

Hydroxypropyl Methylcellulose Based Cephalixin Extended Release Tablets: Influence of Tablet Formulation, Hardness and Storage on *in Vitro* Release Kinetics

Muniyandy SARAVANAN,^{*a} Kalakonda SRI NATARAJ,^a and Kettavarampalayam Swaminath GANESH^b

^aDepartment of Pharmaceutics, Vel's College of Pharmacy; Chennai-600117, India; and ^bShelys Pharmaceuticals Limited; Dar es salaam, Tanzania. Received January 6, 2003; accepted May 12, 2003

The object of this study was to develop hydroxypropyl methylcellulose (HPMC) based cephalixin extended release tablet, which can release the drug for six hours in predetermined rate. Twenty-one batches of cephalixin tablets were prepared by changing various physical and chemical parameters, in order to get required theoretical release profile. The influences of HPMC, microcrystalline cellulose powder (MCCP), granulation technique, wetting agent and tablet hardness on cephalixin release from HPMC based extended release tablets were studied. The formulated tablets were also characterized by physical and chemical parameters. The dissolution results showed that a higher amount of HPMC in tablet composition resulted in reduced drug release. Addition of MCCP resulted in faster drug release. Tablets prepared by dry granulation was released the drug slowly than the same prepared with a wet granulation technique. Addition of wetting agent in the tablets prepared with dry granulation technique showed slower release. An increase in tablet hardness resulted in faster drug release. Tablets prepared with a wet granulation technique and having a composition of 9.3% w/w HPMC with a hardness of 10–12 kg/cm² gave predicted release for 6 h. The *in vitro* release data was well fit in to Higuchi and Korsmeyer–Peppas model. Physical and chemical parameters of all formulated tablets were within acceptable limits. One batch among formulated twenty-one batches was successful and showed required theoretical release. The effect of storage on *in vitro* release and physicochemical parameters of successful batch was studied and was found to be in acceptable limits.

Key words cephalixin; extended release tablet; hydroxypropyl methylcellulose; tablet hardness; release kinetics

Cephalixin is a semi synthetic antibiotic derived from Cephalosporin 'C'. It is absorbed completely (80–100%) after oral administration¹⁾ and having a biological half-life²⁾ of 1 h. To maintain therapeutic range, the drug should be administered 3–4 times a day, which leads to the saw tooth kinetic of the absorption and resulting in ineffective therapy. The conventional oral regimen results in initial high peak plasma level and that fall drastically below the effective concentration before the next dose. Hence, many authors attempted to develop sustained/extended release dosage forms for cephalixin in order to achieve constant effective plasma concentration. Shin and Cho³⁾ studied cephalixin release kinetics from Eudragit-hydroxypropyl cellulose membranes. Martinez-Pacheco *et al.*^{4,5)} formulated double-layer tablets containing small proportions of acrylic resins for cephalixin-controlled release and also studied the effect of compression force on biopharmaceutical characteristics of Eudragit RS-based cephalixin tablets. Schneider *et al.*⁶⁾ evaluated cephalixin prolonged release formulation for better therapy. Dhopeswarkar *et al.*⁷⁾ developed cephalixin sustained release matrix tablet by using xanthan gum and sodium alginate.

In our previous work, we reported⁸⁾ Eudragit L100 based cephalixin tablet with ideal release profile. In the present study we attempted to formulate cephalixin extended release tablet by using hydroxypropyl methylcellulose (HPMC), which is economic and the drug release from HPMC matrix is uniform irrespective of the pH. Since the solubility of HPMC is pH independent, constant release rate throughout the gastrointestinal tract can be expected from the HPMC based tablets than the Eudragit whose solubility is pH dependent. The influences of HPMC, microcrystalline cellulose (MCCP), method of granulation technique, wetting

agent, hardness, and storage on *in vitro* release profile were studied to find out suitable tablet formulation with acceptable physical and chemical parameters. The tablets were characterized by drug content, weight variation, hardness, thickness, friability, and stability. The *in vitro* release of formulated extended release tablet was compared with a marketed sample.

Theory By considering pharmacokinetic parameters, we have calculated and reported⁸⁾ theoretical release profile of cephalixin for an ideal tablet. Briefly, the tablet should release 125 mg of cephalixin initially within first 1 h and 46.7 mg of cephalixin per hour for next 5 h from 375 mg of total dose in order to maintain plasma cephalixin concentration of 4.5 mg/l. The percentage of drug to be released from an ideal tablet containing 375 mg of cephalixin is given in Fig. 1.

Experimental

Materials Cephalixin IP was obtained from Orchid Chemicals and Pharmaceutical Ltd., India. Hydroxypropyl methylcellulose (Methocel 15 cps) was obtained from Dow chemicals India. Polyvinylpyrrolidone was purchased from Shanghai Sun Power New Material Company, China. Magnesium Stearate IP was procured from Sinai Pharma Pvt Ltd., India. Lactose IP was purchased from Lactose India Ltd. All other chemicals used were of analytical grade.

Preparation of Cephalixin Extended Release Tablets by Wet Granulation Technique The cephalixin extended release tablets (batches 1–5) were prepared by wet granulation⁹⁾ technique. Ingredients required for 6000 tablets as per the formula given in Table 1 were weighed and granulated as follows. The drug and HPMC were separately passed through sieve #40 and 60, respectively and mixed with lactose, which was previously passed through sieve #40, in a double cone blender for 5 min. After mixing the powders were transferred to 10 l capacity rapid mixer granulator (Kevin engineers, India) and granulated for 3 min by using 10% w/v Polyvinylpyrrolidone (PVP) in isopropyl alcohol (50% v/v) as binding agent. The wet granules were passed through sieve #18 and dried at 40 °C for 90 min in a tray

* To whom correspondence should be addressed. e-mail: msaravanan72@hotmail.com

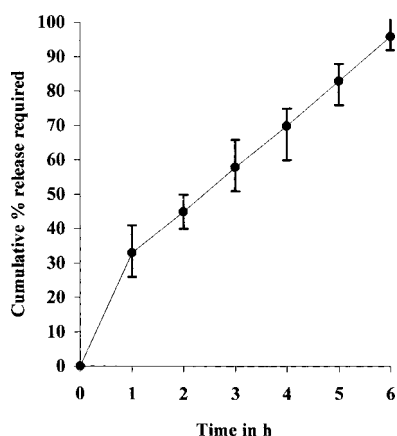


Fig. 1. Theoretical Release Profile of Ideal Extended Release Tablet Containing 375 mg of Cephalexin and Intended to Maintain Effective Concentration for 6 h

The bar indicates theoretical release limits.

drier (Bombay Engineering Works, India). The dried granules were passed through sieve #14 and the moisture content was determined by Karl Fischer method (Karl-Fischer Titrator, Precision V/M MD, India). Then the dried granules were passed through sieve #14, lubricated with magnesium stearate by mixing in rapid mixer granulator (Kevin Engineers, India) at slow speed for 5 min and compressed using 15/32 flat punches in Cadmach tablet compression machine to get tablets. Batches 1, 2 and 5 were prepared by using different ratio of HPMC as per the formula given in Table 1. Batches 2, 3 and 4 were prepared with varying quantity of MCCP. To study the effect of hardness on release profile, in each batch, three sub-batches A, B and C were prepared with a hardness of 6–8, 8–10 and 10–12 kg/cm², respectively.

Preparation of Cephalexin Extended Release Tablets by Dry Granulation Technique The granules required for batch 6 (6000 tablets) were prepared by dry granulation technique⁹ as per the formula given in Table 1. The drug and HPMC were separately passed through sieve #40 and 60, respectively and mixed with lactose, which was previously passed through sieve #40, in a double cone blender. The resulting mixture was compressed into slugs using 12.5 mm flat punches. The slugs thus obtained were crushed in Kalweka dry granulator, passed through sieve #14 and the moisture content were measured (Karl-Fischer Titrator, Precision V/M MD, India). The ultimately obtained granules were lubricated with magnesium stearate by mixing in a rapid mixer granulator (Kevin Engineers, India) at slow speed for 5 min and were compressed to tablets by using 15/32 flat punches.

Batch 7 was prepared with a wetting agent polysorbate 80, in order to study its influence on the *in vitro* release. The tablets were prepared by the procedures similar to that of batch 6. Polysorbate 80 was added to cephalexin/HPMC mixture in the double cone blender and mixed for 5 min. Then the lactose was added and the tablets were prepared as described for batch 6. To study the effect of hardness on release profile, in each batch three sub-batches A, B and C were prepared with hardness of 6–8, 8–10 and 10–12 kg/cm², respectively.

Physical Parameters The formulated tablets were tested^{9,10} for weight variation, thickness, friability (Friability test apparatus, Indian Equipment Corporation) and hardness (DrSchleuniger Pharmatron).

Drug Content Twenty tablets were weighed and powdered. The quantity equivalent to 100 mg of cephalexin was weighed accurately and taken in 100 ml volumetric flask. Fifty milliliters of water was added, sonicated (Sonicator-Branson, SmithKline) for 5 min, made up to 100 ml with water, and filtered. Two milliliters of above solution was diluted to 100 ml in a volumetric flask and the drug was determined at 261 nm⁵ by using UV-visible spectrophotometer (Shimadzu UV-2201).

In Vitro Release Studies The *in vitro* release of cephalexin from formulated tablets was carried out in 0.1 N HCl for 1 h, and continued in 0.01 N HCl for another 1 h and finally in phosphate buffer pH 7.4 for 4 h. The studies were performed in USP dissolution apparatus 1 (Programmable tablet dissolution tester USP XXI and XXII, TDT 067, ELECTROLAB, India) at 37 ± 2 °C and 100 rpm. Samples were taken at hourly interval and analysed for cephalexin content at 261 nm⁵ by using UV-visible spectrophotometer. The same procedure was followed to study the *in vitro* release of cephalexin

from a marketed product.

Stability Studies The formulated cephalexin tablets, batch 2 which gave *in vitro* drug release complying the calculated limits, were kept for a short term accelerated stability study in high density polyethylene sealed cover at 40 ± 2 °C/75 ± 5% RH as per International Conference on Harmonization States (ICH) guidelines. Samples were with-drawn for every month of storage and evaluated⁸ for appearance, hardness, drug content, and dissolution.

Release Kinetics Data obtained from *in vitro* release studies were fitted to various kinetic equations. The kinetic models¹¹ used are zero order, first order and Higuchi equation. The following plots were made: Q_t vs. t (zero order kinetic model); $\log(Q_0 - Q_t)$ vs. t (first order kinetic model) and Q_t vs. square root of t (Higuchi model). Where Q_t is the amount of cephalexin released at time t and Q_0 is the initial amount of cephalexin present in tablets. Further, to find out the mechanism of drug release, first 60% drug release was fitted in Korsmeyer–Peppas model:

$$M_t/M_\infty = kt^n$$

where M_t/M_∞ is fraction of drug released at time t , k is rate constant and n is release exponent. The n value is used to characterize different release mechanisms.¹¹

Results and Discussion

An ideal extended release tablet should release the required quantity of drug with predetermined kinetics in order to maintain effective drug plasma concentration. To achieve this the tablet should be formulated in such a way to release the drug in a predetermined and reproducible manner. By considering the biopharmaceutic and pharmacokinetic profile of the drug, the required release from the tablet can be predetermined.⁸ To achieve the predetermined release profile, various formulation factors like polymer/drug ratio, hardness and additives should be modified to get the required release. Ideally, the tablet should release the drug as per the predetermined rate even under storage conditions.

Cephalexin is effective in wide variety of infections because they have a broad spectrum and high therapeutic/toxic ratio. Cephalexin monohydrate^{2,12} is a white to cream crystalline solid with bitter taste and having molecular weight of 365.4 (347.4 for anhydrous). It is soluble in water (1 g in 100 ml) and in dilute aqueous alkaline solutions. It is very slightly soluble to practically insoluble in alcohol and other organic solvents. Cephalexin is sensitive to moisture, heat and light. Generally tablets, capsules and dry powders should be stored² between 15–30 °C, in a dry and cool place. Reconstituted suspensions² are stable for 7 to 14 d at between 2–8 °C and 6–15 °C. In hydrochloric acid buffer (pH 1.2), cephalexin lost 5% activity in 24 h at 37 °C as compare to a 45% loss in phosphate buffer at pH 6.5.¹³ Cephalexin in serum was found to lose 10%, 50%, 75% activity respectively, after storage at 5 °C, 25 °C and 37 °C for 48 h.^{13,14} The antibiotic retains activity well in serum and urine as no loss in activity was noted after storage at –20 °C for 14 d.¹⁴

Because of shorter biological half-life, cephalexin should be preferably given in extended release dosage forms. In our previous work, we have calculated and reported the required theoretical release profile of cephalexin⁸ from tablets (Fig. 1) and formulated Eudragit L100 based cephalexin tablet with ideal characteristic. In the present work, we tried to develop HPMC based cephalexin tablets, which could release the drug in predetermined rate for 6 h. Twenty-one batches were formulated by changing formulation parameters as per the formula given in Table 1, in order to study the effect on *in vitro* release kinetics and to find out the tablet formulation,

Table 1. Composition of Cephalexin Extended Release Tablets

Ingredients (mg/tablet)	Batch 1	Batch 2	Batch 3	Batch 4	Batch 5	Batch 6	Batch 7
	Wet granulation				Dry granulation		
Cephalexin IP	375	375	375	375	375	375	375
HPMC 15 cps	20	35	35	35	50	35	35
Lactose	40	40	40	40	40	40	40
MCCP			10	20			
Polysorbate 80							5
PVP	7	7	7	7	7		
Magnesium stearate	5	5	5	5	5	5	5
Total weight	447	462	472	482	477	455	460
% of HPMC to cephalaxin	5.3	9.3	9.3	9.3	13.3	9.3	9.3
% of MCCP to cephalaxin			2.7	5.3			

In each batch three sub batches A, B and C were prepared with hardness of 6–8, 8–10 and 10–12 kg/cm², respectively.

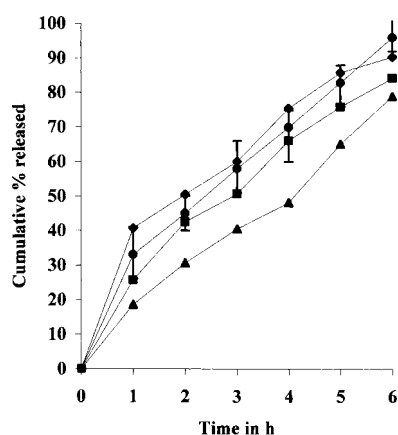


Fig. 2. Influence of HPMC on the *in Vitro* Release of Cephalexin from Formulated Tablets

The figure illustrates the cephalexin release from batch 1A (—◆—), 2A (—■—) and 5A (—▲—) tablets formulated with 5.3, 9.3 and 13.3% of HPMC with respect to cephalaxin and having a hardness of 6–8 kg/cm². The release is compared with theoretical release profile (—●—). Each data represents average of six readings and bar represents theoretical release limits.

which will give required release profile.

Batches 1, 2 and 5 were formulated by using various cephalaxin/HPMC proportions as per formula given in Table 1, in order to study the effect of HPMC on drug release profile. Amount of percentage of polymer added in each batch is shown in Table 1. Figure 2 shows the cephalexin cumulative percentage released *versus* time for tablets formulated with various percentage of HPMC. All the batches showed a release over 4–6 h. As expected, the release rate was slower with higher quantity of HPMC, the tablets having 9.3% of HPMC with respect to drug showed optimum release profile as shown in Fig. 2. At higher percentage of HPMC in tablets, when in contact with release medium, HPMC may swell^{15,16} and form a thick gel, thus may decrease the size of the pores present in the tablet and reducing the drug release.

Using the same formula and changing the granulation technique batch 2 and 6 were formulated in order to find out the change in release kinetics. As shown in Fig. 3, tablets formulated by dry granulation released the drug slowly than the tablet formulated by wet granulation technique. This may be due to slow penetration of dissolution medium into the tablet prepared by dry granulation technique. The presence of more moisture in granules prepared by dry granulation technique

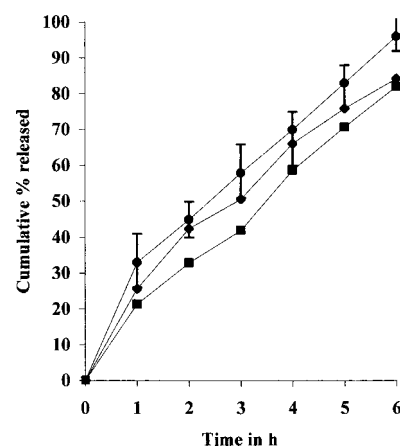


Fig. 3. Influence of Granulation Technique on the *in Vitro* Release of Cephalexin from Formulated Tablets

The figure illustrates the drug release from batch 2A (—◆—) and 6A (—■—), tablets formulated by wet and dry granulation, respectively and having a hardness of 6–8 kg/cm². The release is compared with theoretical release profile (—●—). Each data represents average of six readings and bar represents theoretical release limits.

(Table 2) resulted in the faster swelling of HPMC matrix, which might reduced the pore size through which diffusion of drug towards the dissolution medium^{15,16} occurs and thus slower the release of drug. More over the presence of PVP, highly water soluble additive, in the tablets prepared by wet granulation technique, may undergo rapid dissolution which may favour penetration/contact of dissolution medium inside the swollen HPMC matrix/drug and thus may give faster release.

To study the influence of wetting agent on *in vitro* release profile in batch 7, polysorbate 80 was added as wetting agent and its effect on *in vitro* release was studied. The influence of polysorbate 80 on the *in vitro* release of cephalaxin from the formulated tablets is given Fig. 4. Theoretically, the addition of wetting agent, which enhances the contact of drug and dissolution medium, should result in faster dissolution. Nevertheless, as shown in Fig. 4, the presence of the wetting agent in the HPMC matrix tablet has reduced the drug release considerably. Addition of wetting agent in HPMC tablets will enhance the contact of polymer matrix with the dissolution medium, which in turn may produce rapid swelling of the tablet and thus produced slower release.

In batches 2, 3 and 4 the quantity of MCCP incorporated

Table 2. Physical and Chemical Parameters of Formulated Cephalixin Tablets

Evaluated parameters	Batch 1			Batch 2			Batch 3			Batch 4			Batch 5			Batch 6			Batch 7			Marketed sample					
	A	B	C	A	B	C	A	B	C	A	B	C	A	B	C	A	B	C	A	B	C						
Average weight (mg)	495 ± 25	496 ± 28	498 ± 31	486 ± 31	485 ± 20	486 ± 34	486 ± 31	485 ± 25	489 ± 39	505 ± 29	500 ± 35	502 ± 45	502 ± 49	500 ± 45	492 ± 36	498 ± 25	495 ± 31	498 ± 25	502 ± 40	492 ± 40	502 ± 40	498 ± 25	495 ± 28	492 ± 35	490 ± 45	515 ± 51	
Thickness (mm)	4.2 ± 0.12	4.3 ± 0.15	4.2 ± 0.2	4.4 ± 0.21	4.5 ± 0.25	4.4 ± 0.24	4.0 ± 0.2	4.2 ± 0.28	4.0 ± 0.15	4.2 ± 0.18	4.5 ± 0.25	4.3 ± 0.29	4.5 ± 0.29	4.3 ± 0.22	4.4 ± 0.18	254.2 ± 0.25	4.1 ± 0.12	4.3 ± 0.25	4.3 ± 0.2	4.3 ± 0.2	4.1 ± 0.12	4.5 ± 0.21	4.3 ± 0.21	4.2 ± 0.13	4.4 ± 0.15		
Friability %	0.8 ± 98.6	0.9 ± 97.7	0.9 ± 98.2	0.9 ± 98.6	0.7 ± 101.6	0.8 ± 99.4	0.9 ± 97.6	0.9 ± 99.7	0.9 ± 98.3	0.6 ± 99.8	0.5 ± 101.7	0.6 ± 101.6	0.9 ± 100.3	0.7 ± 101.2	0.9 ± 100.4	0.6 ± 98.6	0.8 ± 102.5	0.6 ± 98.6	0.8 ± 98.1	0.8 ± 98.1	0.8 ± 98.1	0.6 ± 98.6	0.9 ± 97.7	0.5 ± 98.8	1.0 ± 98.6	0.75 ± 99.5	
Assay	2.34 ± 1.98	1.98 ± 0.85	2.4 ± 0.85	2.4 ± 1.87	1.25 ± 1.25	1.87 ± 1.87	0.77 ± 0.77	2.42 ± 2.42	2.56 ± 2.56	1.55 ± 1.55	2.67 ± 2.67	2.88 ± 2.88	3.25 ± 3.25	2.45 ± 2.45	1.96 ± 1.96	0.85 ± 0.85	1.68 ± 1.68	0.85 ± 0.85	0.74 ± 0.74	0.74 ± 0.74	0.74 ± 0.74	0.85 ± 0.85	0.57 ± 0.57	1.25 ± 1.25	1.78 ± 1.78	1.24 ± 1.24	
Disintegration time (min)	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	7 ± 2	—
Moisture content	4.7 ± 0.35			4.7 ± 0.22			4.5 ± 0.76			4.1 ± 0.45			4.9 ± 0.67			5.5 ± 0.96			6.5 ± 0.36			—					

In each batch three sub batches A, B and C were prepared with hardness of 6–8, 8–10 and 10–12 kg/cm², respectively. Results are expressed as mean ± S.D. (n=6). NA: not applicable.

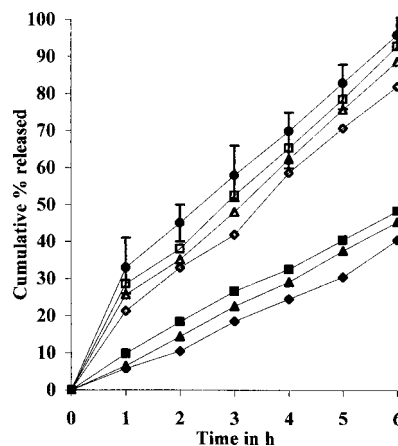


Fig. 4. Influence of Polysorbate 80 and Tablet Hardness on the *in Vitro* Release of Cephalixin from HPMC Tablets

The figure illustrates the drug release from batch 6A (—◇—), 6B (—△—), 6C (—□—), 7A (—◆—), 7B (—▲—) and 7C (—■—) tablets. Sub batches A, B and C indicate a hardness of 6–8, 8–10 and 10–12 kg/cm², respectively. The release is compared with theoretical release profile (—●—). Each data represents average of six readings and bar represents theoretical release limits.

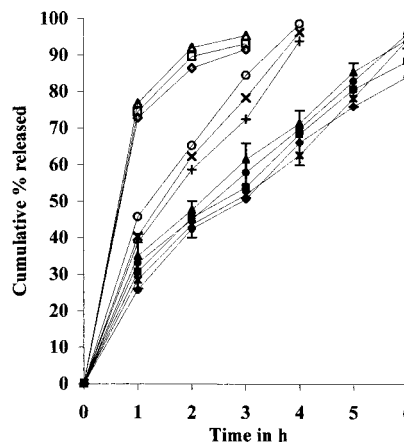


Fig. 5. Influence of Percentage of MCCP and Tablet Hardness on the *in Vitro* Release of Cephalixin from HPMC Tablets

The figure illustrates the drug release from batch 2A (—◆—), 2B (—■—), 2C (—▲—), 3A (—+—), 3B (—×—), 3C (—○—), 4A (—◇—), 4B (—□—) and 4C (—△—) tablets. Batch 2, 3 and 4 were formulated with 0, 2.65 and 5.3% of MCCP with respect to cephalixin. Sub batches A, B and C indicate a hardness of 6–8, 8–10 and 10–12 kg/cm², respectively. The release is compared with theoretical release profile (—●—) and a marketed sample (*). Each data represents average of six readings and bar represents theoretical release limits.

was varied, to find out its effect on the dissolution profile. *In vitro* release of cephalixin from tablet formulation made with different percentage of MCCP was given in Fig. 5. Incorporation of MCCP enhanced the drug release from the HPMC tablets. MCCP, in general, is used in tablet formulation as diluent and disintegrant. The disintegrant MCCP may disintegrate the hydrated layer, which is formed around the HPMC matrix when in contact with dissolution medium, thus may form pores/channels, thereby enhance the contact between drug and dissolution medium to give faster drug release. In all batches, three sub batches were prepared with different hardness (Table 1) in order to study its influence on *in vitro* release and to find out the suitable formulation, which can release the cephalixin in the predetermined rate. In general, increase in hardness in tablet will result in less porosity and slow drug release. As indicated in Figs. 4, 5 and 6, the increase in hardness of tablets formulated with HPMC

results in increased drug release. This may be due to faster swelling of HPMC in tablets having lower hardness and higher porosity. Tablets with low hardness swell immediately forming a gel like layer around the tablet and blocking the surface pores, resulting in slower drug release. Therefore, this is also a considerable factor in optimising drug release. Only batch 2 has showed required release profile among formulated seven batches and it is comparable with the release of a marketed formulation as shown in Fig. 5.

All other evaluation parameters like drug content, hardness, friability, weight variation, thickness and moisture content were studied for all the batches. All batches passed the acceptable limits of their respective parameters as shown in Table 2. The *in vitro* release data obtained were fitted in to

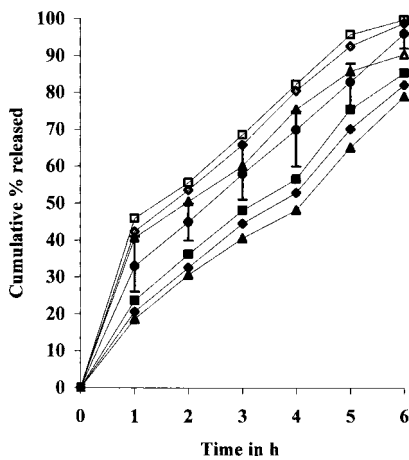


Fig. 6. Influence of Tablet Hardness on the *in Vitro* Release of Cephalexin from HPMC Tablets

The figure illustrates the drug release from batch 1A (—△—), 1B (—◇—), 1C (—□—), 5A (—▲—), 5B (—◆—) and 5C (—■—) tablets. Sub batches A, B and C indicate a hardness of 6–8, 8–10 and 10–12 kg/cm², respectively. The release is compared with theoretical release profile (—●—). Each data represents average of six readings and bar represents theoretical release limits.

various kinetic equations. Correlations of individual batch with applied equation are given in Table 4. The release rates were calculated from the slope of the appropriate plots. Batches 1, 2, 3 and 4 showed higher correlation with Higuchi plot than zero order and first order. Batches 5, 6, 7 showed higher correlation with zero order equation than Higuchi and first order. To find out release mechanism the *in vitro* release data were applied in Korsmeyer–Peppas equation. The release exponent *n* was determined and given in Table 4. Batch 1 tablets formulated with 5.3 percentage of HPMC with respect to drug showed (*n*=0.28 to 0.35) Fickian diffusion. Batches prepared with 9.3 and 13.3% of HPMC showed (*n*=0.51 to 0.7) Anomalous (non-Fickian) diffusion. The tablet formulated with wetting agent (batch 7) showed (*n*=0.88 to 1.20) super case-II diffusion principle.

Accelerated stability studies were performed on batch 2 tablets as per ICH guidelines. The cephalexin content and *in vitro* release were tested at periodic time intervals. The cephalexin contents were decreased periodically as shown in

Table 3. Physical and Chemical Parameters of Batch 2 Tablets during Stability Studies

Batch No.	Stability duration	Appearance	Hardness (kg/cm ²)	% Drug content
2A	Initial	Pale yellow	6–8	98.6±2.4
	After 30 d	Pale yellow	3.5–5.5	95.4±3.2
	After 60 d	Pale yellow	3–3.5	91.8±1.9
	After 90 d	Pale yellow	2–3	87.6±3.5
2B	Initial	Pale yellow	8–10	101.6±1.25
	After 30 d	Pale yellow	6.5–7.5	97.9±2.45
	After 60 d	Pale yellow	5–6	90.5±2.88
	After 90 d	Pale yellow	4–5.5	88.4±3.49
2C	Initial	Pale yellow	10–12	99.4±1.87
	After 30 d	Pale yellow	7.5–9.5	96.5±2.33
	After 60 d	Pale yellow	7–9	93.6±3.48
	After 90 d	Pale yellow	6–8	90.7±2.4

Results are expressed as mean±S.D. (*n*=6).

Table 4. Coefficient, *in Vitro* Release Rate and the Release Exponent of Cephalexin from Formulated Tablets

Batch No.	Zero order		First order		Higuchi		Korsmeyer–Peppas	
	<i>r</i> ²	<i>k</i> ₀ (h ⁻¹)	<i>r</i> ²	<i>k</i> ₁ (h ⁻¹)	<i>r</i> ²	<i>k</i> _H (h ^{-1/2})	<i>r</i> ²	<i>n</i> value
1A	0.9113	13.80	0.8866	0.0249	0.9924	37.03	0.9922	0.35
1B	0.9229	15.14	0.7159	0.0375	0.9948	40.40	1	0.34
1C	0.9067	15.21	0.6028	0.0479	0.9929	40.91	1	0.28
2A	0.9663	13.47	0.9305	0.0211	0.9823	34.90	0.9854	0.63
2B	0.9575	13.87	0.9000	0.0233	0.9874	36.22	0.9942	0.52
2C	0.9500	14.58	0.8374	0.0278	0.9920	38.30	0.9907	0.51
3A	0.9636	22.19	0.8315	0.0274	0.9847	45.37	1	0.61
3B	0.9598	23.10	0.7935	0.0306	0.9909	47.47	1	0.63
3C	0.9418	23.63	0.7142	0.0371	0.9969	49.17	1	0.52
4A	0.7665	28.88	0.8866	0.0287	0.9496	55.10	NA	NA
4B	0.7607	29.48	0.9135	0.0261	0.9467	56.36	NA	NA
4C	0.7559	30.18	0.9223	0.0244	0.9441	57.81	NA	NA
5A	0.9889	12.41	0.9456	0.0192	0.9301	30.94	0.9990	0.69
5B	0.9901	13.07	0.9368	0.0203	0.9435	32.80	0.9989	0.69
5C	0.9840	13.58	0.9239	0.0218	0.9550	34.38	0.9990	0.64
6A	0.9906	13.25	0.9410	0.0187	0.9458	33.28	0.9808	0.70
6B	0.9873	14.07	0.9035	0.0233	0.9544	35.55	0.9723	0.63
6C	0.9818	14.51	0.8587	0.0263	0.9635	36.95	0.9666	0.54
7A	0.9914	6.64	0.9940	0.0127	0.8710	15.98	0.9930	1.10
7B	0.9991	7.63	0.9923	0.0132	0.9002	18.60	0.9989	1.08
7C	0.9963	7.89	0.9903	0.0136	0.9392	19.70	0.9990	0.88

NA: not applicable because more than 70% of the drug has released with in first 1 h.

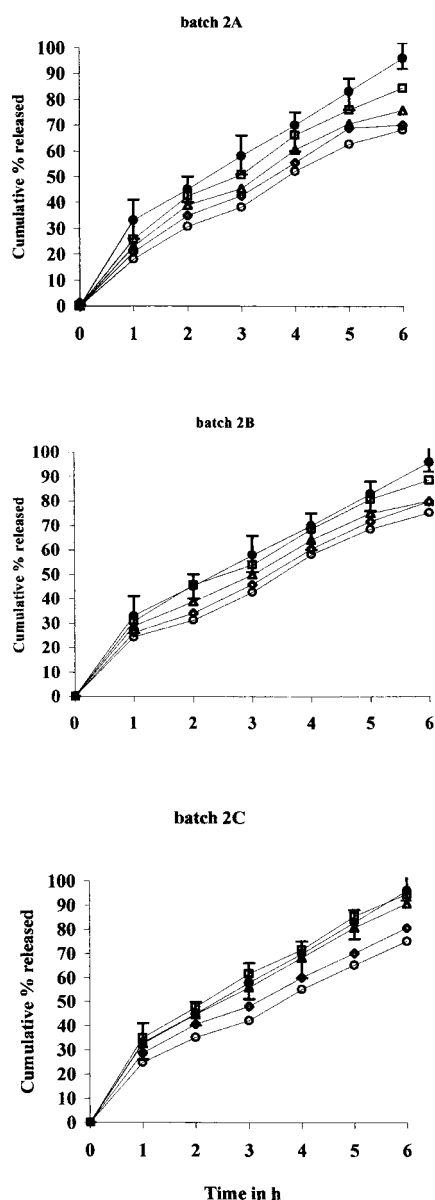


Fig. 7. The *in Vitro* Release Profile of Batch 2 during Stability Studies
Initial (—□—), first month (—△—), second month (—◇—) and third month (—○—). The release is compared with theoretical release profile (—●—). Each data represents average of six readings and bar represents theoretical release limits.

Table 3, this may be due to the presence of moisture and heat.^{2,13,14} The changes of *in vitro* release profile observed during stability studies are shown in Fig. 7. The drug release become slower on storage, this is because of decrease in hardness of the tablet, may be due to ageing of HPMC gel, as given in Table 3. *In vitro* release from the batch 2C with the hardness of 10–12 kg/cm² was found to be within predicted release profile for longer time than the other batches. All other tested parameters of batch 2C were in acceptable limits

Table 5. Change in Coefficient, Release Rate Constant and Release Exponent during Stability Studies

Batch No. and period	Higuchi		Korsmeyer–Peppas	
	r^2	k_H (h ^{-1/2})	r^2	n value
2A _{initial}	0.9823	34.90	0.9854	0.63
2A _{1 month}	0.9790	31.84	0.9932	0.67
2A _{2 month}	0.9706	30.13	0.9906	0.69
2A _{3 month}	0.9616	28.64	0.9942	0.74
2B _{initial}	0.9874	36.22	0.9942	0.52
2B _{1 month}	0.9838	33.29	0.9898	0.49
2B _{2 month}	0.9681	32.63	0.9560	0.59
2B _{3 month}	0.9621	31.07	0.9399	0.61
2C _{initial}	0.9920	38.30	0.9907	0.51
2C _{1 month}	0.9884	36.499	0.9973	0.49
2C _{2 month}	0.9867	32.07	0.9974	0.47
2C _{3 month}	0.9753	30.016	0.982	0.56

as given in Table 3 and only this batch among formulated twenty one batches, was found to the suitable formulation for cephalixin extended release tablet. The *in vitro* release data obtained during stability studies were fitted to Higuchi and Peppas model to find out the change in release rate and mechanism. The correlation coefficients and n value of tested batches remain nearly the same during stability studies as shown in Table 5. No change in release mechanism was observed, but the release rate constants were decreased as given in Table 5.

References

- 1) Thornhill T. S., Levison M. E., Johnson W. E., Kaye D., *Applied Microbiology*, **17**, 457–461 (1969).
- 2) Colin Dollery (ed.), “Therapeutic Drugs,” 2nd ed., Churchill Livingstone, Edinburgh, 1999, pp. c144–c146.
- 3) Shin S. C., Cho S. J., *Drug Dev. Ind. Pharm.*, **22**, 299–305 (1996).
- 4) Martinez-Pacheco R., Vila-Jato J. L., Concherio A., Souto C., Ramos T., *Int. J. Pharmaceut.*, **47**, 37–42 (1988).
- 5) Martinez-Pacheco R., Vila-Jato J. L., Souto C., Ramos T., *Int. J. Pharmaceut.*, **32**, 99–102 (1986).
- 6) Schneider H., Nightingale C. H., Quintiliani R., Flanagan D. R., *J. Pharm. Sci.*, **67**, 1620–1622 (1978).
- 7) Dhopeswarkar V., O’Keefe J. C., Zatz J. L., Deeter R., Horton M., *Drug Dev. Ind. Pharm.*, **20**, 1851–1867 (1994).
- 8) Saravanan M., Nataraj K. S., Ganesh K. S., *Biol. Pharm. Bull.*, **25**, 541–545 (2002).
- 9) Banker G. S., Anderson N. R., “Theory and Practice of Industrial Pharmacy,” 3rd ed., ed. by Lachman L., Lieberman H. A., Kanig J. L., Varghese Publishing House, Mumbai, 1987, pp. 296–329.
- 10) United States Pharmacopoeia 23, United States Pharmacopoeial Convention, INC., 1995, p. 323.
- 11) Costa P., Sousa Lobo J. M., *E. J. Pharm. Sci.*, **13**, 123–133 (2001).
- 12) Nichols W. K., “Anti-infectives, in Remington: The Science and Practice of Pharmacy,” 19th ed., ed. by Gennaro A. R., Mack Publishing Company, Pennsylvania, 1995, pp. 1290–1297.
- 13) Simmons R. J., *Anal. Microbiology*, **11**, 193–195 (1972).
- 14) Wick W. E., *Applied Microbiology*, **15**, 765–766 (1967).
- 15) Mitchell K., Ford J. L., *Int. J. Pharmaceut.*, **100**, 175–179 (1993).
- 16) Meury G. P., *J. Pharm. Sci.*, **85**, 725–731 (1996).