Two New Homoisoflavonoids from *Caesalpinia pulcherrima*

Muchchintala MAHESWARA, Vidavalur SIDDAIAH, and Chunduri VENKATA RAO*

Natural Products Division, Department of Chemistry, Sri Venkateswara University; Tirupati-517 502, India. Received December 26, 2005; accepted April 4, 2006

Two new homoisoflavonoids, (E)-7-methoxy-3-(4'-methoxybenzylidene)chroman-4-one (1) and (E)-7-hydroxy-3-(3',4',5'-trimethoxybenzylidene)chroman-4-one (5), along with three known homoisoflavonoids (Z)-7-hydroxy-3-(4'-methoxybenzylidene)chroman-4-one (isobon ducellin) (2), (E)-7-hydroxy-3-(4'-methoxybenzylidene) chroman-4-one (bonducellin) (3) and (E)-7-hydroxy-3-(2',4'-dimethoxybenzylidene)chroman-4-one (4) were isolated from the whole plant of *Caesalpinia pulcherrima*. The structures of these new compounds were elucidated by electron impact mass spectrometry (EI-MS) and 1D and 2D-NMR spectral studies. Antimicrobial activity of the new compounds was evaluated.

Key words Caesalpinia pulcherrima; Leguminosae; homoisoflavonoid; antimicrobial activity

Caesalpinia pulcherrima (Leguminosae) is a small size plant distributed in the hill tracks of Tirumula Hills, Andhra Pradesh, South India.¹⁾ In traditional Indian Medicine C. pulcherrima is used in the treatment of tridosha, fever, ulcer, abortifacient, emmenagogue, asthma, tumors, vata and skin diseases.²⁾ Previous studies on this plant have resulted in the isolation of several diterpenoids,³⁻⁶ flavanoids,⁷⁾ peltogy-noids⁸⁾ and homoisoflavonoids.⁷⁻⁹⁾ In continuation of our search for new plant secondary metabolites,¹⁰⁾ we have investigated the whole plant of C. pulcherrima, and report here the isolation, structure elucidation and antimicrobial activity of two new homoisoflavanoids, (E)-7-methoxy-3-(4'-methoxybenzylidene)chroman-4-one (1) and (E)-7-hydroxy-3-(3',4',5'-trimethoxybenzylidene)chroman-4-one (5), together with three known homoisoflavonoids, (Z)-7-hydroxy-3-(4'methoxybenzylidene)chroman-4-one (isobonducellin) (2), (E)-7-hydroxy-3-(4'-methoxybenzylidene)chroman-4-one (bonducellin) (3) and (E)-7-hydroxy-3-(2',4'-dimethoxybenzylidene)chroman-4-one (4).

Results and Discussion

Compound 1, was isolated as an amorphous powder. Its molecular formula was assigned as $C_{18}H_{16}O_4$ from its mass spectrum (M⁺ at *m/z* 296), elemental analysis and ¹³C-NMR spectrum, with eleven degree of unsaturation. The strong IR absorptions at 1659 (C=O), 1604 (C=C), 829 cm⁻¹ (*para* substituted benzene ring), UV absorption maxima at 356 and 320 nm and negative ferric chloride test indicated that 1 was a non-phenolic unsaturated homoisoflavanone.⁸⁾

The ¹H-NMR spectrum of **1** revealed the presence of two methoxyl groups as a singlet at δ 3.82 integrating for 6 pro-





* To whom correspondence should be addressed. e-mail: cvr_svu@yahoo.com

tons. The EI-MS of 1 showed two retro-Diels-Alder fragments at m/z 151 and m/z 146 indicating the presence of one methoxyl group in ring-A and another methoxyl group in ring-B, respectively. The correlations observed in the NOESY spectrum of 1 (Fig. 3) indicated that the two methoxyl groups in 1 were placed at C-7 and C-4' position. The ¹H-NMR spectrum of **1** showed an ABX spin coupled system at δ 7.79 (1H, d, J=8.8 Hz), 6.68 (1H, dd, J=8.8, 1.9 Hz) and 6.54 (1H, d, J=1.9 Hz) was assigned to H-5, H-6 and H-8, respectively. Further it also revealed the presence of AA' XX' spin coupled system with two protons centered at δ 7.42 (d, J=8.5 Hz) and another two protons at δ 7.04 (d, J=8.5 Hz), which were assigned to the H-2', 6' and H-3', 5' protons, respectively. The E-geometry of the double bond at C-3 and C-9 in 1 was clearly indicated by δ values of the methylene protons at δ 5.40 for C-2 and the vinylic proton at



Fig. 2. Structure of Compound 2



Fig. 3. Significant HMBC (\longrightarrow) and NOESY (\longleftarrow) Correlations for 1 and 5

Table 1. 13 C-NMR Data (125, 75 MHz, CDCl₃ and DMSO- d_6) of Compounds 1, 5, 2, 3 and 4

Carbon	1	5	2	3	4
2	67.7	67.5	75.0	67.2	67.4
3	126.4	129.5	126.6	126.6	128.3
4	179.6	179.1	181.6	180.2	179.6
4a	105.4	107.8	115.5	114.3	115.3
5	128.9	129.5	129.1	129.2	128.8
6	110.3	111.2	110.6	110.8	110.5
7	165.5	164.7	164.3	164.3	164.1
8	100.6	102.4	102.1	102.4	102.1
8a	162.5	162.6	162.6	162.5	158.9
9	135.6	135.6	138.6	135.5	131.0
1'	128.5	130.2	127.2	128.7	114.0
2'	132.3	107.8	132.5	131.4	161.7
3'	114.3	152.8	112.9	113.7	99.8
4'	160.3	138.4	160.2	159.9	162.2
5'	114.3	152.8	112.9	113.7	104.4
6'	132.3	107.8	132.5	131.4	130.8
7-OMe	55.8	_	_	_	_
2'-OMe	_	_	_	_	55.6
3'-OMe		56.0	_	_	_
4'-OMe	55.8	60.1	54.8	54.9	54.8
5'-OMe	—	56.0	_	—	—

 δ 7.65 for C-9, since in the case of the *Z*-geometry positions the H-9 away from the anisotropic region of the carbonyl group should resonate at a higher field.^{11,12} Therefore the structure of **1** was characterized as (*E*)-7-methoxy-3-(4'methoxybenzylidene)chroman-4-one.

Compound **5** was isolated as a yellow colour solid. It was analyzed for $C_{19}H_{19}O_6$ from its mass spectrum (M⁺ at *m/z* 342), elemental analysis and ¹³C-NMR spectrum with eleven degree of unsaturation. The UV absorption bands at 355 and 310 nm, the characteristic IR absorption bands at 3233 (OH), 1651 (C=O), 1618 (C=C), 847 cm⁻¹ (trisubstituted benzene ring) and positive ferric chloride test indicated that compound **5** was also a unsaturated homoisoflavanone⁸⁾ containing phenolic hydroxyl group.

The fragment peaks at m/z 167 and m/z 137 attributable to a retro-Diels-Alder reaction in the EI-MS of 5, suggested that three methoxyl groups were present in the ring-B and a hydroxyl group in ring-A respectively. The ¹H-NMR spectrum of 5 was closely related to that of 1, suggesting that compound 5 also possessed a homoisoflavanone skeleton with three methoxyl and a hydroxyl groups. However, instead of the AA'XX' coupling system in 1, compound 5 possessed a symmetric two proton singlet at δ 6.72 (2H, s) assigned to C-2' and C-6' positions and two sharp singlets at δ 3.82 (6H, s) and 3.70 (3H, s) assigned for three symmetric methoxyl groups at C-3', C-4', and C-5' positions, respectively. The placements of these three methoxyl groups at C-3', C-4', and C-5' were confirmed by the NOESY and HMBC correlations (Fig. 3). The hydroxyl group was placed at C-7 as this carbon signal was shifted downfield by 29.3 ppm in its ¹³C-NMR spectrum.¹³⁾ Thus the structure of compound 5 was characterized as (E)-7-hydroxy-3-(3',4',5'-trimethoxybenzylidene) chroman-4-one.

The known compounds, (*Z*)-7-hydroxy-3-(4'-methoxybenzylidene)chroman-4-one (isobonducellin) (**2**), (*E*)-7-hydroxy-3-(4'-methoxybenzylidene)chroman-4-one (bonducellin) (**3**) and (*E*)-7-hydroxy-3-(2',4'-dimethoxybenzylidene)- chroman-4-one (4) were identified by comparison of the spectral data in the literatures.⁷⁻⁹

Experimental

General Procedures Melting points were determined on a Kofler hotstage apparatus and are uncorrected. UV spectra were recorded in MeOH on a Shimadzu UV-240 spectrophotometer and IR spectra were recorded in KBr discs on a Bio-rad win FT-IR spectrophotometer. ¹H- and ¹³C-NMR spectra were determined on a Bruker 300, 500 Spectrometer using DMSO-*d*₆, and CDCl₃ with TMS as an internal standard. ¹H–¹H COSY, HSQC, HMBC and the phase-sensitive NOESY (with 150 ms mixing time) spectra were recorded using the standard pulse sequences. EI-MS were recorded at 70 eV (direct probe) on a Nermag R10-10 mass spectrometer. LC-MS was recorded on a AGILENT-1100 periods LC/MSD (VL). Column chromatography (CC) was performed on acme silica gel finer than 200 mesh (0.08 nm).

Plant Material The whole plant of *C. pulcherrima* was collected in December 2004 at Tirumala Hills, Tirupati, Andhra Pradesh, India.

Extraction and Isolation The air-dried and powdered whole plant (3.5 kg) of *C. pulcherrima* was successively extracted with *n*-hexane, Me₂CO and MeOH. The *n*-hexane extract was purified over a silica gel column using *n*-hexane and EtOAc and the fractions eluted with 5% EtOAc in hexane afforded compound 1 (24 mg). Similarly the acetone extract was chromatographed using hexane and EtOAc and their step gradient to obtain a mixture containing **2** and **3** (35 mg). Further elution of the column with 40% EtOAc in hexane afforded **4** (16 mg) and **5** (21 mg). The mixture was rechromatographed using step gradient elution with 25% and 30% EtOAc in hexane to yield **2** (12 mg) and **3** (18 mg) respectively.

(*E*)-7-Methoxy-3-(4'-methoxybenzylidene)chroman-4-one (1): Amorphous powder, mp 130—132 °C (hexane–EtOAc). UV λ_{max} (MeOH) nm (log ε): 356 (4.44), 320 (4.21). IR (KBr) v_{max} cm⁻¹: 2966, 2841, 1659 (C=O), 1604 (C=C), 1587, 1456, 1259, 1157, 1027, 829. ¹H-NMR (DMSO- d_6) δ : 7.79 (1H, d, J=8.8 Hz, H-5), 7.65 (1H, s, H-9), 7.42 (2H, d, J=8.5 Hz, H-2', H-6'), 7.04 (2H, d, J=8.5 Hz, H-3', H-5'), 6.68 (1H, dd, J=8.8, 1.9 Hz, H-6), 6.54 (1H, d, J=1.9 Hz, H-8), 5.40 (2H, s, H-2), 3.82 (6H, s, OMe-7, OMe-4'). ¹³C-NMR (DMSO- d_6): see Table 1. EI-MS m/z (%): 296 [M]⁺ (100), 295 (62), 151 (56), 146 (27), 131 (40), 108 (74), 77 (18). LC-MS m/z 297 [M+H]⁺ and 319 [M+Na]⁺. Anal. Calcd for C₁₈H₁₆O₄: C, 72.97; H, 5.40; Found: C, 72.93; H, 5.45.

(*E*)-7-Hydroxy-3-(3',4',5'-trimethoxybenzylidene)chroman-4-one (**5**): Yellow colour solid, mp 202—204 °C (hexane–EtOAc). UV λ_{max} (MeOH) nm (log ε): 355 (4.60), 310 (3.91). IR (KBr) v_{max} cm⁻¹: 3233 (OH), 2924, 1651 (C=O), 1618 (C=C), 1582, 1468, 1259, 1158, 1008, 847. ¹H-NMR (DMSO- d_6) & 10.6 (1H, s, OH), 7.73 (1H, d, J=8.7 Hz, H-5), 7.63 (1H, s, H-9), 6.72 (2 H, s, H-2', H-6'), 6.55 (1 H, dd, J=8.7, 1.9 Hz, H-6), 6.32 (1H, d, J=1.9 Hz, H-8), 5.41 (2H, s, H-2), 3.82 (6H, s, OMe-3' OMe-4'), 3.70 (3H, s, OMe-5'). ¹³C-NMR (DMSO- d_6): see Table 1. EI-MS m/z (%): 342 [M]⁺ (100), 341 (43), 280 (27), 181 (43), 175 (32), 167 (70), 137 (92), 107 (27), 77 (18). LC-MS m/z 341 [M–H]⁻. *Anal.* Calcd for C₁₉H₁₉O₆: C, 66.66; H, 5.55; Found: C, 66.70; H, 5.61.

References

- Gamble J. S., "Flora of the Presidency of Madras," Vol. 1, Botanical Survey of India, Calcutta, 1956, p. 279.
- Kirtikar K. R., Basu B. D., "Indian Medicinal Plants," Vol. 2, Periodical Experts, New Delhi, 1935, pp. 848–849.
- Che C. T., McPherson D. D., Cordell G. A., Fong, H. H. S., J. Nat. Prod., 49, 561—569 (1986).
- McPherson D. D., Che T. T., Cordell G. A., Soejarto D. D., Pezzuto J. M., Fong H. H. S., *Phytochemistry*, 25, 167–170 (1986).
- Patel A. D., Freyer A. J., Webb R. L., Zuber G., Reichwein R., Bean M. F., Faucettle L., Johnson R. K., *Tetrahedron*, 53, 1583–1592 (1997).
- Ragasa C. Y., Hofilena J. G., Rideout J. A., J. Nat. Prod., 65, 1107– 1110 (2002).
- Srinivas K. V. N. S., Koteswara Rao Y., Mahender I., Das B., Rama Krishna K. V. S., Hara Kishore K., Murty U. S. N., *Phytochemistry*, 63, 789–793 (2003).
- McPherson D. D., Cordell G. A., Soejarto D. D., Pezzuto J. M., Fong H. H. S., *Phytochemistry*, 22, 2835–2838 (1983).
- Zhao P., Iwamoto Y., Kouno I., Egami Y., Yamamoto H., *Phytochem*istry, 65, 2455–2461 (2004).
- 10) Maheswara M., Koteswara Rao Y., Siddaiah V., Venkata Rao C., So-

- mayajulu K. V., Lin F. T., Chem. Pharm. Bull., 52, 974-975 (2004).
- Bohler P., Tamm Ch., *Tetrahedron Lett.*, 36, 3479–3483 (1967).
 Heller W., Tamm C., *Progress in the Chemistry of Organic Natural*

Products, 40, 105-152 (1981).

13) Agrawal P. K., "Carbon-13 NMR of Flavonoids," Elsevier, Amsterdam, 1989, p. 259.