## Golmaenone, a New Diketopiperazine Alkaloid from the Marine-Derived Fungus *Aspergillus* sp.

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A new diketopiperazine alkaloid, golmaenone (1) and related alkaloids, neoechinulin A (2) and L-alanyl-L-tryptophan anhydride (3), have been isolated from the culture broth of the marine-derived fungus *Aspergillus* sp. The structure and absolute stereochemistry of the new compound (1) was assigned by spectroscopic methods and the advanced Marfey's method. Compounds 1 and 2 exhibited a significant radical scavenging activity against 1,1-diphenyl-2-picrylhydrazyl (DPPH) with IC<sub>50</sub> values of 20 and 24  $\mu$ M, respectively, which are similar to the positive control, ascorbic acid (IC<sub>50</sub>, 20  $\mu$ M). Compounds 1 and 2 also showed an ultraviolet-A (UV-A) (320—390 nm) protecting activity with ED<sub>50</sub> values of 90 and 170  $\mu$ M, respectively, which are more active than oxybenzone (ED<sub>50</sub>, 350  $\mu$ M) currently being used as sunscreen.

**Key words** diketopiperazine alkaloid; golmaenone; neoechinulin A; L-alanyl-L-tryptophan anhydride; marine-derived fungus; *Aspergillus* sp.

Diketopiperazines are widespread microbial products commonly found in nutrient rich cultures of both terrestrial<sup>1)</sup> and marine fungi.<sup>2,3)</sup> Diketopiperazines are of interest because of their activity in various pharmacological assay systems.<sup>4)</sup>

As part of a program to explore the bioactive metabolites produced by the fungi isolated from marine habitats,<sup>5)</sup> we investigated the bioactive constituents of the marine algicolous fungus and isolated a new golmaenone (1) in addition to neoechinulin A (2) and L-alanyl-L-tryptophan anhydride (3).

A fungal strain (culture # MFA 212) was isolated from the surface of the marine red alga *Lomentaria catenata* collected at Golmae Village, Ulsan City, Korea in 2002, and it was identified by fatty acid methyl ester anlaysis (FAME) as a *Aspergillus* sp.<sup>6)</sup> The fungus was cultured (101) in a seawater-based medium.<sup>7)</sup>

The culture broth and mycelium were separated, and the broth was extracted with ethyl acetate to provide a crude extract (1.5 g), which was subjected to a combination of column chromatography on silica gel (*n*-hexane/EtOAc) and octadesyl silica (ODS) gel (H<sub>2</sub>O/MeOH) to furnish three fractions containing diketopiperazines 1 (20 mg), 2 (120 mg), and 3 (35 mg). Further purifications of each fraction by HPLC (YMC ODS-A, MeOH) yielded a new golmaenone (1) (12 mg), as well as neoechinulin A (2) (95 mg) and L-alanyl-L-tryptophan anhydride (3) (7 mg).

Golmaenone (1)<sup>8)</sup> was isolated as a yellow solid which was thought to have a molecular composition of  $C_{19}H_{21}N_3O_4$  from the high resolution (HR)-FAB-MS and <sup>13</sup>C-NMR data.

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Since 1 showed eleven unsaturations in HR-FAB-MS, it implied that 1 contained four carbonyl, five double bonds, and two rings. The IR spectrum of 1 showed absorptions for free amide (3433, 1697 cm<sup>-1</sup>) and hydrogen-bonded amide (3242, 1629 cm<sup>-1</sup>) functionality. The UV spectrum of 1 showed the presence of conjugated amide [222 nm (log  $\varepsilon$  1.8), 327 (1.9), 368 (1.7)] chromophores.

In the <sup>1</sup>H-NMR spectrum, three protons were exchanged by D<sub>2</sub>O, suggesting that 1 has three amide protons [ $\delta$  6.61 (1H, s, H-11), 11.57 (1H, s, H-14), 11.44 (1H, s, H-15). Detailed analyses of the <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of 1, including the results from distortionless enhancement by polarization transfer (DEPT), <sup>1</sup>H-detected heteronuclear multiplequantum coherence (HMQC) and heteronuclear multplebond correlation (HMBC) experiments, revealed signals ascribable to a methyl substituted diketopiperazine [ $\delta$  6.61 (1H, br s, H-11), 4.40 (1H, qd, J=7.0, 1.8 Hz, H-12), 11.57 (1H, s, H-14), 1.66 (3H, d, J=7.0 Hz, H<sub>3</sub>-22), 140.0 (C-9), 157.3 (C-10), 51.8 (C-12), 166.4 (C-13), 21.1 (C-22)], 1,2disubstituted benzene [ $\delta$  7.96 (1H, dd, J=8.0, 1.5 Hz, H-3), 7.14 (1H, ddd, J=8.2, 8.0, 1.0 Hz, H-4), 7.57 (1H, ddd, J=8.6, 8.2, 1.5 Hz, H-5), 8.74 (1H, dd, J=8.6, 1.0 Hz, H-6), 141.4 (C-1), 123.6 (C-2), 130.5 (C-3), 122.6 (C-4), 135.4 (C-5), 121.3 (C-6)], 2,2-dimethyl-3-butenamide [ $\delta$  11.44 (1H, s, H-15), 6.12 (1H, dd, J=17.5, 10.5 Hz, H-18), 5.31, 5.37 (each 1H, d, J=10.5, 17.5 Hz, respectively, H<sub>2</sub>-19), 1.43 (6H, s, CH<sub>3</sub>-20/21), 175.9 (C-16), 46.8 (C-17), 142.4 (C-18), 114.9 (C-19), 24.8 (C-20/21)], and 1,3,3-trisubstitited propenone [ $\delta$  7.22 (1H, s, H-8), 195.1 (C-7), 102.3 (C-8), 140.0 (C-9)] (Table 1).

The connection of the functional groups in 1, which led to the planar structure, was achieved on the basis of HMQC and HMBC correlations. Key HMBC correlations between H-15 and C-2, C-6 and C-16; between H-3 and C-7; between H-8 and C-7 and C-10; between H<sub>3</sub>-20/21 and C-16 and C-18; and between H-14 and C-8, C-10 and C-12, clearly estab-



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Table 1. NMR Spectral Data for Golmaenone  $(1)^{a,b}$ 

C#	$\delta_{\mathrm{H}} \left( \mathrm{mult.}, J \right)$	$\delta_{ m C}$ (mult.)	HMBC (H to C)
1		141.4	
2		123.6	
3	7.96 (dd, 8.0, 1.5)	130.5	1, 4, 5, 7
4	7.14 (ddd, 8.2, 8.0, 1.0)	122.6	2, 3, 5, 6
5	7.57 (ddd, 8.6, 8.2, 1.5)	135.4	1, 3, 4
6	8.74 (dd, 8.6, 1.0)	121.3	1, 2, 4
7		195.1	
8	7.22 (s)	102.3	7,10
9		140.0	
10		157.3	
11	6.61 (br s)		9,13
12	4.40 (qd, 7.0, 1.8)	51.8	10, 13, 22
13		166.4	
14	11.57 (s)		10, 12
15	11.44 (s)		2, 6, 16
16		175.9	
17		46.8	
18	6.12 (dd, 17.5, 10.5)	142.4	16, 17, 20/21
19	5.37 (d, 17.5)	114.9	17, 18
	5.31 (d, 10.5)		
20/21	1.43 (s)	24.8	16, 17, 18
22	1.66 (d, 7.0)	21.1	12, 13

a) Recorded in CDCl<sub>3</sub> at 400 MHz (<sup>1</sup>H) and 100 MHz (<sup>13</sup>C). b) Assignments aided by DEPT, HMQC, and HMBC.

lished the planar structure of 1.

The geometry of C-8/C-9 double bond in compound 1 was determined to be (Z) configuration on the basis of the chemical shifts of H-8 [ $\delta$  7.22 (1H, s)] and H-14 [ $\delta$  11.57 (1H, s)], which were shifted to the low field by the deshielding effect of the carbonyl group on  $\beta$ -vinyl proton<sup>9)</sup> and by the hydrogen-bonding with 7-carbonyl group, respectively.

The stereochemistry of the alanine residue was determined by the advanced Marfey's method.<sup>10)</sup> For this analysis two enantiomeric alanine isomers were derivatized with 1-fluoro-2,4-dinitrophenyl-5-L-alaninamide (L-FDAA), and analyzed by reversed-phase HPLC. The retention times of the corresponding enantiomers (2*S* and 2*R*) were observed with 9.6 and 10.6 min, respectively. Analogous derivatization of the acid hydrolyzate of compound **1** followed by HPLC analysis and comparison with the standard derivatives enabled us to deduce 12*S* configuration.<sup>11</sup>

Compounds **2** and **3** have been isolated from the more polar fractions and were identified as neoechinulin A, which was previously isolated as an antioxidative substance from the fungal genera *Aspergillus*,<sup>9,12,13)</sup> and L-alanyl-L-tryptophan anhydride,<sup>14)</sup> respectively.

Compounds 1 and 2 exhibited a significant radical scavenging activity against DPPH with  $IC_{50}$  values of 20 and 24  $\mu$ M, respectively, which are similar to the positive control, ascorbic acid ( $IC_{50}$ , 20  $\mu$ M). Compounds 1 and 2 also showed a UV-A protecting activity with ED<sub>50</sub> values of 90 and 170  $\mu$ M, respectively, which are more active than oxybenzone (ED<sub>50</sub>, 350  $\mu$ M) currently being used as sunscreen.

The further biological evaluation of **1** is in progress.

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## **References and Notes**

- Sorensen D., Larsen T. O., Christophersen C., Nielsen P. H., Anthoni, U., *Phytochemistry*, **51**, 1181–1183 (1999).
- Jiang Z., Boyd K. G., Mearns-Spragg A., Adams D. R., Wright P. C., Burgess J. G., *Nat. Prod. Lett.*, 14, 435–440 (2000).
- Kozlovsky A. G., Vinokurova N. G., Adanin V. M., Burkhardt G., Dahse H., Gräfe U., J. Nat. Prod., 64, 553 (2001).
- Kozlovsky A. G., Marfenina O. G., Vinokurova N. G., Zhelifonova V. P., Adanin V. M., *Mycotoxins*, 48, 37–43 (1998).
- Lee S. M., Li X. F., Jiang H., Cheng J. G., Seong S., Choi H. D., Son B. W., *Tetrahedron Lett.*, 44, 7707–7710 (2003).
- 6) The fungal strain was identified as a *Aspergillus* sp. based on fatty acid methyl ester analysis and growth characteristics (Korean Culture Center of Microorganisms, Seoul, Korea). Their analysis showed a similarity index of 0.62.
- 7) The fungus was cultured (201) for 30 d (static) at 29 °C in SWS medium: soytone (0.1%), soluble starch (1.0%), and seawater (100%).
- 8) Golmaenone (1) was isolated as a yellow solid which showed: mp 160—161 °C (from CHCl<sub>3</sub>). [ $\alpha$ ]<sub>D</sub> +7.1° (c=0.4, CHCl<sub>3</sub>). IR (neat) cm<sup>-1</sup>: 3433, 3242, 1697, 1629, 1578, 1510, 1446, 1378, 1310, 1218, 1164, 1021, 916, 752. UV  $\lambda_{max}$  (CHCl<sub>3</sub>) nm (log  $\varepsilon$ ): 222 (1.8), 247 (2.0), 327 (1.9), 368 (1.7). CD  $\lambda_{max}$  (CHCl<sub>3</sub>) nm ( $\Delta \varepsilon$ ): 230 (+0.5), 239 (-0.2), 253 (-0.4), 305 (+0.1), 346 (-0.1). LR-FAB-MS m/z: 378 [M+Na]<sup>+</sup>, 356 [M+H]<sup>+</sup>; HR-FAB-MS m/z: 378.1428 (Calcd for C<sub>19</sub>H<sub>21</sub>N<sub>3</sub>O<sub>4</sub>Na: 378.1430). See Table 1 for NMR spectral data.
- Marchelli R., Dossena A., Pochini A., Dradi E., J. Chem. Soc. Perkin Trans. 1, 1977, 713–717 (1977).
- Fujii K., Ikai Y., Mayumi T., Oka H., Suzuki M., Harada K.-I., *Anal. Chem.*, **69**, 3346–3352 (1997).
- 11) Samples (0.5 mg) of each compounds 1—3 were subjected to acid hydrolysis with 6 N HCl (1 ml) at 110 °C for 12 h. The hydrolyzates were dried, resuspended in H<sub>2</sub>O (100 μl), and derivatized with L-FDAA. The L-FDAA derivatives, from the hydrolyzates, were compared with similarly derivatized standard amino acids (L-alanine and D-alanine) by HPLC [YMC ODS-A (10×250 mm), 10 μm, flow rate 1 ml/min, UV detection at 340 nm] using an isocratic elution of MeCN–0.1% (v/v) aqueous TFA (1:1). The retention times of L-FDAA derivatives of standard amino acids, L-alanine and D-alanine, were 9.6 and 10.6 min, respectively, and the retention times of L-FDAA derivatives of hydrolyzates were 9.6 min, respectively.
- 12) Neoechinulin A (2) was isolated a colorless solid which showed spectral data virtually identical to that reported in the literature.<sup>9,13)</sup> The NMR data was reassigned as follow: <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ: 8.32 (1H, s, H-1), 7.27 (1H, d, *J*=7.8 Hz, H-4), 7.18 (1H, dd, *J*=7.8, 7.5 Hz, H-5), 7.16 (1H, dd, *J*=7.5, 7.3 Hz, H-6), 7.36 (1H, d, *J*=7.3 Hz, H-7), 7.21 (1H, s, H-8), 7.45 (1H, br s, H-11), 4.30 (1H, qd, *J*=7.0, 1.7 Hz, H-12), 6.40 (1H, s, H-14), 6.07 (1H, dd, *J*=17.5, 10.5 Hz, H-16), 5.23 (1H, d, *J*=10.5 Hz, H<sub>a</sub>-17), 5.19 (1H, d, *J*=17.5 Hz, H<sub>b</sub>-17), 1.53 (6H, s, H<sub>3</sub>-18/19), 1.60 (3H, d, *J*=7.0 Hz, H<sub>3</sub>-20). <sup>13</sup>C-NMR (CDCl<sub>3</sub>) δ: 143.8 (C-2), 102.9 (C-3), 126.0 (C-3a), 118.9 (C-4), 121.0 (C-5), 122.3 (C-6), 111.2 (C-7), 134.3 (C-7a), 111.9 (C-8), 124.5 (C-9), 159.8 (C-10), 51.7 (C-12), 165.7 (C-13), 39.2 (C-15), 144.3 (C-16), 113.3 (C-17), 27.3 (C-18), 27.4 (C-19), 20.9 (C-20).
- 13) Yagi R., Doi M., Biosci. Biotechnol. Biochem., 63, 932-933 (1999).
- 14) Hamasaki T., Nagayama K., Hatsuda Y., Agric. Biol. Chem., 40, 2487 (1976).