

Withangulatin I, a New Cytotoxic Withanolide from *Physalis angulata*

Shwu-Woan LEE,^a Min-Hsiung PAN,^b Chiu-Ming CHEN,^c and Zong-Tsi CHEN^{*,a}

^aDepartment of Applied Chemistry, Chia-Nan University of Pharmacy and Science; Tainan 717, Taiwan, R.O.C.:

^bDepartment of Seafood Science, National Kaohsiung Marine University; Kaohsiung 811, Taiwan, R.O.C.: and

^cDepartment of Chemistry, National Tsing Hua University; Hsinchu 300, Taiwan, R.O.C.

Received October 11, 2007; accepted November 7, 2007; published online November 22, 2007

A new withanolide, withangulatin I (**2**), was isolated from the whole plant of *Physalis angulata*. Its structure was established as (20*S*,22*R*)-15 α -acetoxy-5 β ,6 β -epoxy-14 α -hydroxy-1,4-dioxo-witha-2,16,24-trienolide on the basis of chemical and spectroscopic methods including 2D-NMR and circular dichroism (CD) experiments. Withangulatin A (**1**) and withangulatin I (**2**) were tested for their cytotoxic activities against two human cancer cell lines, colorectal carcinoma COLO 205 and gastric carcinoma AGS, *in vitro*. Compounds **1** and **2** exhibited inhibitory activities against these two human cancer cells with IC₅₀ values of 16.6 and 1.8 and 53.6 and 65.4 μ M, respectively.

Key words *Physalis angulata*; Solanaceae; withanolide; withangulatin; cytotoxic activity

Physalis angulata L. is an annual herb belonging to the family Solanaceae and is widely distributed in tropical and temperate regions. It is a common herbal plant, known in Chinese as kuzhi, which has been used as a folk medicine with antiinflammatory, antitussive, antipyretic, diuretic, antidotal, and antitumor effects in Taiwan.¹ The extracts and components of this plant were found to have antitumor,^{2,3} cytotoxic,^{4–8} inhibition of the ubiquitin-proteasome pathway,⁹ immunomodulatory,¹⁰ antimycobacterial,¹¹ antinociceptive, antiinflammatory, and antiallergic activities.¹²

Physalis angulata is known to contain a wide variety of pharmacologically important steroidal compounds, such as withasteroids.⁴ To date, phytochemical studies of this plant have described the isolation of numerous phytochemicals, including withasteroids,^{4–6,8,13–17} flavonoid glycoside,¹⁸ and alkaloids.¹⁹ In a previous study, we isolated a withanolide, withangulatin A (**1**),¹⁵ with significant biological activities²⁰ from this plant as a part of our search for bioactive constituents from natural sources. Here, we report the isolation and structural elucidation of a minor withanolide, withangulatin I (**2**), from this plant. Its structure was established as (20*S*,22*R*)-15 α -acetoxy-5 β ,6 β -epoxy-14 α -hydroxy-1,4-dioxo-witha-2,16,24-trienolide on the basis of extensive spectroscopic data interpretation, chemical transformation, and circular dichroism (CD) experiments. The cytotoxic activities of compounds **1** and **2** against human colorectal carcinoma COLO 205 and human gastric carcinoma AGS cell lines were also evaluated *in vitro*.

Results and Discussion

As in the previously reported procedure,¹⁵ the fractions

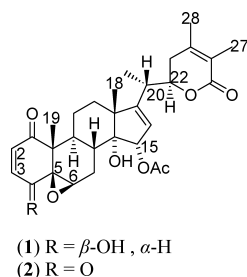


Fig. 1. Structures of Withangulatin A (**1**) and Withangulatin I (**2**)

containing withanolides were combined. The combined fractions were further purified by repeated column chromatography and preparative TLC to afford a minor withanolide, withangulatin I (**2**). The structure of **2** was established based on detailed spectral analyses, chemical transformation, and comparison with the spectral data reported in the literature.

Compound **2** was isolated as a pale yellow, amorphous powder. The IR spectrum of **2** displayed absorption bands of hydroxyl (3450 cm^{-1}), α,β -unsaturated δ -lactone (1710 cm^{-1}), and α,β -unsaturated ketone (1690 cm^{-1}) functions. The UV spectrum of **2** showed absorption at λ_{max} (MeOH) 227 nm, which also implied the presence of α,β -unsaturated δ -lactone and α,β -unsaturated ketone moieties.²¹ The molecular formula of **2** was determined to be C₃₀H₃₆O₈ by high-resolution (HR)-EI-MS at m/z 524.2413 (M⁺, Calcd 524.2410). The ¹H- and ¹³C-NMR spectral data (Table 1) showed four methyl singlets (δ 1.09, 1.38, 1.85, 1.92), a secondary methyl (δ 1.12, d, $J=7.0$ Hz), a 5 β ,6 β -epoxy group [δ_{H} 3.56 (1H, br d, $J=2.2$ Hz); δ_{C} 63.5 (s), 64.0 (d)], an acetoxy group [δ_{H} 1.97 (Me, s); δ_{C} 21.4 (q), 169.6 (s)], an α,β -unsaturated δ -lactone [δ_{C} 148.5 (C-24), 122.2 (C-25), 166.3 (C-26)], two oxygenated methines [δ_{H} 5.22 (d, $J=2.6$ Hz, H-15); δ_{C} 83.6 (C-15)] and [δ_{H} 4.23 (ddd, $J=12.5, 7.0, 3.0$ Hz, H-22); δ_{C} 79.2 (C-22)], a trisubstituted double bond [δ_{H} 5.73 (d, $J=2.6$ Hz, H-16); δ_{C} 121.2 (C-16), 162.3 (C-17)], and a Δ^2 -1,4-dione moiety [δ_{H} 6.85, 6.89 (each 1H, d, $J=10.3$ Hz, H-2 and H-3, respectively); δ_{C} 202.2 (C-1), 139.4 (C-2), 141.2 (C-3), 193.6 (C-4)].²² The EI-MS spectrum showed a base peak at m/z 125 which confirmed the presence of the α,β -unsaturated δ -lactone moiety.²³ These data revealed that compound **2** is a typical withanolide bearing an acetoxy group and a Δ^2 -1,4-dione moiety. The deshielding proton

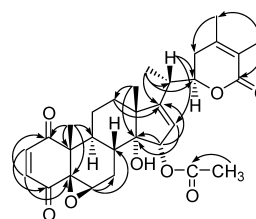


Fig. 2. Key HMBC Correlations Observed in **2**

Table 1. ^1H - (500 MHz) and ^{13}C -NMR (125 MHz) Spectral Data of Withangulatin I (**2**) in CDCl_3

Position	^{13}C	DEPT	^1H	HMBC ($^1\text{H}\rightarrow^{13}\text{C}$)	NOESY
1	202.2	C			
2	139.4	CH	6.85 (d, $J=10.3$ Hz)	C-4	
3	141.2	CH	6.89 (d, $J=10.3$ Hz)	C-1, 5	
4	193.6	C			
5	63.5	C			
6	64.0	CH	3.56 (br d, $J=2.2$ Hz)	C-7, 8	H-7 α , 7 β
7	24.5	CH ₂	2.65 (m, 7 β) 1.67 (m, 7 α)		H-6, 7 α , 8, 15 H-6, 7 β , 9
8	34.5	CH	1.97 (m)		H-7 β , 11 β , 15, 18, 19
9	38.3	CH	2.36 (m)		H-7 α , 11 α
10	50.0	C			
11	23.0	CH ₂	2.02 (m, 11 α) 1.41 (m, 11 β)		H-9, 11 β , 12 β H-8, 11 α , 18, 19
12	37.6	CH ₂	1.80 (m, 12 β) 1.64 (m, 12 α)		H-11 β , 18 H-9
13	52.1	C			
14	81.1	C			
15	83.6	CH	5.22 (d, $J=2.6$ Hz)	C-13, 14, 17	H-8, 16, 18
16	121.2	CH	5.73 (d, $J=2.6$ Hz)	C-13, 14, 15	H-15, 21, 22
17	162.3	C			
18	16.0	CH ₃	1.09 (s)	C-12, 13, 14, 17	H-8, 11 β , 12 β , 15
19	19.2	CH ₃	1.38 (s)	C-1, 5, 9, 10	H-8, 11 β
20	35.4	CH	2.51 (m)	C-16, 17, 21, 22	H-21, 22
21	17.6	CH ₃	1.12 (d, $J=7.0$ Hz)	C-17, 20, 22	H-16, 20, 22
22	79.2	CH	4.23 (ddd, $J=12.5, 7.0, 3.0$ Hz)		H-16, 20, 21, 23 α
23	33.0	CH ₂	2.42 (m, 23 β) 2.18 (dd, $J=17.6, 3.0$ Hz, 23 α)		H-23 α , 28 H-22, 23 β , 28
24	148.5	C			
25	122.2	C			
26	166.3	C			
27	12.4	CH ₃	1.85 (s)	C-24, 25, 26, 28	
28	20.4	CH ₃	1.92 (s)	C-23, 24, 25	H-23 α , 23 β
15-OAc	21.4	CH ₃	1.97 (s)	15-C=O	
	169.6	C			

signals at δ 5.22 (d, $J=2.6$ Hz, H-15) showed NOESY cross-peaks with signals at δ 1.97 (H-8) and 1.09 (H-18) indicating a 15 β -proton and the acetoxy group was located at C-15 with the α -orientation.¹⁵⁾ The above ^1H -NMR data and the ^{13}C -NMR chemical shifts for C-14, C-15, C-16, and C-17 at δ 81.1, 83.6, 121.2, and 162.3, respectively, indicated that **2** comprises a 14 α -hydroxy-15 α -acetoxy-16-ene system in ring D.^{6,8,13,15)} The structure of the Δ^2 -1,4-dione moiety in ring A was further confirmed on the basis of the following HMBC correlations: H-2 with C-4; H-3 with C-1 and C-5; H-19 with C-1, C-5, C-9, and C-10; and H-6 with C-4, C-7, and C-8. The ^1H - and ^{13}C -NMR spectral data of **2** were similar to those of withangulatin A (**1**), except for the signals due to ring A. The Cotton effects of **2** ($\Delta\epsilon+6.03$ at 248 nm and $\Delta\epsilon-0.87$ at 352 nm) were similar to those of withaperuvin E,²²⁾ bearing the same structure of rings A, B, and the α,β -unsaturated δ -lactone moiety, indicating a 22*R* configuration and *cis*-fusion of rings A and B. Based on the above evidence, compound **2** was assumed to be 4-oxowithangulatin A, which was confirmed by manganese dioxide oxidation²¹⁾ of **1** to **2**. Therefore the structure of withangulatin I (**2**) was considered to be (2*S*,22*R*)-15 α -acetoxy-5 β ,6 β -epoxy-14 α -hydroxy-1,4-dioxo-witha-2,16,24-trienolide.

Biological Studies The cytotoxic activities of compounds **1** and **2** isolated from *P. angulata*, along with doxorubicin and camptothecin (as the positive control) against two human cancer cell lines, COLO 205 (colorectal carcinoma)

Table 2. Cytotoxic Activities of Compounds **1** and **2** against Colorectal Carcinoma COLO 205 and Gastric Carcinoma AGS Cell Lines

Compounds	IC ₅₀ [μM] ^{a)}	
	COLO 205	AGS
Withangulatin A (1)	16.6 \pm 0.5	1.8 \pm 0.1
Withangulatin I (2)	53.6 \pm 0.5	65.4 \pm 4.2
Doxorubicin ^{b)}	4.8 \pm 0.5	0.9 \pm 0.1
Camptothecin ^{b)}	>100	1.2 \pm 0.1

a) IC₅₀ is defined as the concentration that required to reduce the number of viable cells by 50% and the results are means \pm standard deviation of three independent experiments. b) Positive control compound.

and AGS (gastric carcinoma) were evaluated *in vitro*. The results (as shown in Table 2) were expressed as the concentration inhibiting 50% of the cell growth (IC₅₀). Compounds **1** and **2** exhibited inhibitory activities against COLO 205 and AGS cancer cells with IC₅₀ values of 16.6 and 1.8 and 53.6 and 65.4 μM , respectively. Previous structure-activity studies of withanolides indicated the importance of the 4 β -hydroxy-2-ene-1-one and 5 β ,6 β -epoxy moieties for biological activities.^{24,25)} Compound **1**, bearing the same moieties mentioned above, showed stronger cytotoxic activities compared with **2**, bearing Δ^2 -1,4-dione and 5 β ,6 β -epoxy moieties. These findings reveal that the 4-hydroxy group is a cytotoxic activity contributor to withanolide and the presence of 4-oxo group

appears to induce a decrease in the cytotoxic activity of withanolide.

Experimental

General Optical rotation was measured on a JASCO DIP-360 digital polarimeter. UV spectra were measured on a Hitachi 200 spectrophotometer. IR spectra were recorded on a Perkin Elmer 781 infrared spectrophotometer. CD spectra were recorded with a Jasco J-720 spectropolarimeter. ¹H-NMR (500 MHz), ¹³C-NMR (125 MHz), and 2D-NMR spectra were recorded on a Bruker AV-500 spectrometer. EI-MS and HR-EI-MS were recorded on a JMS-700 mass spectrometer. Column chromatography was performed on silica gel 60 (Merck, 70–230 mesh), and TLC and preparative TLC were performed on precoated silica gel plates (Merck, Kieselgel 60 F₂₅₄, 0.25 mm and 1.00 mm, respectively).

Plant Material The entire plant of *P. angulata* was collected in Puli, Nantou county, Taiwan, and authenticated by Prof. Chang-Sheng Kuoh, Department of Biology, National Cheng Kung University. A voucher specimen was deposited in the natural product laboratory of the Department of Applied Chemistry, Chia-Nan University of Pharmacy and Science, Tainan, Taiwan.

Extraction and Isolation The dried, powdered whole plant of *P. angulata* (1.0 kg) was extracted with MeOH (9.0 l) under room temperature for 1 week. The concentrated MeOH extract (95 g) was chromatographed on a silica gel column with a CHCl₃/acetone mixture of increasing polarity as eluents, and each fraction was monitored by TLC. The fractions eluted with CHCl₃/acetone (5:3) were combined and evaporated to give a residue (1.8 g) that was further chromatographed on a silica gel column with *n*-hexane/EtOAc/MeOH (8:5:0.5) as eluent. The fractions containing withanolides were collected and further purified with preparative TLC and developed with *n*-hexane/EtOAc/MeOH (4:3:0.3) to give compound **2** (2.1 mg).

Withangulatin I (**2**): Pale yellow, amorphous powder; $[\alpha]_D^{27} +200.1^\circ$ ($c=0.050$ CDCl₃); UV (MeOH) λ_{max} (log ϵ) 227 (4.30) nm; IR (KBr) ν_{max} 3450, 3020, 1740, 1710, 1690, 1465, 1380, 1260, 1250, 1140, 790 cm⁻¹; CD (MeOH, c 1.21 × 10⁻⁴) $\Delta\epsilon$ (nm): +6.03 (248), -0.87 (352); ¹H-NMR (CDCl₃, 500 MHz): see Table 1; ¹³C-NMR (CDCl₃, 125 MHz): see Table 1; EI-MS m/z (75 eV, rel. int. %): 524 (M⁺, 6), 509 (8), 482 (20), 464 (10), 449 (7), 260 (12), 125 (100); HR-EI-MS: m/z 524.2413 (M⁺, Calcd for C₃₀H₃₆O₈ 524.2410).

Oxidation of 1 Compound **1** (5.1 mg) in 15 ml of chloroform was stirred with activated MnO₂ (30 mg) for 48 h. The mixture was filtered and the filtrate was evaporated to dryness. The dried residue was subjected to preparative TLC with *n*-hexane/EtOAc/MeOH (4:3:0.3) as eluent to afford a pale yellow powder (2.6 mg), which was identical to withangulatin I (**2**) (¹H-NMR, IR, and MS).

Cytotoxicity Assay Standard natural product anticancer agents (doxorubicin and camptothecin) were obtained from commercial sources. The cytotoxicities of compounds **1**, **2**, and the two anticancer agents against the two human cancer cell lines COLO 205 and AGS were determined using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay.²⁶ Human cancer cells and selected cells were plated at a density of 1 × 10⁵ cells/ml into 24-well plates. After overnight culture, cells were pre-treated with a series of concentrations of compounds **1** and **2** for 24 h. The final concentration of dimethyl sulfoxide in the culture medium was <0.05%. At the end of treatment, 30 μ l of MTT was added, and the cells were incubated for an additional 4 h. Cell viability was determined by scanning with an enzyme-linked immunosorbent assay reader with a 570-nm filter.

Acknowledgments We thank the Chia-Nan University of Pharmacy and Science, Taiwan, R.O.C., for financial support of this work and Prof. Chang-

Sheng Kuoh, Department of Biology, National Cheng Kung University, for authentication of plant material. The authors also thank Misses J. Z. Wu and L. N. Lai of the Department of Chemistry, National Cheng Kung University, for NMR and MS analyses, respectively.

References

- 1) Chiu N. Y., Chang K. S., "The Illustrated Medicinal Plants of Taiwan," Vol. II, Southern Materials Center, Inc., Taipei, 1991, p. 191.
- 2) Wu S. J., Ng L. T., Chen C. H., Lin D. L., Wang S. S., Lin C. C., *Life Sci.*, **74**, 2061–2073 (2004).
- 3) Magalhaes H. I., Veras M. L., Torres M. R., Alves A. P., Pessoa O. D., Silva E. R., Costa-Lotufo L. V., de Moraes M. O., Pessoa C., *J. Pharm. Pharmacol.*, **58**, 235–241 (2006).
- 4) Damu A. G., Kuo P. C., Su C. R., Kuo T. H., Chen T. H., Bastow K. F., Kee K. H., Wu T. S., *J. Nat. Prod.*, **70**, 1146–1152 (2007).
- 5) Kuo P. C., Kuo T. H., Damu A. G., Su C. R., Lee E. J., Wu T. S., Shu R., Chen C. M., Bastow K. F., Chen T. H., Lee K. H., *Org. Lett.*, **8**, 2953–2956 (2006).
- 6) He Q. P., Maa L., Luo J. Y., He F. Y., Lou L. G., Hu L. H., *Chem. Biodivers.*, **4**, 443–449 (2007).
- 7) Hsieh W. T., Huang K. Y., Lin H. Y., Chung J. G., *Food Chem. Toxicol.*, **44**, 974–983 (2006).
- 8) Nagafuji S., Okabe H., Akahane H., Abe F., *Biol. Pharm. Bull.*, **27**, 193–197 (2004).
- 9) Ausseil F., Samson A., Aussagues Y., Vandenberghe I., Creancier L., Pouny I., Kruczynski A., Massiot G., Bailly C., *J. Biomol. Screen.*, **12**, 106–116 (2007).
- 10) Soares M. B., Brustolim D., Santos L. A., Bellintani M. C., Paiva F. P., Ribeiro Y. M., Tomassini T. C., Dos Santos R., *Int. Immunopharmacol.*, **6**, 408–414 (2006).
- 11) Pietro R. C., Kashima S., Sato D. N., Januario A. H., Franca S. C., *Phytomedicine*, **7**, 335–338 (2000).
- 12) Choi E. M., Hwang J. K., *J. Ethnopharmacol.*, **89**, 171–175 (2003).
- 13) Abe F., Nagafuji S., Okawa M., Kinjo J., *Chem. Pharm. Bull.*, **54**, 1226–1228 (2006).
- 14) Shingu K., Yahara S., Okabe H., Nohara T., *Chem. Pharm. Bull.*, **40**, 2448–2451 (1992).
- 15) Chen C. M., Chen Z. T., Hsieh C. H., Li W. S., Wen S. Y., *Heterocycles*, **31**, 1371–1375 (1990).
- 16) Januario A. H., Filho E. R., Pietro R. C., Kashima S., Sato D. N., Franca S. C., *Phytother. Res.*, **16**, 445–448 (2002).
- 17) Row L. R., Reddy K. S., Sarma S., Matsuura T., Nakashima R., *Phytochemistry*, **19**, 1175–1181 (1980).
- 18) Ismail N., Alam M., *Fitoterapia*, **72**, 676–679 (2001).
- 19) Basey K., McGaw B. A., Woolley J. G., *Phytochemistry*, **31**, 4173–4176 (1992).
- 20) Lee Y. C., Lai Y. K., *J. Cell. Biochem.*, **57**, 150–162 (1995).
- 21) Lavie D., Glotter E., Shov Y., *J. Chem. Soc.*, **1965**, 7517–7531 (1965).
- 22) Bagchi A., Neogi P., Sahai M., Ray A. B., Oshima Y., Hikino H., *Phytochemistry*, **23**, 853–855 (1984).
- 23) Kupchan S. M., Anderson W. K., Bollinger P., Doskotch R. W., Smith R. M., Renaud J. A. S., Schnoes H. K., Burlingame A. L., Smith D. H., *J. Org. Chem.*, **34**, 3858–3866 (1969).
- 24) Kuroyanagi M., Shibata K., Umehara K., *Chem. Pharm. Bull.*, **47**, 1646–1649 (1999).
- 25) Su B. N., Misico R., Park E. J., Santarsiero B. D., Mesecar A. D., Fong H. H. S., Pezzuto J. M., Kinghorn A. D., *Tetrahedron*, **58**, 3453–3466 (2002).
- 26) Mosmann T., *J. Immunol. Methods*, **65**, 55–63 (1983).