeCN–H₂O (3 : 17)] to give seven fractions (frs. B₇₋₁–B₇₋₇), and fr. B₇₋₃ was subjected to HPLC [CHA, MeCN–H₂O (14 : 1)] to give **16** (39 mg). Fraction B₉ (2.40 g) was passed through a Lobar RP-8 column [MeCN–H₂O (3 : 17)] to give nine fractions (frs. B₉₋₁–B₉₋₉), and fr. B₉₋₄ was subjected to HPLC [CHA, MeCN–H₂O (19 : 1)] to give **2** (12 mg).

The following compounds were identified by comparison with authentic compounds: (4*S*)-*p*-menth-1-ene-4,7-diol 4-*O*-β-*D*-glucopyranoside (**7**), (4*R*,6*S*)-*p*-menth-1-ene-4,6-diol 4-*O*-β-*D*-glucopyranoside (**8**) and (4*R*)-*p*-menth-1-ene-7,8-diol 7-*O*-β-*D*-glucopyranoside (**10**).

(8*R*)-9-Hydroxycuminyll β-*D*-Glucopyranoside (1) An amorphous powder, [α]_D²⁴ –44° (*c*=1.5, MeOH). Positive FAB-MS *m/z*: 657 [2M+H]⁺, 367 [M+K]⁺, 351.1428 [M+Na]⁺ (Calcd for C₁₆H₂₄NaO₇; 351.1420), 329 [M+H]⁺, 311 [M–H₂O+H]⁺, 149 [M–C₆H₁₂O₆+H]⁺ (base). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

Enzymatic Hydrolysis of 1 A mixture of **1** (13 mg) and hesperidinase (5 mg, ICN Biomedicals, Inc., Lot 72635) in water (5 ml) was shaken in a water bath at 37 °C for 20 d. The mixture was concentrated *in vacuo* to dryness and the residue was chromatographed over silica gel [CHCl₃–MeOH (9 : 1) and CHCl₃–MeOH–H₂O (7 : 3 : 0.5)] to afford an aglycone **1a** (2 mg) and a sugar fraction. The sugar fraction was passed through Sephadex LH-20 (MeOH) to give a syrup, and HPLC [carbohydrate analysis (Waters), detector; JASCO RI-930 detector and JASCO OR-990 chiral detector, solv.; MeCN–H₂O (17 : 3), 2 ml/min; *t*_R 4.50 min (same location as that of *D*-glucose)] showed the presence of *D*-glucose.

(8*R*)-9-Hydroxycuminyll Alcohol (1a) An amorphous powder, [α]_D²¹ +13° (*c*=0.3, CHCl₃). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

(8*S*)-8,9-Dihydroxycuminyll β-*D*-Glucopyranoside (2) An amorphous powder, [α]_D²² –31° (*c*=0.4, MeOH). Positive FAB-MS *m/z*: 345.1548 [M+H]⁺ (Calcd for C₁₆H₂₅O₇; 345.1550), 327 [M–H₂O+H]⁺, 183 [M–C₆H₁₀O₅+H]⁺ (base), 165 [M–C₆H₁₂O₆+H]⁺. ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-3, C-4, C-6, C-7; H-3/C-1, C-2, C-5, C-8; H-5/C-1, C-3, C-6, C-8; H-6/C-2, C-4, C-5, C-7; H₂-7/C-1, C-2, C-6, Glc C-1; H₂-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-7.

Enzymatic Hydrolysis of 2 A mixture of **2** (4 mg) and β-glucosidase (5 mg) in water (5 ml) was shaken in a water bath at 37 °C for 7 d. The mixture was treated in the same way as described for **1** to afford an aglycone **2a** (1 mg) and a sugar fraction. From the sugar fraction, *D*-glucose was detected as described for **1**.

(8*S*)-8,9-Dihydroxycuminyll Alcohol (2a) An amorphous powder, [α]_D²² +12° (*c*=0.1, MeOH). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

8-Hydroxycuminyll β-*D*-Glucopyranoside (3) An amorphous powder, [α]_D²⁴ –40° (*c*=0.5, MeOH). Positive FAB-MS *m/z*: 351.1418 [M+Na]⁺ (Calcd for C₁₆H₂₄NaO₇; 351.1420), 311 [M–H₂O+H]⁺, 149 [M–C₆H₁₂O₆+H]⁺ (base). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-3, C-4, C-6, C-7; H-3/C-1, C-2, C-5, C-8; H-5/C-1, C-3, C-6, C-8; H-6/C-2, C-4, C-5, C-7; H₂-7/C-1, C-2, C-6, Glc C-1; H₃-10/C-4, C-8, C-9; Glc H-1/C-7.

(3*S*,4*S*,6*R*)-*p*-Menth-1-ene-3,6-diol 6-*O*-β-*D*-Glucopyranoside (4) An amorphous powder, [α]_D²³ –20° (*c*=0.5, MeOH). Positive FAB-MS *m/z*: 665 [2M+H]⁺, 371.1488 [M+K]⁺ (base, Calcd for C₁₆H₂₈KO₇; 371.1472), 355.1717 [M+Na]⁺ (Calcd for C₁₆H₂₈NaO₇; 355.1733), 315 [M–H₂O+H]⁺, 153 [M–C₆H₁₂O₆+H]⁺. ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-4, C-6, C-7; H-4/C-3, C-5, C-6, C-8, C-9, C-10; H-5_{ax}/C-3, C-4; H-5_{eq}/C-1, C-2, C-3, C-4, C-8; H-6/C-1, C-2, C-4, Glc C-1; H₃-7/C-1, C-2, C-6; H-8/C-3, C-4, C-5, C-9, C-10; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9;

Glc H-1/C-6.

Enzymatic Hydrolysis of 4 A mixture of **4** (7 mg) and β-glucosidase in water (5 ml) was shaken in a water bath at 37 °C for 7 d. The mixture was treated in the same way as described for **1** to afford an aglycone **4a** (3 mg) and a sugar fraction. From the sugar fraction, *D*-glucose was detected as described for **1**.

(3*S*,4*S*,6*R*)-*p*-Menth-1-ene-3,6-diol (4a) An amorphous powder, [α]_D²³ +8° (*c*=0.2, MeOH). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

(3*R*,4*S*,6*R*)-*p*-Menth-1-ene-3,6-diol 6-*O*-β-*D*-Glucopyranoside (5) Colorless needles (MeOH), mp 215–217 °C, [α]_D²³ +89° (*c*=0.5, MeOH). Positive FAB-MS *m/z*: 665 [2M+H]⁺, 371 [M+K]⁺, 355 [M+Na]⁺, 333.1897 [M+H]⁺ (base, Calcd for C₁₆H₂₆O₇; 333.1913), 315 [M–H₂O+H]⁺, 153 [M–C₆H₁₂O₆+H]⁺. ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-3, C-4, C-6, C-7; H-3/C-5; H-4/C-3, C-5, C-6, C-8, C-9, C-10; H-5_{ax}/C-3, C-4, C-8; H-5_{eq}/C-1, C-3, C-4, C-6; H-6/C-1, C-2, C-4, C-7, Glc C-1; H₃-7/C-1, C-2, C-6; H-8/C-4, C-9, C-10; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-6.

Enzymatic Hydrolysis of 5 A mixture of **5** (7 mg) and β-glucosidase (5 mg) in water (5 ml) was shaken in a water bath at 37 °C for 7 d. The mixture was treated in the same way as described for **1** to afford an aglycone **5a** (2 mg) and a sugar fraction. From the sugar fraction, *D*-glucose was detected as described for **1**.

(3*R*,4*S*,6*R*)-*p*-Menth-1-ene-3,6-diol (5a) An amorphous powder, [α]_D²³ +240° (*c*=0.1, MeOH). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

(3*R*,4*S*,6*R*)-*p*-Menth-1-ene-3,6-diol 3-*O*-β-*D*-Glucopyranoside (6) Colorless needles (MeOH), mp 215–217 °C, [α]_D²³ +126° (*c*=0.2, MeOH). Positive FAB-MS *m/z*: 371 [M+K]⁺, 333.1930 [M+H]⁺ (Calcd for C₁₆H₂₆O₇; 333.1913), 153 [M–C₆H₁₂O₆+H]⁺ (base). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-3, C-4, C-6, C-7; H-3/C-1, C-2, Glc C-1; H-5_{ax}/C-3, C-4; H-5_{eq}/C-1, C-3, C-4, C-6; H₃-7/C-1, C-2, C-6; H-8/C-4; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-3.

Enzymatic Hydrolysis of 6 A mixture of **6** (4 mg) and β-glucosidase (5 mg) in water (5 ml) was shaken in a water bath at 37 °C for 7 d. The mixture was treated in the same way as described for **1** to afford an aglycone **6a** (1 mg) and a sugar fraction. From the sugar fraction, *D*-glucose was detected as described for **1**.

(4*S*,6*S*)-*p*-Menth-1-ene-4,6-diol 4-*O*-β-*D*-Glucopyranoside (9) Colorless needles (MeOH), mp 78–81 °C, [α]_D²¹ –2° (*c*=0.7, MeOH). Positive FAB-MS *m/z*: 355.1724 [M+Na]⁺ (Calcd for C₁₆H₂₈NaO₇; 355.1733), 315 [M–H₂O+H]⁺, 153 [M–C₆H₁₂O₆+H]⁺ (base). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

Enzymatic Hydrolysis of 9 A mixture of **9** (6 mg) and β-glucosidase (5 mg) in water (5 ml) was shaken in a water bath at 37 °C for 7 d. The mixture was treated in the same way as described for **1** to afford an aglycone **9a** (2.5 mg) and a sugar fraction. From the sugar fraction, *D*-glucose was detected as described for **1**.

(4*S*,6*S*)-*p*-Menth-1-ene-4,6-diol (9a) An amorphous powder, [α]_D²¹ +40° (*c*=0.2, MeOH). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

(4*R*)-*p*-Menth-1-ene-7,8-diol 7-*O*-β-*D*-Glucopyranoside (11) An amorphous powder, [α]_D²⁴ –6° (*c*=0.3, MeOH). Positive FAB-MS *m/z*: 371 [M+K]⁺, 355.1735 [M+Na]⁺ (base, Calcd for C₁₆H₂₈NaO₇; 355.1733), 333 [M+H]⁺, 315 [M–H₂O+H]⁺, 153 [M–C₆H₁₂O₆+H]⁺. ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-4, C-6, C-7; H-3_{ax}/C-1, C-2; H-3_{eq}/C-1, C-2, C-4, C-5; H-4/C-3, C-5, C-6, C-8, C-9, C-10; H-5_{ax}/C-1, C-4, C-8; H-5_{eq}/C-1, C-3, C-4, C-6, C-8; H-6_{ax}/C-1, C-4, C-5; H-6_{eq}/C-1, C-2, C-4, C-5, C-7; H₂-7/C-1, C-2, C-6, Glc C-1; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-7.

Enzymatic Hydrolysis of 11 A mixture of **11** (6 mg) and β-glucosidase (5 mg) in water (5 ml) was shaken in a water bath at 37 °C for 7 d. The mixture was treated in the same way as described for **1** to afford an aglycone **11a** (1.7 mg) and a sugar fraction. From the sugar fraction, *D*-glucose was detected as described for **1**.

(4*R*)-*p*-Menth-1-ene-7,8-diol (11a) An amorphous powder, [α]_D²⁴ +66° (*c*=0.1, MeOH). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

(3*R*,4*R*)-*p*-Menth-1-ene-3,4-diol 3-*O*-β-*D*-Glucopyranoside (12) An amorphous powder, [α]_D²¹ –86° (*c*=0.6, MeOH). Positive FAB-MS *m/z*: 665 [2M+K]⁺, 371 [M+K]⁺, 355.1729 [M+Na]⁺ (Calcd for C₁₆H₂₈NaO₇;

355.1733), 333 [M+H]⁺, 315 [M-H₂O+H]⁺, 153 [M-C₆H₁₂O₆+H]⁺ (base). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

Enzymatic Hydrolysis of 12 A mixture of **12** (5 mg) and β-glucosidase (5 mg) in water (5 ml) was shaken in a water bath at 37 °C for 5 d. The mixture was treated in the same way as described for **1** to afford an aglycone **12a** (2 mg) and a sugar fraction. From the sugar fraction, D-glucose was detected as described for **1**.

(3R,4R)-p-Menth-1-ene-3,4-diol (12a) An amorphous powder, [α]_D²¹ -29° (c=0.2, MeOH). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

(3R,4R,6R)-p-Menth-1-ene-3,4,6-triol 3-O-β-D-Glucopyranoside (13) An amorphous powder, [α]_D²² -97° (c=0.1, MeOH). Positive FAB-MS *m/z*: 697 [2M+K]⁺, 387 [M+K]⁺, 371 [M+Na]⁺, 349.1867 [M+H]⁺ (base, Calcd for C₁₆H₂₉O₈; 349.1863), 331 [M-H₂O+H]⁺, 169 [M-C₆H₁₂O₆+H]⁺ (base). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-4, C-6, C-7; H-3/C-1, C-2, Glc C-1; H-5_{ax}/C-3, C-4, C-6; H-5_{eq}/C-1, C-3, C-4, C-6, C-8; H-6_{eq}/C-1, C-2, C-4, C-7; H₃-7/C-1, C-2, C-6; H-8/C-3, C-4, C-5, C-9, C-10; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-3.

(1S,2R,4R)-p-Menth-5-ene-1,2-diol 2-O-β-D-Glucopyranoside (14) An amorphous powder, [α]_D²³ +8° (c=1.8, MeOH). Positive FAB-MS *m/z*: 665 [2M+K]⁺, 371 [M+K]⁺, 355.1738 [M+Na]⁺ (Calcd for C₁₆H₂₈NaO₇; 355.1733), 333.1902 [M+H]⁺ (Calcd for C₁₆H₂₉O₇; 333.1913), 315 [M-H₂O+H]⁺, 153 [M-C₆H₁₂O₆+H]⁺ (base). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-1, C-3, C-4, C-6, Glc C-1; H₃-3/C-1, C-2, C-4, C-5, C-8; H-4/C-5, C-6, C-8, C-9, C-10; H-5/C-1, C-3, C-4, C-8; H-6/C-1, C-2, C-4, C-7; H₃-7/C-1, C-2, C-6; H-8/C-3, C-4, C-5, C-9, C-10; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-2.

Enzymatic Hydrolysis of 14 A mixture of **14** (12 mg) and β-glucosidase (5 mg) in water (5 ml) was shaken in a water bath at 37 °C for 20 d. The mixture was treated in the same way as described for **1** to afford an aglycone **14a** (5 mg) and a sugar fraction. From the sugar fraction, D-glucose was detected as described for **1**.

(1S,2R,4R)-p-Menth-5-ene-1,2-diol (14a) An amorphous powder, [α]_D²² +24° (c=0.3, MeOH). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

(1S,2R,4R)-p-Menth-5-ene-1,2-diol 1-O-β-D-Glucopyranoside (15) An amorphous powder, [α]_D²¹ -23° (c=0.6, MeOH). Positive FAB-MS *m/z*: 371 [M+K]⁺, 355.1733 [M+Na]⁺ (Calcd for C₁₆H₂₈NaO₇; 355.1733), 333 [M+H]⁺, 315 [M-H₂O+H]⁺, 153 [M-C₆H₁₂O₆+H]⁺ (base). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-1, C-3, C-4, C-6, C-7; H₃-3/C-1, C-2, C-4, C-5, C-8; H-4/C-2, C-3, C-5, C-6, C-8, C-9, C-10; H-5/C-1, C-3, C-4, C-8; H-6/C-2, C-4, C-7; H₃-7/C-1, C-2, C-6; H-8/C-3, C-4, C-5, C-9, C-10; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-1.

Enzymatic Hydrolysis of 15 A mixture of **15** (5 mg) and β-glucosidase (5 mg) in water (5 ml) was shaken in a water bath at 37 °C for 20 d. The mixture was treated in the same way as described for **1** to afford an aglycone **15a** (5 mg) and a sugar fraction. From the sugar fraction, D-glucose was detected as described for **1**.

(1S,2R,4S)-p-Menth-5-ene-1,2,4-triol 2-O-β-D-Glucopyranoside (16) An amorphous powder, [α]_D²³ -60° (c=1.1, MeOH). Positive FAB-MS *m/z*: 387 [M+K]⁺, 371.1693 [M+Na]⁺ (Calcd for C₁₆H₂₈NaO₈; 371.1681), 349 [M+H]⁺, 313 [M-2H₂O+H]⁺, 169 [M-C₆H₁₂O₆+H]⁺ (base). ¹H-NMR

(pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2. HMBC Correlations: H-2/C-1, C-3, C-4, C-6, C-7, Glc C-1; H₂-3/C-1, C-2, C-4, C-5, C-8; H-5/C-1, C-3, C-4, C-6, C-8; H-6/C-1, C-2, C-4, C-5, C-7; H₃-7/C-1, C-2, C-6; H-8/C-3, C-4, C-5, C-9, C-10; H₃-9/C-4, C-8, C-10; H₃-10/C-4, C-8, C-9; Glc H-1/C-2.

Enzymatic Hydrolysis of 16 A mixture of **16** (14 mg) and β-glucosidase (5 mg) in water (5 ml) was shaken in a water bath at 37 °C for 14 d. The mixture was treated in the same way as described for **1** to afford an aglycone **16a** (6 mg) and a sugar fraction. From the sugar fraction, D-glucose was detected as described for **1**.

(1S,2R,4S)-p-Menth-5-ene-1,2,4-triol (16a) An amorphous powder, [α]_D²³ -50° (c=0.5, MeOH). ¹H-NMR (pyridine-*d*₅, 500 MHz) δ: Table 1. ¹³C-NMR (pyridine-*d*₅, 125 MHz) δ: Table 2.

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