Regioselective Synthesis of 6-Alkyl- and 6-Prenylpolyhydroxyisoflavones and 6-Alkylcoumaronochromone Derivatives

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The palladium-catalyzed coupling reaction of 6-iodoisoflavone, prepared from 3'-iodoacetophenone derivative, with 2-methyl-3-butyn-2-ol gave 6-alkynylisoflavone derivative, which was hydrogenated to give 6-alkylhydroxyisoflavone (luteone hydrate) (2). Dehydration of 2 gave 2',4',5,7-tetrahydroxy-6-prenylisoflavone (luteone) (1). Wighteone hydrate (3) was also synthesized from 6-iodotris(benzyloxy)isoflavone in a similar manner. 6-Alkyl-4'5,7-trihydroxy-coumaronochromone (4) was synthesized by oxidative cyclization of 2 with *o*-chloranil.

Key words iodoisoflavone; prenylisoflavone; luteone; regioselective prenylation; coumaronochromone; o-chloranil

Prenyl (=3-methyl-2-butenyl)isoflavones and (3-hydroxy-3-methylbutyl)isoflavones, which contain an alkyl or alkenyl group in the A- and/or B-ring, are widely distributed in nature and have antifungal activity.^{1–4)} Luteone, known as a phytoalexin, was first isolated in 1973 from immature fruits of Lupinus luteus (Leguminosae).⁵⁾ The structure was assigned as 2',4',5,7-tetrahydroxy-6-(3-methyl-2-butenyl)isoflavone (1) by spectroscopic and chemical studies. The same isoflavone 1 was also isolated from healthy leaves or roots of white lupin (Lupinus albus L., cv Kievskij Mutant)^{4,6)} and the roots of yellow lupin (Lupinus leuteus L., cv. Barpin) together with luteone hydrate, the structure of which was assigned to be 2',4',5,7-tetrahydroxy-6-(3-hydroxy-3-methylbutyl)isoflavone (2) by spectroscopic analysis.⁷⁾ Luteone hydrate (2) was isolated as a fungal metabolite of luteone (1) with cultures of Aspergillus flavus and Botrytis cinerea.⁸⁾ Wighteone hydrate [4',5,7-trihydroxy-6-(3-hydroxy-3-methylbutyl)isoflavone] (3) was also isolated as a fungal metabolite of wighteone [4',5,7-trihydroxy-6-(3-methyl-2-butenyl)isoflavone]^{2,6,7,9)} with cultures of A. flavus and B. cinerea.¹⁰⁾ In view of the isolation of 1 and 2 from the same natural source, it is considered that 2 is a precursor of 1 and dehydration of 2 would lead to 1. The total syntheses of isoflavones 1, 2, and 3 have not been achieved yet, although the dimethyl ether of luteone (1) has been synthesized.¹¹⁾ The reason seems to be due to the difficulty in introducing an alkyl or alkenyl group regioselectively into the isoflavone nucleus and the selectivity of protection and consequent deprotection. Furthermore, 6-alkylpolyhydroxyisoflavones are isomerized to the corresponding isomers, 8-alkylpolyhydroxyisoflavones, by bases.^{12,13} We need to solve these problems for the regioselective synthesis of these phloroglucin-type 6-prenylisoflavones. In our previous paper,¹⁴⁾ we reported the regioselective synthesis of prenylisoflavones. As a continuation of our studies on the regioselective synthesis of alkyl- and prenylisoflavones, we wish to report here on the first syntheses of 1, 2, and 3 using the palladium (0)-catalyzed coupling reaction¹⁵⁾ of the corresponding iodoisoflavone with 2-methyl-3-butyn-2-ol.¹⁶) Recently, the new coumaronochromones (=benzofuro[2,3-*b*][1]benzopyran-11-ones) lupilutin (8-alkylcoumaronochromone from the root of yellow lupin)⁷⁾ and lupinalbin B (6-prenylcoumaronochromone from the root of white lupin)¹⁷⁾ have also been isolated. However, 6-alkylcoumaronochromone (4), which is considered to be a precursor of lupinalbin B, has yet to be isolated from natural sources. We have examined the simple and general applicability of DDQ or *o*-chloranil to the synthesis of alkylpolyhydroxycoumaronochromones from the corresponding 2'-hydroxyisoflavones.¹⁸⁾ We wish to report here the synthesis of compound 4 by oxidative cyclization of compound 2 with *o*-chloranil.

Results and Discussion

The catalytic hydrogenation of 2',4'-bis(benzyloxy)-6'methoxymethoxyacetophenone¹⁴⁾ over Pd/C, followed by iodination of the resulting 2',4'-dihydroxyacetophenone 5 with I_2 and $H_5IO_6^{(19)}$ gave the 3'-iodoacetophenone **6** in 92% yield. Compound 6 was converted into bis(benzyloxy)acetophenone 7, the structure of which was determined by direct comparison with a sample of the isomer [2',4'-bis(benzyloxy)-5'iodo-6'-methoxymethoxyacetophenone (mp 99-100 °C)¹⁴] and ¹H-NMR-NOE analysis. The mixture of 7 with the isomer showed a marked decrease in the melting point relative to that of each compound. Compound 7 was not obtainable by the I₂-CF₃CO₂Ag method.¹⁴⁾ Condensation of 7 with 2,4bis(benzyloxy)benzaldehyde in the presence of sodium hydroxide gave 6'-methoxymethoxychalcone 8, and then the methoxymethyl group in the chalcone was cleaved by treatment with dilute HCl to give 3'-iodo-6'-hydroxychalcone 9 in 86% yield. Oxidative rearrangement of acetate 10, prepared from 9, with thallium(III) nitrate trihydrate (TTN),²⁰⁾ followed by hydrolysis of the resulting mixture 11 with aqueous sodium hydroxide gave the desired 6-iodoisoflavone 12 in 40% yield and the chalcone 9, the structures of which were identified by ¹H-NMR spectral analysis. On the basis of the results, it was shown that deacetylation of 10 with TTN took place more easily than the oxidative rearrangement of the phenyl group of 10 to give the chalcone 9. Therefore 9 was converted into benzoate 13, which was oxidatively rearranged with TTN to give the corresponding acetal 14 easily. The crude acetal 14 was hydrolyzed with aqueous sodium hydroxide to give the 6-iodoisoflavone 12 in 70% yield via two steps from 13. The coupling reaction of 12 with 2methyl-3-butyn-2-ol in the presence of Pd(0) in triethylamine gave 6-(3-hydroxy-3-methylbutynyl)isoflavone 15 in 71% yield. The catalytic hydrogenation of 15 gave 2',4',5,7-

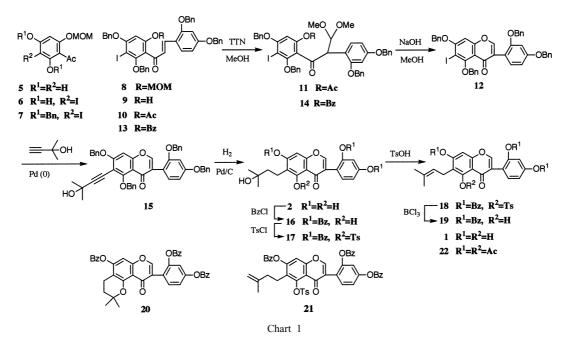


Table 1. ¹H-NMR (400 MHz, CD₃COCD₃) Data for Prenyl- and Alkylisoflavones 1, 2, Luteone, and Luteone Hydrate^a)

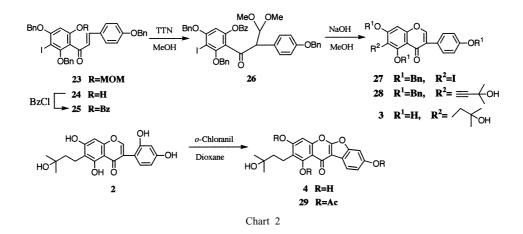
Compound	2-Н	8-H	3'-Н	5'-H	6'-Н 2'-Н	Me	CH ₂	CH=C	ОН
1	8.14 s	6.53 s	6.48 d (<i>J</i> =2.4)	6.44 dd (J=2.4, 8.3)	7.12 d (J=8.3)	1.65 s 1.78 s	3.37 d (<i>J</i> =7.3)	5.28 t (J=7.3)	8.31 br s, 8.43 br s 9.23 br s, 13.06 s
Natural product ⁷⁾ (1)	8.14 s	6.53 s	6.48 d (Incomplete)	6.43 dd (J=2.4, 8.9)	7.12 d (<i>J</i> =8.9)	1.65 s 1.78 s	3.37 br d (<i>J</i> =7.3)	5.28 br t $(J=7)$	13.05 s
2	8.15 s	6.52 s	6.49 d (<i>J</i> =2.4)	6.44 dd (<i>J</i> =2.4, 8.3)	7.12 d (<i>J</i> =8.3)	1.26 s (6H)	1.71 m 2.79 m		3.59 br s 8.32 s, 8.43 s 9.29 br s, 13.05 s
Natural product ⁷⁾ (2)	8.14 s	6.51 s	6.48 d (Incomplete)	6.43 dd (<i>J</i> =2.4, 8.8)	7.12 d (<i>J</i> =8.8)	1.25 s (6H)	1.71 m 2.81 m		13.04 s
3	8.15 s	6.47 s	6.90 d (<i>J</i> =8.8)	6.90 d (<i>J</i> =8.8)	7.45 d (J=8.8) 7.45 d (J=8.8)	1.26 s (6H)	1.71 m 2.78 m		8.50 br s, 9.65 br s 13.33 s

a) s, singlet; d, doublet; dd, double doublet; t, triplet; br, broad; m, multiplet.

tetrahydroxy-6-(3-hydroxy-3-methylbutyl)isoflavone (2) in 96% yield. The ¹H-NMR spectrum of **2** was identical to that of a natural sample of luteone hydrate⁷⁾ (Table 1) and other physical properties (see Experimental). On the basis of these results, the structure of luteone hydrate was confirmed by the first synthesis of 2',4',5,7-tetrahydroxy-6-(3-hydroxy-3-methylbutyl)isoflavone (**2**).

Exhaustive benzoylation (7h) of **2** afforded partly the isomer [2',4',7-tris(benzoyloxy)-5-hydroxy-8-(3-hydroxy-3-methylbutyl)isoflavone]. To prevent the isomerization, the partial benzoylation of compound **2** gave 2',4',7-tris(benzoyloxy)isoflavone **16** for 30 min in 85% yield, and subsequently compound **16** was tosylated for 20 min to give 5-tosyloxy-isoflavone **17** in 91% yield. Compound **17** was dehydrated with BF₃·OEt₂ at room temperature to give 6-prenyl-5-tosyloxyisoflavone **18** (20%), 5-hydroxy-6-prenylisoflavone **19** (25%), and dihydropyran derivative **20** (45%), respectively. In this reaction, it was shown that part of **18** was initially detosylated, and the resulting compound **19** was subsequently cyclized to give **20**. The formation of **20** strongly supported the structure of **2** and decreased the yield of **19**.

The tosylate 17 was dehydrated with TsOH \cdot H₂O to give a mixture of 6-prenylisoflavone 18 and the regioisomer 6-(3methyl-3-butenyl)isoflavone 21. The ¹H-NMR spectrum of the tosylate mixture (18 and 21) showed the ratio of 18 to 21 to be 85:15 [peaks due to CH₂CH=C(CH₃)₂ at δ =3.36 (2H, d) and CH₂CH₂C(CH₂)=CH₂ at δ =4.57 (2H, s)]. The mixture (18 and 21) reacted quantitatively with benzohydroximoyl chloride¹⁴⁾ in dry CH₂Cl₂ at room temperature to give a mixture of the unchanged 6-prenylisoflavone 18 and the terminal alkene-cyclic adduct, and then 18 was purified by silica gel column chromatography. The detosylation of 18 with BCl₃, followed by hydrolysis of the resultant compound 19 with 10% NaOH in a mixture of methanol and dioxane at room temperature, gave 2',4',5,7-tetrahydroxy-6-(3-methyl-2-butenyl)isoflavone (1) in 66% yield (¹H-NMR in Table 1), which was converted into the tetraacetate derivative 22. The ¹H-NMR, IR, and UV spectral data for 1 were completely identical to those of a natural sample of luteone.^{5,6)} On the basis of these results, the structure of luteone was confirmed for the first time by the synthesis of 2',4',5,7-tetrahydroxy-6-(3-methyl-2-butenyl)isoflavone (1).



Condensation of 7 with 4-benzyloxybenzaldehyde and the subsequent hydrolysis of the resultant chalcone 23 gave 6'hydroxychalcone 24, which was converted into the corresponding benzoate 25. The oxidative rearrangement of 25 with TTN for 20 h, followed by the hydrolysis of the resultant acetal 26, gave the corresponding 4'-benzyloxy-6-iodoisoflavone 27 in 30% yield via three steps from 24. The yield of compound 27 was lower than that of the 2',4'-bis(benzyloxy)-6-iodoisoflavone 12. The reason depends on the number of the electron-releasing subtituents in the B-ring.^{12,21)} The coupling reaction of 27 with 2-methyl-3-butyn-2-ol in the presence of Pd(0) gave 6-(3-hydroxy-3-methyl-1-butynyl)isoflavone 28 in 57% yield. The catalytic hydrogenation of 28 over Pd/C gave 4',5,7-trihydroxy-6-(3-hydroxy-3methylbutyl)isoflavone (3) in 92% yield. The ¹H-NMR spectrum and other physical properties were identical to those of a natural sample of wighteone hydrate¹⁰ (Table 1). On the basis of these results, the structure of wighteone hydrate was confirmed by the synthesis of 4',5,7-trihydroxy-6-(3-hydroxy-3-methylbutyl)isoflavone (3).

The oxidative cyclization of the 2'-hydroxyisoflavone **2** with *o*-chloranil (2.2 eq) [the reduction potential (0.83 V) is lower than that (1.0 V) of DDQ]^{22,23)} in dioxane gave the desired coumaronochromone **4** at 80 °C for 2 h in good yield. Compound **4** was easily converted into diacetate **29** at 0 °C using the acetic anhydride-pyridine method.

The present regioselective synthesis of iodoisoflavones and the palladium (0)-catalyzed coupling reaction of iodoisoflavones with 2-methyl-3-butyn-2-ol have been shown to be efficient and useful procedures for the syntheses of prenyl- and alkylpolyhydroxyisoflavones and *O*-alkylated prenylisoflavones.

Experimental

All the melting points were measured on a Yanaco MP-J3 micro meltingpoint apparatus and are uncorrected. The ¹H-NMR spectra were measured with a JEOL EX400 spectrometer (400 MHz), using tetramethylsilane as internal standard (δ , ppm). The IR spectra were recorded on a Hitachi 215 spectrophotometer, and the UV spectra were recorded on a Hitachi 124 spectrophotometer. Elemental analyses were performed with a Yanaco CHN corder model MT-5. Column chromatography and thin-layer chromatography (TLC) were carried out on Kieselgel 60 (70–230 mesh) and Kieselgel 60 F-254 (Merck).

2',4'-Dihydroxy-6'-methoxymethoxyacetophenone (5) 2',4'-Bis(benzyloxy)-6'-methoxymethoxyacetophenone (4.5 g, 11.46 mmol) was hydrogenolyzed over Pd/C (5%) (450 mg) in MeOH (100 ml) and AcOEt (100 ml) at 18—23 °C until uptake of hydrogen ceased. After removal of the solvent under reduced pressure, the resulting compound was purified by silica gel column chromatography (AcOEt : hexane=2:1 as a solvent) and recrystallized from a mixture of AcOEt and hexane to give dihydroxyace-tophenone **5** (2.28 g, 94%) as colorless needles, mp 118—120 °C; ¹H-NMR (CDCl₃) δ =2.65 (3H, s, COCH₃), 3.52 (3H, s, OCH₃), 5.25 (2H, s, OCH₂O), 5.25 (1H, s, C₄-OH), 6.04 (1H, d, *J*=2.4 Hz, Ar-H), 6.14 (1H, d, *J*=2.4 Hz, Ar-H), 13.79 (1H, s, C₆-OH). *Anal.* Calcd for C₁₀H₁₂O₅: C, 56.60; H, 5.70. Found: C, 56.61; H, 5.60.

2',**4'-Dihydroxy-3'-iodo-6'-methoxymethoxyacetophenone (6)** The acetophenone **5** (3.16 g, 14.8 mmol) was dissolved in EtOH (30 ml), and iodine (1.88 g, 7.4 mmol) and periodic acid (674 mg, 3 mmol) in water (10 ml) were added to the solution; the mixture was then stirred for 30 min at 40 °C. The mixture was cooled and diluted with water to give compound **6** as colorless needles (4.63 g, 92%), mp 160—161 °C; ¹H-NMR (CDCl₃) δ =2.69 (3H, s, COCH₃), 3.52 (3H, s, OCH₃), 5.28 (2H, s, OCH₂O), 5.98 (1H, s, C₄-OH), 6.44 (1H, s, C₅-H), 14.97 (1H, s, C₂-OH). *Anal.* Calcd for C₁₀H₁₁IO₅: C, 35.52; H, 3.28. Found: C, 35.23; H, 3.17.

2',4'-Bis(benzyloxy)-3'-iodo-6'-methoxymethoxyacetophenone (7) To a mixture of **6** (1.0 g, 2.95 mmol), K₂CO₃ (2.03 g, 15 mmol), and DMF (8 ml), a solution of benzyl chloride (0.75 ml, 6.5 mmol) in DMF (1 ml) was added dropwise with stirring under nitrogen at 70 °C for 30 min. The reaction mixture was extracted with CHCl₃, washed with diluted HCl and water, and dried (Na₂SO₄). The resulting compound was recrystallized from a mixture of MeOH and AcOEt to yield **7** (623 mg, 80%) as colorless needles, mp 96—98 °C; ¹H-NMR (CDCl₃) δ =2.47 (3H, s, COCH₃), 3.46 (3H, s, OCH₃), 4.97 (2H, s, ArCH₂O), 5.15 (2H, s, OCH₂O), 5.18 (2H, s, ArCH₂O), 6.65 (1H, s, C₅--H), 7.32—7.60 (10H, m, Ar-H×10). *Anal.* Calcd for C₂₄H₂₃IO₅: C, 55.61; H, 4.47. Found: C, 55.66; H, 4.48.

2,4,2',4'-Tetrakis(benzyloxy)-3'-iodo-6'-methoxymethoxychalcone (8) and 2,4,2',4'-Tetrakis(benzyloxy)-6'-hydroxy-3'-iodochalcone (9) A mixture of the acetophenone 7 (1.84 g, 3.54 mmol) and 2,4-bis(benzyloxy)benzaldehyde (1.70 g, 5.3 mmol) was stirred in the presence of KOH (2.0 g, 35 mmol) in EtOH (120 ml) at 80 °C for 45 min. Ice-water and 10% HCl were added to the reaction mixture to give 6'-methoxymethoxychalcone 8 as yellow precipitates. The collected crude solid 8 was dissolved in a mixture of CHCl₂ (60 ml) and MeOH (60 ml). Concentrated HCl (3 ml) was added to the solution, and then the mixture was stirred at 40 °C for 1 h. The whole solution was extracted with CHCl₃, and the chloroform extract was washed with water and dried (Na2SO4). After removal of the solvent, the resulting compound was recrystallized from a mixture of CHCl3 and MeOH to give 6'-hydroxychalcone 9 (2.35 g, 86% from 7) as yellow needles, mp 158-160 °C; ¹H-NMR (CDCl₃) δ =4.82, 5.00, 5.07, and 5.20 (each 2H, s, PhCH₂), 6.38 (1H, dd, J=2.4 and 8.8 Hz, C₅-H), 6.41 (1H, s, C₅-H), 6.54 (1H, d, J=2.4 Hz, C₃-H), 7.1-7.53 (21H, m, Ar-H×20, C₆-H), 7.90, and 8.29 (each 1H, d, J=15.6 Hz, CH=), 13.77 (1H, s, C2-OH). Anal. Calcd for C43H35IO6: C, 66.67; H, 4.55. Found: C, 66.43; H, 4.62.

6'-Acetoxy-2,4,2',4'-tetrakis(benzyloxy)-3'-iodochalcone (10) and 2',4',5,7-Tetrakis(benzyloxy)-6-iodoisoflavone (12) The chalcone **9** (540 mg, 0.70 mmol) was converted into acetate **10** at 70 °C for 30 min using an acetic anhydride (20 ml)-pyridine (2 ml) method. After the addition of water to the reaction mixture, the whole mixture was extracted with CHCl₃ and the extract was washed with water and dried (Na₂SO₄). After the resulting acetate **10** (530 mg) and TTN (420 mg, 0.9 mmol) were stirred in a solution of MeOH (25 ml) and CHCl₃ (10 ml) at 30 °C for 4h, 10% HCl (15 ml) was added to the reaction mixture, and the whole solution was stirred at room temperature for 1 h to give white precipitates. After removal of the precipi tates by filtration, the filtrate was extracted with CHCl₃, and the extract was washed with water and dried (Na₂SO₄). The resulting crude acetal **11** in dioxane (10 ml) and MeOH (10 ml) was stirred with 10% aqueous NaOH (8 ml) at room temperature for 2 h, and then water and 10% HCl were added to the reaction mixture to give precipitates. The collected precipitates were extracted with CHCl₃, washed with water, and dried (Na₂SO₄). The resulting compound was chromatographed over a silica gel flash column (CHCl₃ as a solvent) to give the isoflavone **12** (218 mg, 40% from **9**), and recrystallized from a solution of MeOH and CHCl₃) as pale yellow needles, mp 174–176.5 °C.

2,4,2',4'-Tetrakis(benzyloxy)-6'-benzoyloxy-3'-iodochalcone (13) A mixture of the chalcone **9** (3.0 g, 3.87 mmol), benzoyl chloride (0.68 ml, 5.9 mmol), and K₂CO₃ (2.68 g, 19 mmol) in DMF (35 ml) was stirred under nitrogen at 60 °C for 30 min. After removal of K₂CO₃ and the solvent under reduced pressure, the residue was extracted with CHCl₃, washed with 10% HCl and water, and dried (Na₂SO₄). The resulting compound was purified by silica gel column chromatography (CHCl₃: hexane=10:1 as a solvent) and further recrystallized from MeOH–Me₂CO to give **13** (3.18 g, 92%) as pale yellow needles, mp 126–128 °C; ¹H-NMR (CDCl₃) δ =4.99, 5.01, 5.04, and 5.18 (each 2H, s, Ph<u>CH</u>₂), 6.51 (1H, s, C₃-H), 6.53 (1H, s, C₅-H), 6.77 (1H, s, C₅-H), 7.03 and 7.79 (each 1H, d, J=16.1 Hz, CH=), 7.20–8.10 (25H, m, Ar-H×25). *Anal.* Calcd for C₅₀H₃₉IO₇: C, 68.34; H, 4.47. Found: C, 68.13; H, 4.70.

1-[6-Benzoyloxy-2,4-bis(benzyloxy)-3-iodophenyl]-2-[2,4-bis(benzyloxy)phenyl]-3,3-dimethoxypropan-1-one (14) and 2,4,2',4'-Tetrakis(benzyloxy)-6-iodoisoflavone (12) A mixture of the chalcone 13 (1.61 g, 1.83 mmol) and TTN (1.24g, 2.8 mmol) was stirred in a solution of MeOH (150 ml) and CHCl₃ (60 ml) at 40 °C for 4 h, then 10% HCl (35 ml) was added to the mixture at room temperature and the whole was further stirred at that temperature for 1 h to give white precipitates. After removal of the precipitates by filtration, the filtrate was extracted with CHCl₂ and the extract was washed with water and dried (Na2SO4). The organic solvent was removed under reduced pressure, and the resulting crude acetal 14 was dissolved in a mixture of dioxane (80 ml) and MeOH (100 ml), and then hydrolyzed with 10% aqueous NaOH at room temperature for 2 h. Water and 10% HCl were added to the reaction mixture to give precipitates. The collected precipitates were extracted with CHCl₃, washed with water, and dried (Na₂SO₄). The resulting compound was chromatographed over a silica gel column (CHCl₃ as a solvent) to give the 6-iodoisoflavone 12, which was recrystallized from a mixture of MeOH and CHCl₃ as pale yellow needles (995 mg, 70% from 13), mp 174—176 °C; ¹H-NMR (CDCl₃) δ =5.02, 5.03, 5.05, and 5.25 (each 2H, s, Ph<u>CH₂</u>), 6.63 (1H, dd, J=2.4 and 8.5 Hz, C_{5'}-H), 6.67 (1H, d, J=2.4 Hz, C₃-H), 6.73 (1H, s, C₈-H), 7.20–7.75 (21H, m, Ar-H×21), 7.78 (1H, s, C2-H). Anal. Calcd for C43H33IO6: C, 66.85; H, 4.30. Found: C, 66.64; H, 4.58.

Acetal **14**: mp 56—58 °C; ¹H-NMR (CDCl₃) δ =2.92 and 3.14 (each 3H, s, OCH₃), 4.7—5.0 (6H, m, Ph<u>CH₂</u>×3), 5.09 (2H, s, Ph<u>CH₂</u>), 5.15 and 5.47 (each 1H, d, *J*=8.8 Hz, CH), 6.25 (1H, dd, *J*=3 and 8.6 Hz, Ar-H), 6.37 (1H, d, *J*=3 Hz, Ar-H), 6.54 (1H, s, Ar-H), 7.09 (1H, d, *J*=8.6 Hz, Ar-H), 7.05—8.15 (25H, m, Ar-H×25).

2',**4'**,**5**,**7-Tetrakis(benzyloxy)-6-(3-hydroxy-3-methyl-1-butynyl)isoflavone (15)** To a solution of **12** (1.5 g, 1.94 mmol) and 2-methyl-3-butyn-2-ol (0.56 ml, 6 mmol) in a mixture of NEt₃ (25 ml) and DMF (9 ml), were added PdCl₂ (17 mg, 0.1 mmol), PPh₃ (51 mg, 0.19 mmol), and CuI (18 mg, 0.1 mmol); the mixture was then stirred under nitrogen at 75 °C for 2 h. The reaction mixture was filtered through charcoal and the filtrate was concentrated under reduced pressure and extracted with AcOEt; the extract was then washed with 2% HCl and water and dried (Na₂SO₄). The resulting compound was purified by silica gel column chromatography (CHCl₃: AcOEt =10:1 as a solvent) and further recrystallized from a mixture of MeOH and Me₂CO to give **15** (1.32 g, 71%) as colorless prisms, mp 173—174°C; ¹H-NMR (CDCl₃) δ =1.51 (6H, s, CH₃×2), 5.04, 5.06, 5.15, and 5.20 (each 2H, s, Ph<u>CH₂</u>), 6.68 (1H, d, J=2.4 Hz, C₃-H), 6.69 (1H, s, C₈-H), 7.20—7.70 (21H, m, Ar-H×21), 7.77 (1H, s, C₂-H). *Anal.* Calcd for C₄₈H₄₀O₇: C, 79.10; H, 5.53. Found: C, 78.86; H, 5.63.

2',4',5,7-Tetrahydroxy-6-(3-hydroxy-3-methylbutyl)isoflavone (Luteone Hydrate) (2) The isoflavone 15 (2.31 g, 3.17 mmol) was hydrogenolyzed over Pd/C (5%) (400 mg) in MeOH (120 ml) and dioxane (100 ml) at room temperature until uptake of hydrogen ceased. After removal of the solvent under reduced pressure, the resulting compound was recrystallized from a mixture of MeOH and CH₂Cl₂ to give 2 (1.13 g, 96%) as pale yellow prisms, mp 229–231 °C; IR (KBr) v 3350, 2975, 1645, 1620, 1460, 1310, 1065, and 830 cm⁻¹; UV λ_{max} nm (log ε) (MeOH) 265 (4.45), 290sh (4.19), and 347sh (3.58); (+AlCl₃) 208sh (3.53), 240sh (4.10), and 267 (4.46);

(+NaOAc) 273sh, 269 (4.44), and 340 (3.93). Anal. Calcd for $C_{20}H_{20}O_7$: C, 64.51, H, 5.41. Found: C, 64.22; H, 5.46.

2',4**'**,7-**Tris(benzoyloxy)-5-hydroxy-6-(3-hydroxy-3-methylbutyl)isoflavone (16)** A mixture of **2** (420 mg, 1.12 mmol), benzoyl chloride (0.47 ml, 4 mmol), and K₂CO₃ (1.55 g, 11 mmol) in Me₂CO (20 ml) was refluxed with stirring under nitrogen for 30 min. After removal of K₂CO₃ and the solvent, the residue was extracted with ethyl acetate, washed with 5% HCl and water, and dried (Na₂SO₄). The resulting compound was purified by silica gel column chromatography (CHCl₃:Me₂CO=10:1 as a solvent) and further recrystallized from a mixture of MeOH and AcOEt to give 16 (650 ml, 85%) as pale yellow needles, mp 154—155 °C; ¹H-NMR (CDCl₃) δ =1.19 (6H, s, CH₃×2), 1.69 and 2.72 (each 2H, m, CH₂), 6.84 (1H, s, C₈-H), 7.26—7.70 (13H, m, Ar-H×13), 7.98 (1H, s, C₂-H), 8.1—8.25 (5H, m, Ar-H×5), 12.92 (1H, s, C₅-OH). *Anal.* Calcd for C₄₁H₃₂O₁₀: C, 71.92; H, 4.71. Found: C, 71.91; H, 4.78.

2',4',7-Tris(benzoyloxy)-6-(3-hydroxy-3-methylbutyl)-5-tosyloxyisoflavone (17) A mixture of 16 (820 mg, 1.19 mmol), TsCl (342 mg, 1.8 mmol), and K₂CO₃ (1.66 g, 12 mmol) in Me₂CO (35 ml) was refluxed with stirring under nitrogen for 20 min. After removal of K₂CO₃ and the solvent, the residue was extracted with AcOEt, washed with 5% HCl and water, and dried (Na₂SO₄). The resulting compound was recrystallized from a mixture of CHCl₃ and MeOH to give 17 (906 mg, 91%) as colorless needles, mp 116—119 °C; ¹H-NMR (CDCl₃) δ =1.13 (6H, s, CH₃×2), 1.39 (1H, s, OH), 1.71 and 2.82 (each 2H, m, CH₂), 2.45 (3H, s, Ar-CH₃), 7.26—7.70 (15H, m, Ar-H×15), 7.87 (1H, s, C₂-H), 7.9—8.2 (8H, m, Ar-H×8). *Anal.* Calcd for C₄₈H₃₈O₁₂S: C, 68.73; H, 4.57. Found: C, 68.47; H, 4.76.

Dehydration of 2',4',7-Tris(benzoyloxy)-6-(3-hydroxy-3-methylbutyl)-5-tosyloxyisoflavone (17) with Boron Trifluoride Diethyl Etherate To a solution of 17 (50 mg, 0.06 mmol) in dry CH₂Cl₂ (2 ml) was added a solution of BF₃·OEt₂ (0.05 mmol) in CH₂Cl₂ (0.4 ml); the mixture was stirred under nitrogen at room temperature for 2 h. To the reaction mixture was added aqueous NH₄Cl, and the whole solution was extracted with chloroform, washed with water, and dried (Na₂SO₄). The resulting compound was chromatographed over a silica gel column (CHCl₃ as a solvent) to give 5-tosyloxyisoflavone 18 [10 mg (20%) as colorless needles, mp 188—189 °C], 5hydroxyisoflavone 19 [10 mg (25%) as pale yellow needles, mp 177— 179 °C], and dihydropyranoisoflavone 20 [18 mg (45%) as colorless prisms], mp 187—190 °C; ¹H-NMR (CDCl₃) δ =1.41 (6H, s, CH₃×2), 1.81 and 2.68 (each 2H, t, *J*=6.8 Hz, CH₂), 6.81 (1H, s, C₈-H), 7.2—7.7 (12H, m, Ar-H), 7.81 (1H, s, C₂-H), 8.09—8.23 (6H, m, Ar-H). *Anal.* Calcd for C₄₁H₃₀O₉: C, 73.87; H, 4.54. Found: C, 73.65; H, 4.80.

2',4',7-Tris(benzoyloxy)-6-(3-methyl-2-butenyl)-5-tosyloxyisoflavone (18) To a solution of 17 (1.0 g, 1.19 mmol) in dry toluene (6 ml) was added TsOH \cdot H₂O (1.81 ml of a 5.25×10⁻¹ mol dm⁻³ in acetic acid); the mixture was stirred under nitrogen at 110 °C for 45 min. The reaction mixture was extracted with ether, washed with 5% aqueous NaHCO₃, 2% HCl, and water, and dried (Na2SO4). The resulting compound was chromatographed over a silica gel-flashed column (CHCl₃ as a solvent) to give a mixture of 6alkenylisoflavones (745 mg) as colorless needles. The ¹H-NMR spectrum of the mixture was shown to be an 85:15 mixture of the 6-(3-methyl-2butenyl)isoflavone 18 and the isomer 6-(3-methyl-3-butenyl)isoflavone 21. The mixture (18 and 21) in CH₂Cl₂ (0.5 ml) was added to a solution of benzohydroximoyl chloride (63 mg, 0.4 mmol) and NEt₃ (0.13 ml, 0.94 mmol) in CH₂Cl₂ (2 ml) in an ice-bath, and then the reaction mixture was stirred at room temperature for 20 h. The reaction mixture was quenched with saturated aqueous NH4Cl and extracted with CH2Cl2, washed with water, and dried (Na₂SO₄). The resulting compound was chromatographed over a silica gel column (CHCl₃: Me₂CO=15:1 as a solvent) to give the 6prenylisoflavone 18, which was recrystallized from a mixture of CHCl₃ and hexane as colorless needles (500 mg, 68% from 17), mp 188-189 °C; ¹H-NMR (CDCl₃) δ =1.38 and 1.45 (each 3H, s, CH₃), 2.44 (3H, s, Ar-CH₃), 3.36 (2H, d, J=6.4 Hz, CH₂), 4.92 (1H, brt, J=6.4 Hz, =CH), 7.2-7.7 (15H, m, Ar-H×15), 7.86 (1H, s, C₂-H), 8.0-8.2 (8H, m, Ar-H×8). Anal. Calcd for C₄₈H₃₆O₁₁S: C, 70.23; H, 4.42. Found: C, 70.19; H, 4.59.

2',4',7-**Tris(benzoyloxy)-5-hydroxy-6-(3-methyl-2-butenyl)isoflavone** (19) A mixture of 18 (160 mg, 0.19 mmol) and BCl₃ (0.14 ml; 1 mol solution: Aldrich) on dry CH₂Cl₂ (2 ml) was stirred under argon at 15 °C for 15 min. The reaction mixture was quenched with saturated NH₄Cl and extracted with CH₂Cl₂, washed with water, and dried (Na₂SO₄). The resulting compound was purified by silica gel column chromatography (CHCl₃ as a solvent) and crystallized fron CHCl₃-hexane to give 19 (123 mg, 95%) as pale yellow needles, mp 177—179 °C; ¹H-NMR (CDCl₃) δ =1.55 and 1.59 (each 3H, s, CH₃), 3.34 (2H, d, J=7.3 Hz, CH₂), 5.12 (1H, t, J=7.3 Hz, =CH), 6.80 (1H, s, C₈-H), 7.3—7.7 (15H, m, Ar-H×15), 7.97 (1H, s, C₂- H), 8.1–8.2 (6H, m, Ar-H×6), 12.90 (1H, s, C_5 -OH). Anal. Calcd for $C_{41}H_{30}O_9$: C, 73.87; H, 4.54. Found: C, 73.73; H, 4.76.

2',4',5,7-Tetrahydroxy-6-(3-methyl-2-butenyl)isoflavone (Luteone) (1) Compound 19 (150 mg, 0.22 mmol) in MeOH (2 ml) and dioxane (2 ml) was hydrolyzed with 10% aqueous NaOH (2 ml) under argon at room temperature for 1 h. To the reaction mixture water and diluted HCl were added; the organic solvent was then evaporated under reduced pressure. The residue was extracted with ether, washed with water, and dried (Na₂SO₄). The compound was chromatographed over a silica gel column (CHCl₃ : AcOEt=2 : 1 as a solvent) to give 6-prenylisoflavone 1 (52 mg, 66%), which was crystallized from CH₂Cl₂-hexane as pale yellow prisms, mp 223—225°C; IR (KBr) v 3425, 3300, 3100 br, 1650, 1615, 1590, 1550, 1215, 1060, 815 cm⁻¹; UV λ_{max} nm (log ε) (MeOH) 266 (4.56), 280 (4.33), 340 (3.63), (+AlCl₃) 271 (4.41), (+NaOAc) 269 (4.55), 340 (3.83). *Anal.* Calcd for C₂₀H₁₈O₆: C, 67.79; H, 5.12. Found: C, 67.55; H, 5.21.

2',4',5,7-Tetraacetoxy-6-(3-methyl-2-butenyl)isoflavone (22) Compound 1 (159 mg, 0.45 mmol) was converted into tetraacetate 22 by treatment with acetic anhydride (2 ml)-pyridine (0.3 ml) at 110 °C for 2 h. The resulting compound was purified by silica gel column chromatography (AcOEt: hexane=2:1 as a solvent) to give 22 as colorless pastes (188 mg, 80%); ¹H-NMR (CDCl₃) δ =1.68 and 1.75 (each 3H, s, CH₃), 2.16, 2.30, 2.36, and 2.40 (each 3H, s, COCH₃), 3.23 (2H, br d, CH₂), 5.00 (1H, br t, =CH), 7.04—7.29 (4H, m, Ar-H), 7.80 (1H, s, C₂-H).

4,2',4'-Tris(benzyloxy)-6'-hydroxy-3'-iodochalcone (24) A mixture of the acetophenone 7 (300 mg, 0.58 mmol) and 4-benzyloxybenzaldehyde (188 mg, 0.9 mmol) in EtOH (50 ml) was stirred in the presence of KOH (330 mg, 6 mmol) at 80 °C for 1 h. The resulting 6'-methoxymethoxychalcone **23** was worked up in the same manner as in the case of the 6'-hydroxy-chalcone **8** to 6'-hydroxy-4-benzyloxychalcone **24**, which was recrystallized from a mixture of CHCl₃ and MeOH as yellow needles (334 mg, 87% *via* two steps from 7), mp 124—125 °C; ¹H-NMR (CDCl₃) δ =4.86, 5.10 and 5.21 (each 2H, s, Ph<u>CH₂</u>), 6.24 (1H, s, C₅-H), 6.84 (2H, d, *J*=8.8 Hz, C₃-and C₅-H), 6.99—7.53 (17H, m, Ar-H), 7.83 and 7.88 (each 2H, d, *J*=15.4 Hz, CH=), 13.57 (1H, s, C₆-OH). *Anal.* Calcd for C₃₆H₂₉IO₅: C, 64.68; H, 4.37. Found: C, 64.47; H, 4.61.

4',5,7-Tris(benzyloxy)-6-iodoisoflavone (27) A mixture of 24 (300 mg, 0.45 mmol), benzoyl chloride (0.08 ml, 0.7 mmol), and K₂CO₃ (433 mg, 3.1 mmol) in DMF (8 ml) was stirred under the same conditions as in the case of the benzoate 13 to give 6'-benzoyloxy-3'-iodochalcone 25. A mixture of 25 and TTN (300 mg, 0.68 mmol) was stirred in a solution of MeOH (25 ml) and CHCl₃ (10 ml) at 40 °C for 20 h. The resulting crude acetal 26 in dioxane (10 ml) and MeOH (10 ml) was hydrogenated with 10% aqueous NaOH at room temperature for 2 h. The reaction mixture was worked up in the same manner as in the case of the 6-iodoisoflavone 12 to give 4',5,7tris(benzyloxy)-6-iodoisoflavone 27, which was recrystallized from a mixture of CHCl₃ and MeOH as pale yellow needles (86 mg, 30% via three steps from 24), mp 154—157 °C; ¹H-NMR (CDCl₃) δ =5.07, 5.10 and 5.27 (each 2H, s, PhCH₂), 6.75 (1H, s, C₈-H), 7.04 (2H, d, J=8.8 Hz, C_{3'}- and C5'-H), 7.30 (15H, m, Ar-H), 7.77 (2H, d, J=8.8 Hz, C2'- and C6'-H), 7.81 (1H, s, C2-H). Anal. Calcd for C37H27IO5: C, 64.87; H, 4.08. Found: C, 64.65: H. 4.23

4',5,7-Tris(benzyloxy)-6-(3-hydroxy-3-methylbutynyl)isoflavone (28) To a solution of 27 (640 mg, 0.96 mmol) and 2-methyl-3-butyn-2-ol (0.28 ml, 3 mmol) in a mixture of NEt₃ (15 ml) and DMF (5 ml) were added PdCl₂ (8.5 mg, 0.05 mmol), PPh₃ (25 mg, 0.1 mmol), and CuI (9.1 mg, 0.05 mmol); the mixture was then stirred under nitrogen at 80 °C for 1 h. The reaction mixture was worked up in the same manner as in the case of the 6-alkynylisoflavone 15 to give 6-(3-hydroxy-3-methylbutynyl)isoflavone 28, which was recrystallized from a mixture of MeOH and Me₂CO as colorless needles (340 mg, 57%), mp 171–173 °C; ¹H-NMR (CDCl₃) δ =1.50 (6H, s, CH₃×2), 1.78 (1H, s, OH), 5.10 and 5.21 (6H, s, Ph<u>CH₂×3</u>), 6.72 (1H, s, C₈-H), 7.03 (2H, d, J=8.3 Hz, C₃- and C₅-H), 7.30–7.52 (15H, m, Ar-H), 7.67 (2H, d, J=8.3 Hz, C₂- and C₆-H), 7.79 (1H, s, C₂-H). Anal. Calcd for C₄₁H₃₄O₆: C, 79.08; H, 5.50. Found: C, 78.94; H, 5.53.

4',5,7-Trihydroxy-6-(3-hydroxy-3-methylbutyl)isoflavone (Wighteone Hydrate) (3) The isoflavone 28 (720 mg, 1.15 mmol) in a solution of MeOH (30 ml) and dioxane (30 ml) was hydrogenolyzed over Pd/C (5%) (80 mg) at room temperature until uptake of hydrogen ceased. The resulting compound was recrystallized from a mixture of water and MeOH to give 3 (367 mg, 92%) as colorless needles, mp 230–232 °C (lit.,⁷⁾ 225–228 °C); ¹H-NMR (see Table 1). *Anal.* Calcd for $C_{20}H_{20}O_6$: C, 67.41; H, 5.66. Found: C, 67.22; H, 5.48.

4',7-Diacetoxy-5-hydroxy-6-(3-hydroxy-3-methylbutyl)coumarono-

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chromone (29) To a dioxane solution (12 ml) of the 2'-hydroxyisoflavone 2 (350 mg; 0.93 mmol), o-chloranil (303 mg, 1.2 mmol) was added and stirred at 80 °C for 10 min, and subsequently o-chloranil (305 mg, 1.2 mmol) was again added to the mixture, and the whole was stirred at 80 °C for 2 h. After removal of the solvent, unreacted o-chloranil was removed by silica gel column chromatography (AcOEt: CHCl₃=2:1 as a solvent) and the obtained coumaronochromone 4 was converted into the diacetate 29 by treatment with acetic anhydride (20 ml)-pyridine (1.5 ml) at 0 °C for 30 min. The resulting compound was purified by silica gel column chromatography (AcOEt: CHCl₃=2:1 as a solvent) to give 29 (205 mg, 61% via two steps from 2) as colorless needles, mp 271–274 °C; ¹H-NMR (CDCl₃) δ =1.31 (6H, s, CH₃×2), 1.55 (1H, s, OH), 1.71 and 2.73 (each 2H, m, CH₂), 2.36 and 2.39 (each 3H, s, COCH₃), 6.87 (1H, s, C₈-H), 7.20 (1H, dd, J=1.9 and 8.3 Hz, $C_{5'}$ -H), 7.39 (1H, d, J=1.9 Hz, $C_{3'}$ -H), 8.08 (1H, d, J=8.3 Hz, $C_{6'}$ -H), 13.16 (1H, s, C5-OH). Anal. Calcd for C24H22O9: C, 63.43; H, 4.88. Found: C. 63.53: H. 4.85.

4',5,7-Trihydroxy-6-(3-hydroxy-3-methylbutyl)coumaronochromone (4) Compound 29 (120 mg, 0.26 mmol) in a mixture of MeOH–dioxane was hydrolyzed with 10% aqueous NaOH at room temperature for 1 h. The resulting compound was recrystallized from a mixture of H₂O and MeOH to give 4 (66 mg, 67%) as colorless needles, mp 260–262 °C; ¹H-NMR (DMSO- d_6) δ =1.16 (6H, s, CH₃×2), 1.52 and 2.61 (each 2H, m, CH₂), 4.18 (1H, br s, OH), 6.61 (1H, s, C₈-H), 6.92 (1H, dd, J=2.0 and 8.9 Hz, C₅--H), 7.11 (1H, d, J=2.0 Hz, C₃--H), 7.74 (1H, d, J=8.9 Hz, C₆--H), 9.94 and 10.80 (each 1H, br s, OH), 13.13 (1H, s, C₅-OH). *Anal.* Calcd for C₂₀H₁₈O₇: C, 64.86; H, 4.90. Found: C, 64.69; H, 4.82.

References and Notes

- Dewick P. M., "The Flavonoids: Advances in Research Since 1980," ed. by Harborne J. B., Chapman and Hall, London, 1988.
- Ingham J. L., Tahara S., Harborne J. B., Z. Naturforsch. 38c, 194–200 (1983).
- 3) Woodward M. D., Phytochemistry, 18, 363-365 (1979).
- Harborne J. B., Ingham J. L., King L., Payne M., *Phytochemistry*, 15, 1485–1487 (1976).
- Fukui H., Egawa H., Koshimizu K., Mitsui T., Agric. Biol. Chem., 37, 417–421 (1973).
- Tahara S., Ingham J. L., Nakahara S., Mizutani J., Harborne J. B., *Phytochemistry*, 23, 1889–1900 (1984).
- Hashidoko Y., Tahara S., Mizutani J., Agric. Biol. Chem., 50, 1797– 1807 (1986).
- Tahara S., Nakahara S., Mizutani J., Ingham J. L., Agric. Biol. Chem., 48, 1471–1477 (1984).
- Ingham J. L., Keen N. T., Hymowitz T., *Phytochemistry*, 16, 1943– 1946 (1977).
- Tahara S., Nakahara S., Ingham J. L., Mizutani J., Nippon Nôgeikagaku Kaishi, 59, 1039–1044 (1985).
- 11) Jain A. C., Kumar A., Gupta R. C., J. Chem. Soc., Perkin Trans. 1, 1979, 279–287 (1979).
- Tsukayama M., Kawamura Y., Tamaki H., Kubo T., Horie T., Bull. Chem. Soc. Jpn., 62, 826–832 (1989).
- 13) Varady J., Tetrahedron Lett., 48, 4273-4275 (1965).
- 14) Tsukayama M., Li H., Tsurumoto K., Nishiuchi M., Kawamura Y., Bull. Chem. Soc. Jpn., 71, 2673—2680 (1998).
- Sonogashira K., Tohda Y., Hagihara N., *Tetrahedron Lett.*, 50, 4467–4470 (1975).
- 16) Tsukayama M., Wada H., Kishida M., Nishiuchi M., Kawamura Y., *Chem. Lett.*, **2000**, 1362—1363 (2000).
- 17) Tahara S., Ingham J. L., Mizutani J., Agric. Biol. Chem., 49, 1775– 1783 (1985).
- Tsukayama M., Oda A., Kawamura Y., Nishiuchi M., Yamashita K., *Tetrahedron Lett.*, 42, 6163—6166 (2001).
- 19) Ahluwalia V. K., Prakash C., Jolly R. S., J. Chem. Soc., Perkin Trans. 1, 1981, 1697—1702 (1981).
- 20) Farkas L., Gottsegen À., Nórgrádi M., Antus S., J. Chem. Soc., Perkin Trans. 1, 1974, 305—312 (1974).
- Horie T., Shibata K., Yamashita K., Fujii K., Tsukayama M., Ohtsuru Y., *Chem. Pharm. Bull.*, 46, 222–230 (1998).
- Becker H.-D., "The Chemistry of the Quinoid Compounds, Part 1," ed. by Patai S., John Wiley & Sons, London, 1974, pp. 335–423.
- Becker H.-D., Björk A., Adler E., J. Org. Chem., 45, 1596–1600 (1980).