

Constituents of Holothuroidea, 17.¹⁾ Isolation and Structure of Biologically Active Monosialo-Gangliosides from the Sea Cucumber *Cucumaria echinata*

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Three new monosialo-gangliosides, CEG-3 (3), CEG-4 (4), and CEG-5 (5), were obtained, together with two known gangliosides, SJG-1 (1) and CG-1 (2), from the lipid fraction of the chloroform/methanol extract of the sea cucumber *Cucumaria echinata*. The structures of the new gangliosides were determined on the basis of chemical and spectroscopic evidence to be 1-*O*-[4-*O*-acetyl- α -L-fucopyranosyl-(1 \rightarrow 11)]-(*N*-glycolyl- α -D-neuraminosyl)-(2 \rightarrow 6)- β -D-glucopyranosyl]-ceramide (3) and 1-*O*-[α -L-fucopyranosyl-(1 \rightarrow 11)]-(*N*-glycolyl- α -D-neuraminosyl)-(2 \rightarrow 6)- β -D-glucopyranosyl]-ceramide (4, 5). The ceramide moieties of each compound were composed of heterogeneous sphingosine or phytosphingosine bases, and 2-hydroxy or nonhydroxylated fatty acid units. These gangliosides showed neuritogenic activity toward the rat pheochromocytoma cell line PC-12 in the presence of nerve growth factor.

Key words glycosphingolipid; ganglioside; sea cucumber; *Cucumaria echinata*; neuritogenic activity

In our continuing research on biologically active glycosphingolipids (GSLs) from echinoderms, a series of studies on the isolation and structural elucidation of the GSLs from sea cucumber species have been performed in our laboratory.^{2–13)} In the study of the GSLs of the sea cucumber *Cucumaria echinata* (*gumi* in Japanese), we reported the isolation and structure of eight glucocerebrosides and a ganglioside.^{2,4)} In a continuation of the preceding studies,⁴⁾ the further isolation and characterization of the biologically active gangliosides from the sea cucumber *C. echinata* were carried out to develop novel medicinal resources from natural marine products. In this paper, we report the isolation and characterization of three new gangliosides from the whole bodies of *C. echinata*. The biological activities of the gangliosides are also reported.

The polar lipid fraction, which was obtained from the chloroform/methanol extract of the whole bodies of *C. echinata*, was subjected to repeated silica gel column chromatography to yield five compounds (1–5), each showing a single spot on TLC. Compounds 1 and 2 were identified as the known gangliosides SJG-1⁶⁾ and CG-1,⁴⁾ respectively, previously obtained from the sea cucumber *Stichopus japonicus* and *C. echinata* on the basis of chemical and spectroscopic evidence (Figs. 1, 2, Table 1, and Experimental).

In its ¹³C-NMR spectrum (Fig. 3, Table 1), compound 4 exhibits the characteristic signals of a sphingosine-type ceramide, with a nonhydroxylated fatty acid and a sugar moiety at C-1 [δ : 70.2 (C-1), 53.9 (C-2), 72.2 (C-3), 131.2 (C-4), 132.8 (C-5), and 175.7 (C-1')]. The ¹³C-NMR spectrum of 4 also features signals due to three anomeric carbons at δ 105.2, 101.3, and 100.6, one of which (δ 101.3) is a quaternary carbon signal, indicating the presence of a sialic acid residue. Therefore 4 is suggested to be a sphingosine-type ganglioside, with nonhydroxylated fatty acid groups and three monosaccharide units. Furthermore, 4 is presumed to have normal-type fatty acids and *ante*-iso-type long-chain bases (LCBs), since the carbon signals for the terminal methyl groups are observed at δ 14.1 (normal form) and δ 11.5 and 19.2 (*ante*-iso form) in the ¹³C-NMR spectrum (Fig.

3, Table 1).

The structure of the ceramide moiety was examined first. When 4 was methanolized with methanolic hydrochloric acid, a mixture of fatty acid methyl esters (FAMs) and an LCB was obtained, together with methyl glucoside and fucoside. The FAM mixture was analyzed using GC-MS, which revealed the presence of 10 components (see Experimental), and the major one was characterized as methyl octadecanoate (stearate). The LCB was found to be 2-amino-1,3-dihydroxy-4-heptadecene based on GC-MS analysis of its TMS derivative.

The stereochemistry of the ceramide moiety of 4 is presumed to be (2*S*,3*R*,4*E*) the same as that of the precursor compound, glucocerebroside CE-1,⁴⁾ co-existing in the same organism.

The structure of the trisaccharide moiety of 4 was established as follows. The presence of glucose (Glc) and fucose (Fuc) was obvious from the results of the methanolysis of 4 (*vide supra*). A detailed analysis of the ¹³C-NMR spectrum of 4 revealed the characteristic signals [δ 173.1 (C-1), 101.3 (C-2), 40.3 (C-3), 54.5 (C-5), 63.3 (C-9), 175.7 (C-10), 67.5 (C-11)] of an *N*-glycolylneuraminic acid (NeuGc) residue (Table 1). In the negative FAB-MS of 4, the molecular ion and fragment ion peaks arising from cleavage of the glycosidic linkages of the major component are observed at *m/z* 1165, 1019, 712, and 550, indicating the presence of a trisaccharide moiety, deoxyhexose \rightarrow NeuGc \rightarrow hexose, as shown in Fig. 2.

Methylation of 4, following the Ciucanu–Kerek method,¹⁴⁾ afforded the permethylated product 4-Me. Partially methylated alditol acetates (S-1, S-2) prepared from 4-Me were analyzed by GC-MS and identified as the alditols derived from 6-linked hexopyranose and terminal 6-deoxy-hexopyranose, respectively. On the other hand, 4-Me was methanolized, the methanolysate was acetylated, and the acetate of partially methylated NeuGc (S-3) derived from 11-linked NeuGc was detected by means of GC-MS analysis. On the basis of the above evidence, the trisaccharide moiety of 4 must be Fuc-(1 \rightarrow 11)-NeuGc-(2 \rightarrow 6)-Glc. The configurations of Fuc,

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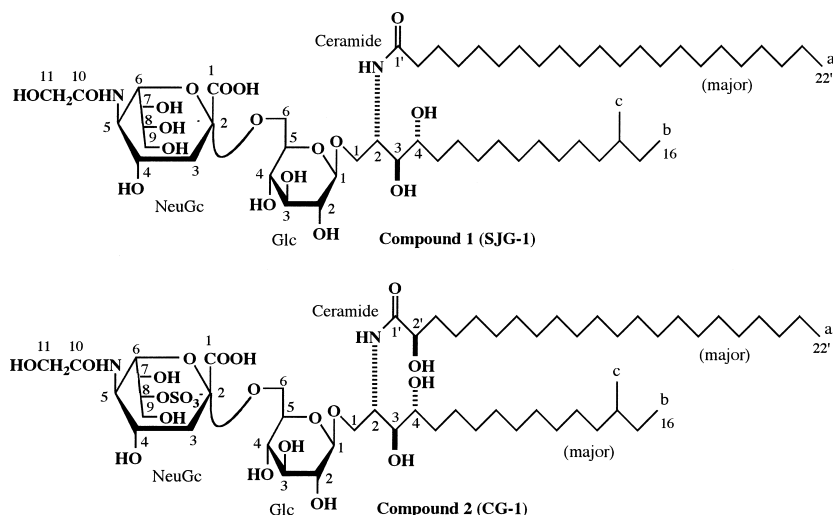


Fig. 1. Structure of Compounds 1 and 2

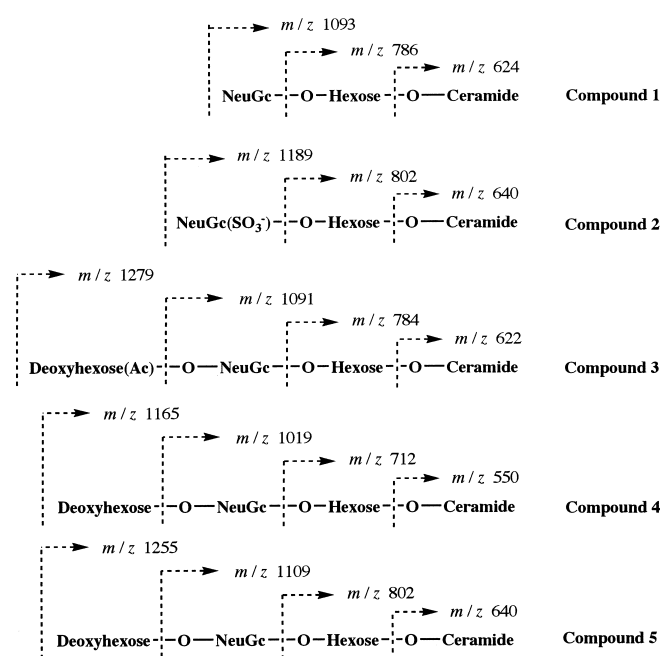


Fig. 2. Negative FAB Mass Fragmentation of the Major Component of Compounds 1–5

NeuGc, and Glc are believed to be α , α , and β on the basis of their anomeric carbon signals (δ 100.6, 101.3,¹⁵ 105.2) in the ¹³C-NMR spectrum of 4. In addition, the absolute configurations of the glucose and fucose units were verified as being the D- and L-form using the Hara method.¹⁶⁾

Consequently, if NeuGc is assumed to belong to the most commonly found D-series, then compound 4 is the α -L-fucopyranosyl-(1 \rightarrow 11)-(N-glycolyl- α -D-neuraminosyl)-(2 \rightarrow 6)- β -D-glucopyranoside of a ceramide, composed of (2S,3R,4E)-C₁₇-sphingosine and heterogeneous nonhydroxylated fatty acids (stearic acid as the major component), as shown in Fig. 3.

Compound 5 exhibits characteristic signals due to the phytosphingosine-type ceramide, with a 2-hydroxy fatty acid and a sugar moiety at C-1 [δ 70.6 (C-1), 51.3 (C-2), 75.9 (C-3), 72.5 (C-4), 176.5 (C-1') and 72.4 (C-2')] in its ¹³C-NMR

spectrum (Fig. 3, Table 1). The ¹³C-NMR spectrum of 5 also shows three anomeric carbon signals at δ 104.9, 101.1, and 100.4, one of which (δ 101.1) is a quaternary carbon signal derived from one sialic acid moiety (Table 1). Accordingly, 5 is suggested to be a phytosphingosine-type ganglioside, with 2-hydroxy fatty acid groups and three monosaccharide units. The terminal methyl groups of the ceramide moiety of 5 must be the same as that of 4 based on their carbon atom signals (Table 1).

Methanolysis of 5 afforded a mixture of FAM, LCB, and methyl glycosides identical to Glc and Fuc. The FAM mixture was analyzed using GC-MS and seven components were detected (see Experimental). The major component was methyl 2-hydroxydocosanoate. On the other hand, the LCB component was found to be 2-amino-1,3,4-trihydroxy-heptadecane by means of GC-MS analysis of its TMS derivative. Furthermore, the stereochemistry of the ceramide moiety of 5 is presumed to be (2S,3S,4R,2'R) because the ceramide part of glucocerebroside CE-3⁴⁾ coexisting in the same organism has the same absolute configuration.

The structure of the oligosaccharide moiety of 5 was indicated to be the same trisaccharide as that of 4 by comparison of the signals due to sugar moieties in their ¹³C-NMR spectra (Table 1) and the fragmentations in their negative FAB-MS (Fig. 2). Furthermore, chemical degradation of 5-Me, permethylated 5, providing S-1, S-2, and S-3, verified the above suggestion (Fig. 3). Since the absolute configurations (D- and L-form) of the Glc and Fuc units were confirmed, if NeuGc is assumed to belong to the D-series, then 5 is the α -L-fucopyranosyl-(1 \rightarrow 11)-(N-glycolyl- α -D-neuraminosyl)-(2 \rightarrow 6)- β -D-glucopyranoside of a ceramide, composed of (2S,3S,4R)-C₁₇-phytosphingosine and heterogeneous (2R)-2-hydroxy fatty acids (docosanoic acid as the major component), as shown in Fig. 3.

In its ¹³C-NMR spectrum (Fig. 4, Table 1), compound 3 exhibits the characteristic signals of a sphingosine-type ceramide, with a 2-hydroxy fatty acid and a sugar moiety at C-1 [δ 70.2 (C-1), 53.8 (C-2), 72.2 (C-3), 131.2 (C-4), 133.2 (C-5), 175.8 (C-1'), and 72.4 (C-2')]. The ¹³C-NMR spectrum of 3 also features signals due to three anomeric carbons at δ 104.8, 101.0, and 100.4, one of which (δ 101.0) is a

Table 1. ^{13}C -NMR Spectral Data (δ Values) of Compounds 1–5 in $\text{C}_5\text{D}_5\text{N}-\text{D}_2\text{O}$ [95 : 5 (1–3), 9 : 1 (4, 5)]

C		1	2	3	4	5
Ceramide						
1	(t)	70.6	70.5	70.2	70.2	70.6
2	(d)	53.7	51.1	53.8	53.9	51.3
3	(d)	76.0	75.7	72.2 ^{e)}	72.2	75.9
4	(d)	72.5	72.2 ^{d)}	131.2	131.2	72.5 ^{e)}
5	(d)			133.2	132.8	
1'	(s)	176.5	175.5	175.8	175.7	176.5
2'	(d)		72.4 ^{d)}	72.4 ^{e)}		72.4 ^{e)}
$\text{CH}_3^{\text{a)}$	(q)	14.1	14.1	14.1	14.1	14.1
$\text{CH}_3^{\text{b)}$	(q)	11.5	11.4	11.4	11.5	11.5
$\text{CH}_3^{\text{c)}$	(q)	19.2	19.1	19.2	19.2	19.2
Glc						
1	(d)	104.8	104.7	104.8	105.2	104.9
2	(d)	74.4	74.7	74.9	74.2	74.4
3	(d)	76.8	77.3	77.2	77.2	77.0
4	(d)	70.0	69.9	70.0	71.1	70.8
5	(d)	76.0	77.0	76.5	76.0	76.0
6	(t)	70.8	68.2	67.6	67.5	68.2
NeuGc						
1	(s)	174.5	173.8	172.3 ^{b)}	173.1	173.8
2	(s)	101.0	101.3	101.0	101.3	101.1
3	(t)	42.6	41.6	40.1	40.3	40.3
4	(d)	66.0	68.5	69.3	67.6	68.6
5	(d)	53.7	52.2	53.9	54.5	54.3
6	(d)	76.0	72.0	74.5	74.6	74.6
7	(d)	68.6	70.8	69.8	69.6	69.8
8	(d)	77.4	80.4	77.2	77.5	77.2
9	(t)	63.9	61.9	63.1	63.3	63.3
10	(s)	176.5	175.8	175.8	175.7	175.7
11	(t)	62.2	62.3	67.6	67.5	67.6
Fuc						
1	(d)			100.4	100.6	100.4
2	(d)			69.8	69.6	70.0
3	(d)			67.2	71.0	71.5
4	(d)			78.8	72.4	72.6
5	(d)			67.0	68.6	68.7
6	(q)			16.8	16.8	16.8
COCH_3	(s)			172.6 ^{b)}		
COCH_3	(q)			22.8		

a–c) Terminal methyl groups in the normal and *ante*-iso type of side chain (see Figs. 1, 3, 4). *d–g*) Assignments may be interchanged in each vertical column.

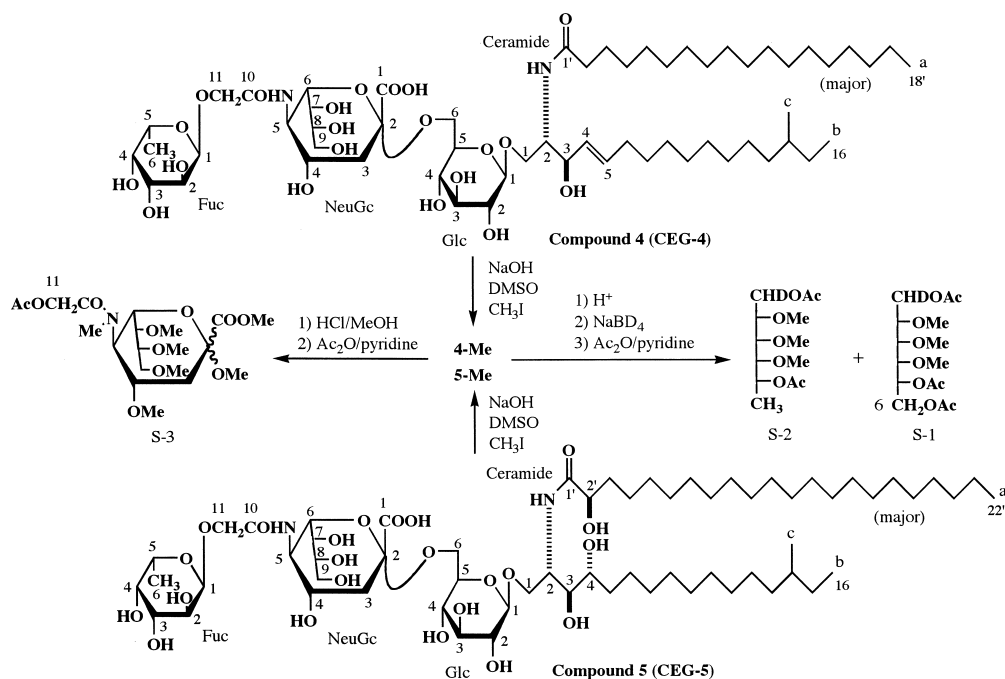


Fig. 3. Structure of Compounds 4 and 5

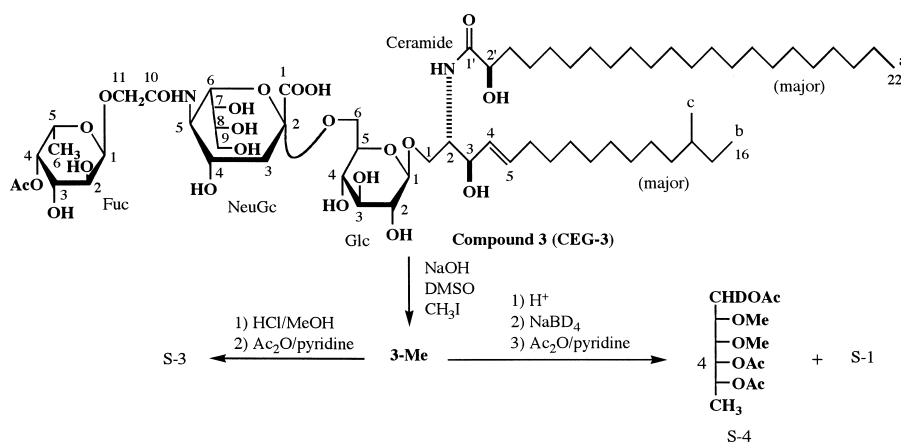


Fig. 4. Structure of Compound 3

quaternary carbon signal, indicating the presence of a sialic acid residue, and an acetyl group at δ 172.6 and 22.8. Therefore **3** is suggested to be a sphingosine-type ganglioside, with 2-hydroxy fatty acid groups, three monosaccharide units, and an acetyl group. Furthermore, the terminal methyl groups of the ceramide moiety of **3** are also same as those of **4** from their carbon atom signals (Table 1).

When compound **3** was methanolized, a mixture of FAM, LCB, and methyl glycosides, Glc and Fuc, was obtained. The major components of the FAM and LCB mixture were identified as methyl 2-hydroxydocosanoate and 2-amino-1,3-dihydroxy-4-heptadecene, respectively, by means of GC-MS analysis. The stereochemistry of the ceramide moiety of **3** is presumed to be (2*S*,3*R*,4*E*,2'*R*) based on the existence of glucocerebroside CE-2² with the same absolute configurations in the same organism.

The presence of the trisaccharide moiety deoxyhexose (Ac) \rightarrow NeuGc \rightarrow hexose, was suggested on the basis of the molecular ion and fragment ion peaks at m/z 1279, 1091, 784, and 622 arising from cleavage of the glycosidic linkages of the major component in the negative FAB-MS of **3**, as shown in Fig. 2. When the signals due to the sugar moieties of **3** and **4** in their ¹³C-NMR spectra were compared, they were nearly identical except for the signals ascribable to C-4 of Fuc (Table 1). The downfield signal for C-4 (δ 78.8) of the Fuc unit resulting from esterification^{17,18} in the ¹³C-NMR spectrum of **3** indicates the presence of the acetyl group at C-4 of the Fuc unit. Therefore the trisaccharide moiety of **3** must be 4-*O*-Ac-Fuc-(1 \rightarrow 11)-NeuGc-(2 \rightarrow 6)-Glc. Furthermore, chemical degradation of permethylated **3**, **3-Me**, yielding S-1, S-3, and the alditol derived from the 4-linked 6-deoxyhexopyranose (S-4), confirmed the structure of the trisaccharide (Fig. 4).

On the basis of the above facts, the anomeric carbon signals, and the absolute configurations (D-, D- and L-form) of Glc, NeuGc (assuming) and Fuc units, compound **3** is regarded as 4-*O*-acetyl- α -L-fucopyranosyl-(1 \rightarrow 11)-(*N*-glycolyl- α -D-neuraminosyl)-(2 \rightarrow 6)- β -D-glucopyranoside of a ceramide, composed of (2*S*,3*R*,4*E*)-C₁₇-sphingosine and (2*R*)-2-hydroxydocosanoic acid as the major components, as shown in Fig. 4.

The effects of the isolated gangliosides (**1**–**5**) on the neuritogenesis of a rat pheochromocytoma cell line (PC-12 cells) were investigated. The results show that they displayed neuro-

togenic activity in the presence of nerve growth factor (NGF). The proportion of cells with neurites longer than the diameter of the cell body of compounds **1**–**5** at a concentration of 10 μ M was 39.1%, 43.0%, 50.8%, 34.0%, and 35.7% when compared with the control (NGF 5 ng/ml: 7.5%). Furthermore, the effects of **1**, **2**, and **3** were greater than that of the mammalian ganglioside GM₁ (35.6%). Interestingly, compound **3** with an acetyl group at the terminal Fuc unit showed the most potent activity.

Compound **3** is, to the best of our knowledge, the first ganglioside with an *O*-acetyl group from echinoderms. Although a ganglioside with the same sugar moiety as that of **4** and **5** has been obtained from the sea cucumber *Stichopus chloronotus*,¹¹ they differ in the basic structure of their ceramide moieties. Three new monosialo-gangliosides (**3**, **4**, **5**) obtained in this study were designated CEG-3, -4, and -5, respectively. The isolation and characterization of such neuritogenically active gangliosides are attracting considerable attention with regard to the development of new medicines from natural marine products.

Experimental

IR spectra were obtained on a Jasco FT/IR-410 infrared spectrophotometer. ¹H- and ¹³C-NMR spectra were recorded on a Jeol GX-270 spectrometer (270, 67.8 MHz) or a Varian Unity-500 spectrometer (500, 125 MHz). Negative-ion FAB-MS spectra were acquired with a Jeol JMS-SX-102 mass spectrometer (xenon atom beam; matrix, triethanolamine). GC and GC-MS were recorded with a Shimadzu QP-5050A [EI mode; ionizing potential, 120 eV; column, TC-1701 (0.25 mm \times 30 m, GL Science Inc.); carrier gas, He].

Separation of Compounds 1–5 The whole bodies of the sea cucumber *C. echinata* (126 kg), which was collected from the Sea of Genkai, Japan, in 1997, were chopped and extracted with CHCl₃-MeOH [1 : 4 (54), 1 : 2 (54) \times 2]. The combined extracts were concentrated *in vacuo* to give an aqueous solution (120 l), which was extracted three times with *n*-hexane (80 l). The *n*-hexane phase was concentrated *in vacuo* to give a residue (3.7 kg). The residue was dissolved in acetone. The acetone-insoluble part (2.0 kg), which was the polar lipid fraction, was chromatographed on silica gel (solvent CHCl₃-MeOH-H₂O, 95 : 15 : 0 to 4 : 6 : 2) to give nine fractions. Successive column chromatography of fraction 7 (silica gel, solvent CHCl₃-MeOH-H₂O, 7 : 3 : 0.5 to 5 : 5 : 1) afforded compound **1** (4.5 mg) (R_f =0.45) and compound **2** (112 mg) (R_f =0.33). On the other hand, fraction 8 was further chromatographed on silica gel (solvent CHCl₃-MeOH-H₂O, 7 : 3 : 0.5 to 5 : 5 : 1) to afford seven fractions. Fraction 6 of the seven fractions was chromatographed successively on silica gel (solvent CHCl₃-MeOH-AcOEt-H₂O, 3 : 4 : 3 : 0.6 to CHCl₃-MeOH-H₂O, 5 : 5 : 1) to give compound **3** (14.0 mg) (R_f =0.25), compound **4** (5.6 mg) (R_f =0.22), and compound **5** (1.7 mg) (R_f =0.20) [silica gel TLC, solvent CHCl₃-MeOH-2% CaCl₂ aq. (6 : 4 : 0.8)].

Compound **1** (SJG-1): Amorphous powder. IR (KBr) cm⁻¹: 3375 (OH),

1646, 1548 (amide). Negative-ion FAB-MS: see Fig. 2. $^1\text{H-NMR}$ ($\text{C}_5\text{D}_5\text{N}:\text{D}_2\text{O}$, 95:5) δ : 0.85 (9H, m, terminal methyl groups). $^{13}\text{C-NMR}$: see Table 1.

Compound 2 (CG-1): Amorphous powder. IR (KBr) cm^{-1} : 3388 (OH), 1643, 1541 (amide). Negative-ion FAB-MS: see Fig. 2. $^1\text{H-NMR}$ ($\text{C}_5\text{D}_5\text{N}:\text{D}_2\text{O}$, 95:5) δ : 0.86 (9H, m, terminal methyl groups). $^{13}\text{C-NMR}$: see Table 1.

Compound 3 (CEG-3): Amorphous powder. IR (KBr) cm^{-1} : 3419 (OH), 1644, 1550 (amide), 1710 (ester group). Negative-ion FAB-MS: see Fig. 2. $^1\text{H-NMR}$ ($\text{C}_5\text{D}_5\text{N}:\text{D}_2\text{O}$, 95:5) δ : 2.00 (3H, s, COCH_3), 1.42 (3H, d, $J=4.0\text{ Hz}$, CH_3 of Fuc), 0.87 (9H, m, terminal methyl groups). $^{13}\text{C-NMR}$: see Table 1.

Compound 4 (CEG-4): Amorphous powder. IR (KBr) cm^{-1} : 3378 (OH), 1645, 1554 (amide). Negative-ion FAB-MS: see Fig. 2. $^1\text{H-NMR}$ ($\text{C}_5\text{D}_5\text{N}:\text{D}_2\text{O}$, 9:1) δ : 1.41 (3H, d, $J=4.0\text{ Hz}$, CH_3 of Fuc), 0.85 (9H, m, terminal methyl groups). $^{13}\text{C-NMR}$: see Table 1.

Compound 5 (CEG-5): Amorphous powder. IR (KBr) cm^{-1} : 3419 (OH), 1650, 1549 (amide). Negative-ion FAB-MS: see Fig. 2. $^1\text{H-NMR}$ ($\text{C}_5\text{D}_5\text{N}:\text{D}_2\text{O}$, 9:1) δ : 1.42 (3H, d, $J=4.0\text{ Hz}$, CH_3 of Fuc), 0.87 (9H, m, terminal methyl groups). $^{13}\text{C-NMR}$: see Table 1.

Methanolysis of 1 Compound **1** (0.5 mg) was heated with 10% HCl in MeOH (0.5 ml) at 70 °C for 18 h. The reaction mixture was then extracted with *n*-hexane, and the extract was concentrated *in vacuo* to yield a mixture of FAM. The MeOH layer was neutralized with Ag_2CO_3 , filtered, and the filtrate was concentrated *in vacuo* to give a mixture of LCB and methyl glycoside.

GC-MS Analysis of FAM from 1 A FAM mixture from **1** was subjected to GC-MS [column temperature: 150–300 °C (rate of temperature increase 5 °C/min)]. The results were as follows: methyl octadecanoate, t_R [min] (ratio of peak areas)=11.8 (18), m/z : 298 (M^+), 255 ($\text{M}-43$) $^+$; methyl nonadecanoate, $t_R=13.5$ (4), m/z : 312 (M^+), 269 ($\text{M}-43$) $^+$; methyl icosanoate, $t_R=15.0$ (6), m/z : 326 (M^+), 283 ($\text{M}-43$) $^+$; methyl heneicosanoate, $t_R=16.5$ (11), m/z : 340 (M^+), 297 ($\text{M}-43$) $^+$; methyl docosanoate, $t_R=18.2$ (9), m/z : 352 (M^+), 320 ($\text{M}-32$) $^+$; methyl docosanoate, $t_R=18.5$ (39), m/z : 354 (M^+), 311 ($\text{M}-43$) $^+$; methyl tricosanoate, $t_R=20.8$ (13), m/z : 368 (M^+), 325 ($\text{M}-43$) $^+$.

GC-MS Analysis of TMS Ethers of LCB from 1 The mixture of LCB and methyl glycoside from **1** was heated with 1-(trimethylsilyl)-imidazole–pyridine (1:1) for 15 min at 70 °C and the reaction mixture (TMS ethers) was analyzed using GC-MS [column temperature: 150–300 °C (rate of temperature increase 5 °C/min)]. The results were as follows: 2-amino-1,3,4-trihydroxy-heptadecane, t_R [min]=16.4, m/z : 326 ($\text{M}-193$) $^+$, 285 ($\text{M}-234$) $^+$, 132.

Methanolysis of 2 In the same manner as described for **1**, compound **2** was methanolized and the reaction mixture was worked up to give a mixture of FAM and a residue composed of LCB and methyl glycoside.

GC-MS Analysis of FAM from 2 A FAM mixture from **2** was subjected to GC-MS under the same conditions as for **1**. The results were as follows: methyl 2-hydroxyoctadecanoate, t_R [min] (ratio of peak areas)=14.5 (9), m/z : 314 (M^+), 255 ($\text{M}-59$) $^+$; methyl 2-hydroxyicosanoate, $t_R=17.9$ (7), m/z : 342 (M^+), 283 ($\text{M}-59$) $^+$; methyl 2-hydroxydocosanoate, $t_R=22.3$ (26), m/z : 370 (M^+), 311 ($\text{M}-59$) $^+$; methyl 2-hydroxytricosanoate, $t_R=25.7$ (17), m/z : 384 (M^+), 325 ($\text{M}-59$) $^+$; methyl 2-hydroxytetracosanoate, $t_R=29.4$ (12), m/z : 396 (M^+), 337 ($\text{M}-59$) $^+$; methyl 2-hydroxytetracosanoate, $t_R=29.9$ (23), m/z : 398 (M^+), 339 ($\text{M}-59$) $^+$; methyl octadecanoate, $t_R=11.5$ (4); methyl docosanoate, $t_R=18.4$ (2).

GC-MS Analysis of TMS Ethers of LCB from 2 The mixture of LCB and methyl glycoside from **2** was treated in the same way as **1** and the reaction mixture (TMS ethers) was analyzed using GC-MS under the same conditions as for **1**. The results were as follows: 2-amino-1,3,4-trihydroxy-heptadecane, t_R [min] (ratio of peak areas)=16.0 (75); 2-amino-1,3,4-trihydroxy-nonadecane, $t_R=19.5$ (25), m/z : 354 ($\text{M}-193$) $^+$, 313 ($\text{M}-234$) $^+$, 132.

Methanolysis of 4 Compound **4** was methanolized and the reaction mixture was worked up in the same manner as described for **1** to yield a mixture of FAM and a residue composed of LCB and methyl glycosides.

FAM: $^1\text{H-NMR}$ (CDCl_3) δ : 3.67 (3H, s, COOCH_3), 0.87 (3H, t, $J=6.7\text{ Hz}$, terminal CH_3).

GC-MS Analysis of FAM from 4 A FAM mixture from **4** was analyzed using GC-MS under the same conditions as for **1**, and the results were as follows: methyl hexadecanoate, t_R [min] (ratio of peak areas)=8.2 (14), m/z : 270 (M^+), 227 ($\text{M}-43$) $^+$; methyl heptadecanoate, $t_R=9.7$ (3), m/z : 284 (M^+), 241 ($\text{M}-43$) $^+$; methyl octadecanoate, $t_R=11.4$ (26); methyl icosanoate, $t_R=14.7$ (10); methyl heneicosanoate, $t_R=16.1$ (3); methyl docosanoate, $t_R=18.2$ (15); methyl tricosanoate, $t_R=20.1$ (3), m/z : 366 (M^+),

334 ($\text{M}-32$) $^+$; methyl tricosanoate, $t_R=20.5$ (9); methyl tetracosanoate, $t_R=22.9$ (5), m/z : 380 (M^+), 348 ($\text{M}-32$) $^+$; methyl tetracosanoate, $t_R=23.3$ (12), m/z : 382 (M^+), 339 ($\text{M}-43$) $^+$.

GC-MS Analysis of TMS Ethers of LCB from 4 The TMS ethers of the mixture of LCB and methyl glycosides from **4** prepared in the same way as **1** were analyzed using GC-MS under the same conditions as for **1**. The results were as follows: 2-amino-1,3-dihydroxy-4-heptadecene, t_R [min]=13.4, m/z : 326 ($\text{M}-103$) $^+$, 297 ($\text{M}-132$) $^+$, 132.

GC Analysis of TMS Ethers of Methyl Glycosides from 4 The mixture of TMS ethers of LCB and methyl glycosides from **4** was analyzed using GC-MS [column temperature: 100–300 °C (rate of temperature increase 5 °C/min)]: t_R [min]=15.2, 16.1, and 17.0 (methyl Fuc); 17.9 and 18.0 (methyl Glc).

Determination of Absolute Configuration of Fuc and Glc Moieties of 4 (Hara Method) Compound **4** (0.5 mg) was heated with 4 N H_2SO_4 (0.5 ml) at 100 °C for 24 h. The reaction mixture was then extracted with *n*-hexane, and the acidic aqueous phase was neutralized with $\text{Ba}(\text{OH})_2$, centrifuged, and the clear supernatant solution was concentrated. The residue (sugar fraction) was heated with L-cysteine methyl ester hydrochloride (1.0 mg) and pyridine (0.05 ml) at 70 °C for 1 h. Then, 0.05 ml of 1-(trimethylsilyl)imidazole was added and the mixture was heated at 70 °C for a further 15 min to yield trimethylsilyl ether of the methyl (4*R*)-thiazolidine-4-carboxylate derivatives. The derivatives were analyzed using GC-MS [column temperature: 180–300 °C (rate of temperature increase 2.5 °C/min)]; t_R [min]=19.7 and 25.1 (derivative of D-Fuc, 21.8; L-Fuc, 19.7; D-Glc, 25.1; L-Glc, 25.9).

Methylation of 4 (Ciucanu–Kerek Method) NaOH–dimethylsulfoxide (DMSO) solution, which was prepared from powdered NaOH (80 mg) and DMSO (2 ml), and MeI (0.2 ml) were added to **4** (1 mg), and the mixture was stirred for 30 min. The reaction mixture was then diluted with water (20 ml), extracted with Et_2O (10 ml \times 3), the Et_2O phases were washed with water, and the solvent was evaporated *in vacuo* to give permethylated **4**, denoted **4-Me**.

Preparation and GC-MS Analysis of Partially Methylated Alditol Acetates from 4-Me Compound **4-Me** (0.5 mg) was heated with 90% HCOOH –10% CF_3COOH (1:1) (0.5 ml) at 100 °C for 18 h in a small-volume sealed vial, and then the solvents were evaporated *in vacuo*. The residue was alkalinized with 7% NH_3 aq. and NaBD_4 (10 mg) was added. After allowing the mixture to stand at room temperature for 7 h, it was acidified with AcOH and concentrated *in vacuo*. H_3BO_3 present in the residue was removed by distillation with MeOH (three times). The residue was heated with Ac_2O – $\text{C}_5\text{H}_5\text{N}$ (1:1, 0.5 ml) at 70 °C for 2 h. The reaction mixture was evaporated *in vacuo* with toluene to give partially methylated alditol acetates. The acetates were subjected to GC-MS [column temperature 150–300 °C (rate of temperature increase 5 °C/min)]. The results were as follows: S-1, t_R [min]=15.2, m/z : 118, 162, 189, 233 [1,5,6-tri-*O*-acetyl-2,3,4-tri-*O*-methylhexitol (derived from 6-linked hexopyranose)]; S-2, $t_R=9.1$, m/z : 118, 131, 162, 175 [1,5-di-*O*-acetyl-6-deoxy-2,3,4-tri-*O*-methylhexitol (derived from terminal 6-deoxyhexopyranose)].

Preparation and GC-MS Analysis of Acetate of Partially Methylated Sialic Acid from 4-Me Compound **4-Me** (0.5 mg) was heated with 10% HCl in MeOH (0.5 ml) at 70 °C for 18 h in a small-volume sealed vial. The reaction mixture was then neutralized with Ag_2CO_3 , filtered, and the filtrate was concentrated *in vacuo*. The residue (methanolysate) was heated with Ac_2O – $\text{C}_5\text{H}_5\text{N}$ (1:1, 0.5 ml) at 70 °C for 2 h. The resulting mixture was evaporated *in vacuo* with toluene and the residue was subjected to GC-MS [column temperature 200–300 °C (rate of temperature increase 5 °C/min)]: S-3, t_R [min]=30.9, m/z : 187, 201, 376, 406 [methyl *N*-glycolyl-11-*O*-acetyl-*N*-methyl-2,4,7,8,9-penta-*O*-methylneuraminic acid (derived from 11-linked NeuGc)].

Methanolysis of 5 In the same manner as described above, compound **5** was methanolized and the reaction mixture was worked up to give a mixture of FAM and a residue composed of LCB and methyl glycoside.

GC-MS Analysis of FAM from 5 A FAM mixture from **5** was subjected to GC-MS under the same conditions as described above. Methyl octadecanoate (ratio of peak areas, 7), methyl 2-hydroxyoctadecanoate (15), methyl 2-hydroxyicosanoate (14), methyl 2-hydroxydocosanoate (24), methyl 2-hydroxytricosanoate (12), methyl 2-hydroxytetracosanoate (10), and methyl 2-hydroxytetracosanoate (18) were detected.

FAM: $^1\text{H-NMR}$ (CDCl_3) δ : 3.67 (3H, s, COOCH_3), 0.88 (3H, t, $J=6.8\text{ Hz}$, terminal CH_3).

GC-MS Analyses of TMS Ethers of LCB and Methyl Glycosides from 5 The residue (mixture of LCB and methyl glycosides) from **5** was trimethylsilylated and the reaction mixture was analyzed using GC-MS in

the same manner as above. 2-Amino-1,3,4-trihydroxy-heptadecane (LCB) and methyl Fuc and Glc (methyl glycosides) were detected.

Determination of the Absolute Configuration of Fuc and Glc Moieties of 5 Compound **5** was subjected to acid hydrolysis and the sugar fraction was treated in the same manner as described for **4**, thereby affording the trimethylsilyl ether of the methyl (4*R*)-thiazolidine-4-carboxylate derivatives. The derivatives were analyzed using GC-MS under the same conditions as above, and L-Fuc and D-Glc were detected.

Preparation and GC-MS Analyses of Partially Methylated Alditol Acetates and Acetate of Partially Methylated Sialic Acid from 5-Me Compound **5** was methylated according to the Ciucanu-Kerek method and the reaction mixture was worked up in the same manner as described for **4**, thereby yielding permethylated **5**, denoted **5-Me**. Compound **5-Me** was hydrolyzed, reduced, and then acetylated, and the partially methylated alditol acetates were analyzed using GC-MS in the same manner as described for **4-Me**, whereupon S-1 and S-2, derived from 6-linked hexopyranose and terminal 6-deoxyhexopyranose, respectively, were detected. On the other hand, **5-Me** was methanolized and then acetylated as above, and the acetate was subjected to GC-MS under the same conditions as described above, and S-3 (derived from 11-linked NeuGc) was detected.

Analyses of FAM, LCB, and Methyl Glycosides from 3 Experiments were conducted as before, leading to a mixture of FAM and a residue composed of LCB and methyl glycosides derived from compound **3**. The FAM mixture was subjected to GC-MS under the same conditions as described above, and methyl hexadecanoate (ratio of peak areas, 1), methyl heptadecanoate (1), methyl octadecanoate (1), methyl octadecanoate (6), methyl nonadecanoate (2), methyl icosanoate (5), methyl heneicosanoate (4), methyl docosanoate (5), methyl docosanoate (9), methyl tricosanoate (5), methyl 2-hydroxyoctadecanoate (11), methyl 2-hydroxydocosanoate (21), methyl 2-hydroxytricosanoate (10), methyl 2-hydroxytetracosanoate (5), and methyl 2-hydroxytetracosanoate (13) were detected.

FAM: ¹H-NMR (CDCl₃) δ: 3.67 (3H, s, COOCH₃), 0.88 (3H, t, *J*=6.6 Hz, terminal CH₃).

The mixture of LCB and methyl glycosides was trimethylsilylated and analyzed using GC-MS as above, and 2-amino-1,3-dihydroxy-4-heptadecene (ratio of peak area, 69), 2-amino-1,3,4-trihydroxy-heptadecane (31), and methyl Fuc and Glc were detected.

Determination of the Absolute Configuration of Fuc and Glc Moieties of 3 Conducted in the same manner as above, L-Fuc and D-Glc were detected.

Preparation and GC-MS Analyses of Partially Methylated Alditol Acetates and Acetate of Partially Methylated Sialic Acid from 3-Me The partially methylated alditol acetates prepared from **3-Me**, permethylated **3**, were analyzed using GC-MS in the same manner as described above, and S-1 and S-4, *t*_R [min]=10.7, *m/z*: 118, 203 [1,4,5-tri-*O*-acetyl-6-deoxy-2,3-di-*O*-methylhexitol (derived from 4-linked 6-deoxyhexopyranose)] were detected. On the other hand, methanolysis followed by acetylation of **3-Me** under the same conditions as above afforded S-3.

Biological Assay The neurotogenic activity of compounds **1**, **2**, **3**, **4**, and **5** on PC-12 cells was observed according to a method previously reported.¹⁰⁾

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