



Table 1. <sup>1</sup>H- and <sup>13</sup>C-NMR Spectral Data of Compounds **1** and **1a** in CD<sub>3</sub>OD

	<sup>1</sup> H		<sup>13</sup> C	
	<b>1</b>	<b>1a</b>	<b>1</b>	<b>1a</b>
2			163.28 (s)	163.58 (s)
3	6.17 (1H, d, <i>J</i> =9.5)	6.18 (1H, d, <i>J</i> =9.5)	112.10 (d)	111.67 (d)
4	7.81 (1H, d, <i>J</i> =9.5)	7.84 (1H, d, <i>J</i> =9.5)	146.08 (d)	146.31 (d)
5	7.34 (1H, d, <i>J</i> =8.5)	7.34 (1H, d, <i>J</i> =8.5)	128.20 (d)	127.97 (d)
6	6.81 (1H, d, <i>J</i> =8.3)	6.84 (1H, d, <i>J</i> =8.5)	113.55 (d)	113.92 (d)
7			160.37 (s)	161.12 (s)
8			114.32 (s)	115.30 (s)
9			155.11 (s)	154.88 (s)
10			113.38 (s)	113.24 (s)
1'a	3.14 (1H, dd, <i>J</i> =14.2, 5.2)	3.18 (1H, dd, <i>J</i> =13.9, 2.4)	26.11 (t)	26.33 (t)
1'b	3.18 (1H, dd, <i>J</i> =14.5, 7.3)	2.92 (1H, dd, <i>J</i> =13.9, 10.3)		
2'	4.06 (1H, dd, <i>J</i> =6.7, 5.5)	3.67 (1H, dd, <i>J</i> =10.3, 2.4)	85.94 (d)	79.39 (d)
3'			73.81 (s)	73.86 (s)
gem-(CH <sub>3</sub> ) <sub>2</sub>	1.22 (3H, s)	1.29 (3H, s)	24.70 (q)	25.42 (q)
	1.31 (3H, s)	1.30 (3H, s)	26.27 (q)	25.49 (q)
G-1	4.15 (1H, d, <i>J</i> =7.3)		103.46 (d)	
G-2	3.15 (1H, dd, <i>J</i> =9.2, 7.4)		74.87 (d)	
G-3	3.18 (1H, dd, <i>J</i> =9.2, 8.6)		77.67 (d)	
G-4	3.26 (1H, dd, <i>J</i> =9.2, 9.0)		71.37 (d)	
G-5	3.10–3.15 (1H, m)		77.84 (d)	
G-6a	3.61 (1H, dd, <i>J</i> =12.1, 5.7)		62.49 (t)	
G-6b	3.76 (1H, dd, <i>J</i> =11.9, 2.4)			

δ in ppm from TMS, *J* value in Hz, r.t.

The structure of **1** was further established by 2D-NMR studies, particularly <sup>1</sup>H–<sup>1</sup>H correlated spectroscopy (COSY), heteronuclear multiple quantum coherence (HMQC) and nuclear Overhauser enhancement and exchange spectroscopy (NOESY). Correlations in HMBC and NOESY revealed proton–carbon sequences from H-2' to C-8 and G-C-1, H-1' to C-7, C-8 and C-9, respectively.

Compounds **2** and **3** are also simple coumarin glycosides. **2** and **3** were identified as apiosylskimmmin<sup>28</sup>) and hymexelsin<sup>29</sup>) by comparison of their structural data with those of authentic samples.

Nonglycosidic simple coumarins **4**, **5**, **6**, and **7** were identified as umbelliferone, scopoletin,<sup>30</sup>) isofraxidin,<sup>31</sup>) and 8-carboxy-7-hydroxy coumarin<sup>32</sup>) based on analysis of their physical and spectral data (MS, <sup>1</sup>H- and <sup>13</sup>C-NMR spectra, <sup>1</sup>H–<sup>1</sup>H COSY, HMQC, HMBC experiments) and by comparison with literature values.

This is the first reported identification of simple coumarin (**7**) as a constituent of plant materials.

## Experimental

**General Experimental Procedures** Melting points were recorded using a Yanagimoto melting point apparatus without correction. UV spectra were recorded using Shimadzu 1600. <sup>1</sup>H- and <sup>13</sup>C-NMR spectra were recorded on a JNM-LA 500 spectrometer and a JEOL AL 400. FAB-MS spectra were obtained using a JMS-DX 302 mass spectrometer. Optical rotations were determined for solutions in MeOH using a JASCO DIP-140 polarimeter.

**Plant Material** *Peucedanum praeruptorum* DUNN. (Umbelliferae) was collected from Ringan Town, Zhejiang Province, P. R. China, in August 2000 during the flowering season. A voucher specimen was identified by Prof. Xiao Luwei at Zhejiang College of Traditional Chinese Medicine and deposited at the herbarium of Meiji Pharmaceutical University.

**Extraction and Isolation** Pulverized roots of *Peucedanum praeruptorum* (2.75 kg) were extracted with water under reflux to give 638.6 g of extract. The water extract (638 g) was fractionated with Diaion HP-20 and then HW-40F to give seven fractions, followed by HPLC with ODS (C18) and/or Develosil (C30) using a MeOH–H<sub>2</sub>O solvent system to yield **1** (98.6 mg), **2** (87.7 mg), **3** (6.3 mg), **4** (15.5 mg), **5** (9.7 mg), **6** (3.9 mg) and **7** (23.4 mg).

**Praeroside VI (1)** White powder; mp 130.5–132.5 °C (decomp.), [α]<sub>D</sub><sup>25</sup> +1.0° (*c*=1.0, MeOH). UV (MeOH): λ<sub>max</sub> (log ε): 326.0 nm (3.70), (MeOH+CH<sub>3</sub>CO<sub>2</sub>Na) 376.5 nm. EI-MS (70 eV): *m/z* (rel. int., %): 426 (M<sup>+</sup>, 1), 408 (2), 264 (26), 247 (38), 246 (34), 229 (29), 205 (64), 176 (100), 175 (86). HR-EI-MS: *m/z*: 426.1525 (Calcd for C<sub>20</sub>H<sub>26</sub>O<sub>10</sub>, 426.1526). <sup>1</sup>H- and <sup>13</sup>C-NMR: see Table 1.

**Enzymatic Hydrolysis of 1** Enzymatic hydrolysis of **1** (20 mg) with cellulase (20 mg) in H<sub>2</sub>O (5 ml) at 40 °C under stirring for 2 d afforded an aglycone (**1a**) and sugar moiety. **1a** (4.5 mg) was isolated from *n*-BuOH soluble part of reaction mixture.

**(2'R)-7-Hydroxy-8-(2',3'-dihydroxy-3'-methylbutyl)-coumarin (1a)** White powder; mp 135.0–137.0 °C (decomp.); [α]<sub>D</sub><sup>25</sup> +46.0° (*c*=0.50, MeOH).

**Apiosylskimmmin (2)** White powder; mp 197.0–198.0 °C (decomp.); [α]<sub>D</sub><sup>25</sup> –69.5° (*c*=1.0, pyridine). UV (MeOH): λ<sub>max</sub> (log ε): 319.0 nm (4.04); FAB-MS: *m/z*: 457 [M+H]<sup>+</sup>, 325 [M–api+H]<sup>+</sup>, 163 [M–(api+glc)+H]<sup>+</sup>.

**Hymexelsin (3)** White powder; [α]<sub>D</sub><sup>25</sup> –104.4° (*c*=0.50, EtOH); UV (MeOH): λ<sub>max</sub> (log ε): 337.5 nm (3.75), 291.0 nm (3.68); FAB-MS: *m/z*: 485 [M–H]<sup>–</sup>.

**Umbelliferone (4)** Colorless needles; mp 226.0–227.5 °C (decomp.); UV (MeOH): λ<sub>max</sub> (log ε): 325.5 nm (3.88). EI-MS (70 eV): *m/z* (rel. int., %): 162 (M<sup>+</sup>, 100), 134 (89), 105 (15), 78 (16), 77 (8).

**Scopoletin (5)** Colorless needles; mp 202.0–203.0 °C (decomp.); UV (MeOH): λ<sub>max</sub> (log ε): 344.0 nm (4.14), 297.5 nm (3.86), 253.5 nm (3.86). EI-MS (70 eV): *m/z* (rel. int., %): 192 (M<sup>+</sup>, 100), 177 (54), 164 (21), 149 (35), 135 (1), 121 (11).

**Isofraxidin (6)** Colorless needles; mp 149.0–150.0 °C (decomp.); UV (MeOH): λ<sub>max</sub> (log ε): 342.0 nm (3.74). EI-MS (70 eV): *m/z* (rel. int., %): 222 (M<sup>+</sup>, 100), 207 (25), 194 (15), 179 (18), 161 (5), 151 (10), 149 (4), 133 (5), 123 (13), 108 (7), 113 (100).

**8-Carboxy-7-hydroxy Coumarin (7)** White powder; mp 210 °C (decomp.); UV (MeOH): λ<sub>max</sub> (log ε): 333.0 nm (4.03), 258.0 nm (3.50). IR: ν<sub>max</sub><sup>KBr</sup> 3600–2600, 1730, 1715, 1610, 1505, 1410, 1360, 1250, 1230, 840 cm<sup>–1</sup>. EI-MS (70 eV): *m/z* (rel. int., %): 206 (M<sup>+</sup>, 7), 188 (23), 162 (97), 160 (19), 134 (100), 132 (6), 105 (18), 78 (20), 77 (11).

**Acknowledgement** The authors are grateful to medical doctor Qian Xiao Hua at Ringan Town Hospital, Zhejiang Province, P. R. China, for support in the collection of Bai-Hua Qianhu, *Peucedanum praeruptorum*.

## References and Notes

- Part XXVI: Okada Y., Ishii H., Zhang Y., Baba M., Okuyama T., *Phar-*

- maceut. Biol.*, to be published.
- 2) Present address: *Food Science & Technology Research Group, Nichirei Foods Inc., Japan.*
  - 3) Okuyama T., Shibata S., *Planta Med.*, **42**, 89—96 (1981).
  - 4) Kozawa T., Sakai K., Uchida M., Okuyama T., Shibata S., *J. Pharm. Pharmacol.*, **33**, 317—320 (1981).
  - 5) Okuyama T., Sakakibara I., Shibata S., *Syoyakugaku-Zasshi*, **35**, 331—339 (1981).
  - 6) Sakakibara I., Okuyama T., Shibata S., *Planta Med.*, **44**, 199—203 (1982).
  - 7) Sakakibara I., Okuyama T., Shibata S., *Planta Med.*, **50**, 117—120 (1984).
  - 8) Asahara T., Sakakibara I., Okuyama T., Shibata S., *Planta Med.*, **50**, 488—492 (1984).
  - 9) Kawasaki C., Okuyama T., Shibata S., Iitaka Y., *Planta Med.*, **50**, 492—496 (1984).
  - 10) Matano Y., Okuyama T., Shibata S., Hoson M., Kawada T., Osada H., Noguchi T., *Planta Med.*, **52**, 135—138 (1986).
  - 11) Nishino H., Nishino A., Okuyama T., Shibata S., *J. Kyoto Pref. Univ. Med.*, **96**, 391—394 (1987).
  - 12) Takata M., Okuyama T., Shibata S., *Planta Med.*, **54**, 323—327 (1988).
  - 13) Okuyama T., Takata M., Shibata S., *Planta Med.*, **55**, 64—67 (1989).
  - 14) Shibata S., Okuyama T., *Abstr. Chin. Med.*, **3**, 214—230 (1989).
  - 15) Takata M., Shibata S., Okuyama T., *Planta Med.*, **56**, 307—311 (1990).
  - 16) Takata M., Shibata S., Okuyama T., *Planta Med.*, **56**, 133 (1990).
  - 17) Nishino H., Okuyama T., Takata M., Shibata S., Tokuda H., Takayasu J., Hasegawa T., Nishino A., Ueyama H., Iwashima T., *Carcinogenesis*, **11**, 1557—1561 (1990).
  - 18) Chang T.-H., Li J.-M., Sun X.-D., Yu Y.-F., Feng W.-Y., Hao L.-Y., Wang Y.-P. W., Zhang K.-Y., Okuyama T., *Shoyakugaku Zasshi*, **47**, 279—282 (1993).
  - 19) Chang T.-H., Adachi H., Mori N., Saito I., Okuyama T., *Eur. J. Pharmacol.*, **258**, 77—84 (1994).
  - 20) Fulan G., Wanbao, J. Xinhua Z., Zhao N., Okuyama T., *J. China Medical University*, **23**, 549—552 (1994).
  - 21) Mizuno A., Okada Y., Nishino H., Okuyama T., *J. Tradit. Med.*, **11**, 220—224 (1994).
  - 22) Zhao N.-C., Jin W.-B., Zhang X.-H., Guan F.-L., Sun Y.-B., Adachi H., Okuyama T., *Biol. Pharm. Bull.*, **22**, 984—987 (1999).
  - 23) Chang H., Okada Y., Okuyama T., Tu P., *Magn. Reson. Chem.*, **45**, 611—614 (2007).
  - 24) Chang H.-T., Okada Y., Ma T.-G., Okuyama T., Tu P.-F., *J. Asian Nat. Prod. Res.*, **10**, 577—581 (2008).
  - 25) Lemmich J., *Phytochemistry*, **38**, 427—432 (1995).
  - 26) Ceccherelli P., *J. Nat. Prod.*, **53**, 536—538 (1990).
  - 27) Derek R. B., *Chem. Commun.*, **2002**, 3070—3071 (2002).
  - 28) Satyanarayana P., *Phytochemistry*, **24**, 1862—1863 (1985).
  - 29) Rao P. S., Asheervadam Y., *J. Nat. Prod.*, **51**, 959—961 (1988).
  - 30) Ito C., Furukawa H., *Chem. Pharm. Bull.*, **35**, 4277—4286 (1987).
  - 31) Tsukamoto H., Hisada S., Nishibe S., *Chem. Pharm. Bull.*, **33**, 4069—4073 (1985).
  - 32) Soine T. O., Erhardt P., Zheleva A., Bubeva-Ivanova L., Mahandru M. M., *J. Pharm. Sci.*, **62**, 1879—1880 (1973).