# Syntheses and Doxorubicin-Inclusion Abilities of $\beta$ -Cyclodextrin Derivatives with a Hydroquinone $\alpha$ -Glycoside Residue Attached at the Primary Side

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This paper describes syntheses and doxorubicin-inclusion abilities of  $\beta$ -cyclodextrin (CyD) derivatives with a hydroquinone  $\alpha$ -glycoside residue attached at the primary side. The hydroquinone glycoside having an  $\alpha$ -Dglucosidic or 2-acetamido-2-deoxy- $\alpha$ -D-glucosidic linkage became a useful component for providing an  $\alpha$ -D-glucose- or 2-acetamido-2-deoxy- $\alpha$ -D-glucose- $\beta$ -CyD conjugate. The surface plasmon resonance analyses of these  $\beta$ -CyD derivatives for the anticancer agent, doxorubicin, indicated that they had excellent inclusion associations on the order of  $10^5 \text{ m}^{-1}$  for the immobilized doxorubicin.

Key words synthesis;  $\beta$ -cyclodextrin; hydroquinone; surface plasmon resonance; saccharide-attached cyclodextrin; doxorubicin

Our recent study showed that the  $\beta$ -glucose- $\beta$ -cyclodextrin (CyD) conjugates 1 and 2 as shown in Fig. 1 were successfully prepared from  $\beta$ -CyD and arbutin (4-hydrox-yphenyl  $\beta$ -glucopyranoside, 3).<sup>1-3)</sup> Arbutin 3 is a naturally occurring hydroquinone glycoside containing a  $\beta$ -D-glucosidic linkage. An appropriate linker necessary to connect 3 with a  $\beta$ -CyD derivative was conveniently introduced into the phenolic alcohol of 3. The inclusion association constants  $(K_{a})$  of 1 and 2 for the immobilized doxorubicin (DXR, anticancer agent) measured using a surface plasmon resonance (SPR) optical biosensor were  $10^5$ — $10^6$  M<sup>-1</sup>. These K<sub>a</sub> values are remarkably high when compared with the inclusion association of the lactose– $\beta$ -CyD conjugate  $4^{4}$  or normal  $\beta$ -CyD which was on the order of  $10^3 \,\mathrm{M}^{-1}$ . The  $\beta$ -CyD derivatives 1 and 2 were characterized by the existence of the phenyl group in the spacer between the glucose and  $\beta$ -CyD. The  $\pi - \pi$  stacking interaction effect between the phenyl group and DXR would increase their inclusion associations for DXR.<sup>3)</sup> Thus, our former study showed that the hydroquinone glycoside could be a synthetically useful component for providing a saccharide- $\beta$ -CyD conjugate which is expected to have an excellent DXR-inclusion ability.5)

The CyD derivatives attached to saccharide moieties are expected to carry drug molecules to specific cells, because they have both the drug-inclusion ability of CyDs and the cell-recognition of saccharides.<sup>4,6–22)</sup> Then, our next objective was the synthesis of a novel  $\beta$ -CyD conjugate having another hydroquinone glycoside. This paper describes the synthesis of two novel  $\beta$ -CyD derivatives attached to a hydroquinone glycoside containing an  $\alpha$ -D-glucopyranosidic or 2-acetamido-2-deoxy- $\alpha$ -D-glucopyranosidic linkage and the evaluation of their DXR-inclusion abilities.

## **Results and Discussion**

We designed two novel  $\beta$ -CyD derivatives **5** and **6** attached to a hydroquinone glycoside containing an  $\alpha$ -D-glucosidic or 2-acetamido-2-deoxy- $\alpha$ -D-glucosidic linkage as shown in Fig. 1.

Chart 1 shows the synthetic procedure of **5** using 4-hydroxyphenyl  $\alpha$ -glucopyranoside ( $\alpha$ -arbutin, 7). The reaction of allyl bromide (1.2 eq) with the dry sodium salt of 7 in N,N-dimethylformamide (DMF) for 70 h gave the allylated compound 8 in 84% yield. The benzylation of 8 using benzyl bromide (4.8 eq) and NaH (16 eq) in DMF for 21 h afforded compound 9 in 99% yield. The hydroboration of 9 with 9borabicyclo[3.3.1]nonane (9-BBN) (2 eq) in tetrahydrofuran (THF) at 0 °C for 5 h, followed by oxidation using aq.  $H_2O_2$ (10 eq) and by hydrolysis with aq. NaOH (3 eq) for 48 h gave compound 10 in 94% yield. The iodination of 10 with iodine (4 eq) in the presence of Ph<sub>3</sub>P (4 eq) in DMF at 35 °C for 2 h quantitatively gave compound 11 in 89% yield. The condensation of 11 (3.8 eq) with the benzylated  $\beta$ -CyD 12<sup>23)</sup> using KOH (ca. 120 eq) and tetra-n-butylammonium iodide (n- $Bu_4NI$ ) (0.5 eq) in DMF for 64 h gave 13 in 69% yield. The treatment of 13 with H<sub>2</sub>-Pd(OH)<sub>2</sub> in DMF for 6 h provided the desired compound 5, which was purified by gel-filtration using LH 20 (MeOH) in 98% yield.



Fig. 1. Saccharide- $\beta$ -CyD Conjugates (1, 2, 4, 5, 6), Arbutin (3) and DXR



a) 1) 0.5 м NaOH aq., H<sub>2</sub>O, 2) AllBr, DMF, 84%, b) BnBr, NaH, DMF, 99%, c) 1) 9-BBN, THF, 2) H<sub>2</sub>O<sub>2</sub> aq., 0.5 м NaOH aq., 94%, d) Ph<sub>3</sub>P, I<sub>2</sub>, DMF, 35 °C, 89%, e) KOH, *n*-Bu<sub>4</sub>NI, DMF, 69%, f) Pd(OH)<sub>2</sub>, H<sub>2</sub> gas, DMF, 98%.

Chart 1. Synthetic Approach to the D-Glucose- $\beta$ -CyD Conjugate (5)



a)  $Yb(OTf)_3$ ,  $BF_3 \cdot OEt_2$ , 4-allyloxyphenol,  $CH_2Cl_2$ , 61%, b) 1)  $O_3$  gas,  $Ph_3P$ , 2)  $NaClO_2$ ,  $NaH_2PO_4$ , 2-methyl-2-butene, *t*-BuOH,  $H_2O$ , 70%, c)  $Me_2P(S)Cl$ , DIEA, DMF, d)  $Pd(OH)_2$ ,  $H_2$  gas, DMF, 56%.

Chart 2. Synthetic Approach to the 2-Acetamido-2-deoxy- $\alpha$ -D-glucose- $\beta$ -CyD Conjugate (6)

Chart 2 shows the synthetic procedure of **6**. The glycosylation of 2-acetamido-3,4,6-tri-*O*-benzyl-2-deoxy- $\alpha$ -D-glucosyl acetate (14)<sup>24</sup> to 4-allyloxyphenol (0.84 eq) was carried out using Yb(OTf)<sub>3</sub> (0.84 eq) and BF<sub>3</sub>·OEt<sub>2</sub> (0.025 eq) in CH<sub>2</sub>Cl<sub>2</sub> at room temperature for 24 h that gave the hydroquinone glycoside derivative **15** in 61% yield.<sup>25)</sup> The ozonization of **15** in the presence of Ph<sub>3</sub>P (10 eq), followed by oxidation using NaClO<sub>2</sub> (10 eq)–NaH<sub>2</sub>PO<sub>4</sub> (1.8 eq) in the presence of 2-methyl-2-butene in *t*-BuOH–H<sub>2</sub>O afforded compound **16** in 70% yield. The condensation of **16** with 6monoamino-6-monodeoxy- $\beta$ -CyD (**17**) using Me<sub>2</sub>P(S)Cl (1.6 eq) in the presence of *N*,*N*-diisopropylethylamine (DIEA) (2 eq) in DMF for 24 h, followed by the debenzylation using H<sub>2</sub>–Pd(OH)<sub>2</sub> produced the desired product **6**, which was purified by gel-filtration using DIAION HP 20 (MeOH) in 56% yield.

Next, the inclusion associations of **5** and **6** with immobilized DXR were estimated by using an SPR optical biosensor according to the reported method.<sup>22)</sup> Figure 2 indicates the kinetic linear plots of **5** and **6**. The association rate constant  $(k_a)$  and dissociation rate constant  $(k_d)$  of **5** (or **6**) were  $1.4 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$   $(1.3 \times 10^3 \text{ M}^{-1} \text{ s}^{-1})$  and  $4.1 \times 10^{-2} \text{ s}^{-1}$   $(2.7 \times 10^{-3} \text{ s}^{-1})$ , and  $K_a$  of **5** (or **6**), calculated by the relationship of  $K_a = k_a/k_d$ , was  $3.5 \times 10^5 \text{ M}^{-1}$  ( $4.6 \times 10^5 \text{ M}^{-1}$ ). The conjugates **5** and **6** indicated high inclusion associations for immobilized DXR. The  $K_a$  values of **5** and **6** were about 100 times higher than that of the  $\beta$ -CyD conjugate **4** having no phenyl group and almost corresponded to those of **1** and **2** of  $10^5 - 10^6 \text{ m}^{-1}$  levels. These results suggested the formation of the  $\pi - \pi$  stacking inclusion complex between the conjugate **5** (or **6**)



Fig. 2. Kinetic Linear Plots for the Immobilized DXR and the Conjugates  $5(\bigcirc; r=0.998)$  and  $6(\Box; r=0.988)$ .



Fig. 3. NOE Interactions Observed in the Inclusion Complex of  ${\bf 5}$  and DXR

and DXR. In the measurement of the <sup>1</sup>H-NOESY spectra of the mixed sample of **5** and DXR (1:1) in D<sub>2</sub>O, the NOE interactions between the protons (H<sub>x</sub>; d; 6.72 ppm and H<sub>y</sub>; d; 7.01 ppm) on the phenyl group and the protons (H<sub>a</sub> and H<sub>c</sub>; br; 7.20 ppm and H<sub>b</sub>; br; 7.46 ppm) on DXR were observed as shown in Fig. 3.<sup>3</sup>) The observation indicated that these protons were located close to each other and there was a strong possibility of the formation of the stacking complex between the phenyl group of **5** and the included DXR, as we speculated. The phenyl group in the conjugates **5** and **6** would also contribute to the increase in the inclusion associations for the immobilized DXR.

### Conclusions

We demonstrated the synthesis of two novel  $\beta$ -CyD derivatives **5** and **6** attached to a hydroquinone glycoside. The hydroquinone glycosides **7** and **15** containing an  $\alpha$ -D-glucosidic or 2-acetamido-2-deoxy- $\alpha$ -D-glucosidic linkage were synthetically useful components for providing these CyDs. The SPR analyses indicated that the conjugates **5** and **6**, which had the phenyl group derived from a hydroquinone glycoside in the spacers between the saccharide and  $\beta$ -CyD, indicated the excellent inclusion associations of  $3.5-4.6 \times 10^5 \text{ M}^{-1}$  for the immobilized DXR. The newly synthesized  $\beta$ -CyD derivatives **5** and **6**, which have high DXR-transportabilities, are promising models of drug-carrying molecules.

#### Experimental

**General** The <sup>1</sup>H-NMR (600 MHz) and <sup>13</sup>C-NMR (150 MHz) spectra were recorded by a JEOL ECA-600 spectrometer. The NOESY experiment was done at 25 °C using a mixing time of 2 ms. Melting points (mp) were measured by a B-545 (BÜCHI Labortechnik AG) and are uncorrected. Optical rotations were recorded by a JASCO DIP-360 digital polarimeter. The HR-MS were obtained using a Mariner spectrometer (PerSeptive Biosystems Inc.). The MALDI-TOF-MS spectra were recorded by a Voyager DE STR spectrometer. Preparative TLC was performed on Merck silica gel 60GF254. The column chromatography was conducted using silica gel 60N (40— 50  $\mu$ m, Kanto Chemical Co., INC.) All anhydrous solvents were purified according to the standard methods. The immobilization of DXR on the sensor cuvette of the SPR optical biosensor was carried out under the same conditions as previously reported.<sup>1–3)</sup> The amount of immobilized DXR was 1.1 ng/mm<sup>2</sup>. The interactions of **5** and **6** with the immobilized DXR were measured at the concentration of  $10^{-6}$ — $10^{-5}$  M in acetate buffer, pH 5.3 at 25 °C.

4-Allyloxyphenyl α-D-Glucopyranoside (8) A 0.5 M NaOH aqueous solution (8 ml, 4 mmol) was added to 7 (1000 mg, 3.7 mmol). After the reaction mixture was stirred for 3 h at room temperature, the solvent was evaporated under reduced pressure and the reaction residue was crystallized. The reaction residue was dissolved in DMF (80 ml) and allyl bromide (413  $\mu$ l, 4.8 mmol) was added. After the reaction mixture was stirred for 70 h, the solvent was evaporated under reduced pressure. The crude product was purified by flash column chromatography on silica-gel (chloroform/methanol= 9/1) to afford **8** (963 mg, 84%) as white crystals. mp: 159.0—160.2 °C;  $[\alpha]_{D}^{23}$ +154° (c=1.1, CH<sub>3</sub>OH); <sup>1</sup>H-NMR (CD<sub>3</sub>OD)  $\delta$ : 3.40 (1H, t, J=11.0 Hz, H-4), 3.53 (1H, m, dd, J=3.4 Hz, J=9.7 Hz, H-2), 3.68—3.72 (2H, m, H-5, H<sub>a</sub>-6), 3.76 (1H, dd, J=4.1 Hz, J=9.6 Hz, H<sub>b</sub>-6), 3.82 (1H, t, J=8.9 Hz, H-3), 4.48 (2H, dt, J=1.3 Hz, J=5.5 Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.21 (1H, dd, J=1.4 Hz, J=10.3 Hz, CH=CH, H<sub>b</sub>), 5.32 (1H, d, J=4.1 Hz, H-1), 5.36 (1H, dd, J=1.4 Hz, J=18.5 Hz, CH=CH<sub>a</sub>H<sub>b</sub>), 6.03 (1H, m, CH=CH<sub>2</sub>), 6.84 (2H, dd, J=2.7 Hz, J=9.6 Hz, Ph), 7.08 (2H, dd, J=2.1 Hz, J=13.1 Hz, Ph); <sup>13</sup>C-NMR (CD<sub>3</sub>OD) δ: 62.4 (C-6), 70.3 (<u>C</u>H<sub>2</sub>CH=CH<sub>2</sub>), 71.6 (C-4), 73.4 (C-2), 74.3 (C-5), 75.0 (C-3), 100.3 (C-1), 116.6 (Ph), 119.6 (Ph), 117.3 (CH=<u>C</u>H<sub>2</sub>), 135.1 (<u>C</u>H=CH<sub>2</sub>), 152.8 (Ph), 155.6 (Ph); HR-MS (ESI) m/z: 335.1124 (Calcd for C<sub>15</sub>H<sub>20</sub>O<sub>7</sub>: M<sup>+</sup>+Na, 335.1101).

4-Allyloxyphenyl 2,3,4,6-Tetra-O-benzyl-α-p-glucopyranoside (9) To a solution of 8 (458 mg, 1.5 mmol) in DMF (30 ml) was added sodium hydride (585 mg, 24.4 mmol) at 0 °C. After the reaction mixture was stirred for 45 min, benzyl bromide (9 ml, 7 mmol) was added. After the reaction mixture was stirred for 21h at room temperature, the reaction was then quenched by adding methanol (50 ml) and water (50 ml). The mixture was extracted with EtOAc (three times) and the combined organic solvent was dried over anhydrous Na2SO4. The organic solvent was filtered and evaporated under reduced pressure. The crude product was purified by flash column chromatography on silica-gel (hexane/ethyl acetate=8/1) to afford 9 (973 mg, 99%) as white crystals. mp: 57.6—58.2 °C;  $[\alpha]_{D}^{23}$  +73° (c=1.3, CHCl<sub>2</sub>); <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 3.59 (1H, dd, J=2.0 Hz, J=10.3 Hz, H<sub>a</sub>-6), 3.69-3.73 (2H, m, H-2, H<sub>b</sub>-6), 3.75 (1H, t, J=9.0 Hz, H-4), 3.91-3.93 (1H, m, H-5), 4.18 (1H, t, J=8.9 Hz, H-3), 4.40 (1H, d, J=12.4 Hz, CH<sub>2</sub>Ph), 4.47-4.50 (3H, m, CH<sub>2</sub>CH=CH<sub>2</sub>, CH<sub>2</sub>Ph), 4.58 (1H, d, J=11.7 Hz,  $CH_2Ph$ ), 4.68 (1H, d, J=12.3 Hz,  $CH_2Ph$ ), 4.79 (1H, d, J=11.7 Hz,  $CH_2Ph$ ), 4.85 (1H, d, J=11.0 Hz, CH<sub>2</sub>Ph), 4.87 (1H, d, J=11.7 Hz, CH<sub>2</sub>Ph), 5.04 (1H, d, J=11.0 Hz, CH<sub>2</sub>Ph), 5.27 (1H, dd, J=1.3 Hz, J=10.3 Hz, CH=CH<sub>2</sub>H<sub>4</sub>), 5.36 (1H, d, J=3.4 Hz, H-1), 5.40 (1H, dd, J=1.4 Hz, J=17.2 Hz, CH= CH<sub>a</sub>H<sub>b</sub>), 6.04 (1H, m, CH=CH<sub>2</sub>), 6.82 (2H, dd, J=2.8 Hz, J=6.9 Hz, Ph), 7.00 (2H, dd, J=2.1 Hz, J=6.9 Hz, Ph), 7.24–7.39 (20H, m, Ph); <sup>13</sup>C-NMR (CDCl<sub>3</sub>) δ: 68.3 (C-6), 69.3 (<u>C</u>H<sub>2</sub>CH=CH<sub>2</sub>), 70.7 (C-5), 73.2 (<u>C</u>H<sub>2</sub>), 73.4 (<u>CH</u><sub>2</sub>), 75.1 (<u>CH</u><sub>2</sub>), 75.7 (<u>CH</u><sub>2</sub>), 77.4 (C-4), 79.7 (C-2), 81.9 (C-3), 96.3 (C-1), 115.4 (Ph), 117.5 (CH=CH<sub>2</sub>), 118.1 (Ph), 127.6–128.4 (Ph), 133.4 (CH=CH<sub>2</sub>), 137.8 (Ph), 138.0 (Ph), 138.2 (Ph), 138.8 (Ph), 150.8 (Ph), 154.0 (Ph); HR-MS (ESI) *m/z*: 695.2985 (Calcd for C<sub>43</sub>H<sub>44</sub>O<sub>7</sub>: M<sup>+</sup>+Na, 695.2979)

4-O-(3-Hydroxypropyl)phenyl 2,3,4,6-Tetra-O-benzyl-α-D-glucopyranoside (10) To a solution of 9 (303 mg, 0.45 mmol) in THF (7 ml) was added in 0.5 M 9-BBN THF solution (2 ml, 1 mmol) at 0 °C. After the reaction mixture was stirred for 5 h at room temperature, a 0.5 M NaOH aqueous solution (2.7 ml, 1.4 mmol) and a 30% H<sub>2</sub>O<sub>2</sub> aqueous solution (0.46 ml, 4.5 mmol) were added. After the reaction mixture was stirred for 48 h, the reaction was then quenched by adding water (10 ml). The mixture was extracted with EtOAc (three times) and the combined organic solvent was dried over anhydrous Na2SO4. The organic solvent was filtered and evaporated under reduced pressure. The crude product was purified by flash column chromatography on silica-gel (hexane/ethyl acetate=3/1) to afford 10 (291 mg, 94%) as a colorless oil.  $[\alpha]_{D}^{23}$  +11° (c=1.1, CHCl<sub>3</sub>); <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ: 1.74 (1H, s, OH), 1.99 (2H, m, CH<sub>2</sub>CH<sub>2</sub>OH), 3.53 (1H, dd, J=1.3 Hz, J=10.3 Hz, H<sub>a</sub>-6), 3.64-3.69 (2H, m, H-2, H<sub>b</sub>-6), 3.71 (1H, t, J=8.9 Hz, H-4), 3.82 (2H, m, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>OH), 3.85-3.88 (1H, m, H-5), 4.03 (2H, t, J=5.5 Hz, CH<sub>2</sub>OH), 4.13 (1H, t, J=9.6 Hz, H-3), 4.36 (1H, d,  $J=11.7 \text{ Hz}, CH_2Ph$ ), 4.44 (1H, d,  $J=10.3 \text{ Hz}, CH_2Ph$ ), 4.53 (1H, d, J=12.3 Hz, CH<sub>2</sub>Ph), 4.63 (1H, d, J=11.7 Hz, CH<sub>2</sub>Ph), 4.75 (1H, d,

4-O-(3-Iodoxypropyl)phenyl 2,3,4,6-Tetra-O-benzyl-α-D-glucopyranoside (11) To a solution of 10 (100 mg, 0.14 mmol) in DMF (3 ml) were added triphenylphosphine (156 mg, 0.6 mmol) and iodine (148 mg, 0.58 mmol) at 35 °C under argon. After the reaction mixture was stirred for 2 h, the reaction was then quenched by adding water (10 ml). The mixture was extracted with EtOAc (three times) and the combined organic solvent was dried over anhydrous Na2SO4. The organic solvent was filtered and evaporated under reduced pressure. The crude product was purified by preparative silica-gel TLC (hexane/ethyl acetate=6/1) to afford 11 (103 mg, 89%) as a colorless oil.  $[\alpha]_{D}^{23} + 102^{\circ}$  (c=1.1, CHCl<sub>3</sub>); <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 2.25 (2H, m, CH<sub>2</sub>CH<sub>2</sub>I), 3.37 (2H, t, J=6.9 Hz, CH<sub>2</sub>I), 3.59 (1H, dd, J=2.0 Hz, J=10.3 Hz, H<sub>a</sub>-6), 3.69—3.74 (2H, m, H-2, H<sub>b</sub>-6), 3.75 (1H, t, J=10.3 Hz, H-4), 3.91 (1H, m, H-5), 3.98 (2H, t, J=5.5 Hz, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>I), 4.18 (1H, t, J=8.9 Hz, H-3), 4.41 (1H, d, J=12.4 Hz, CH<sub>2</sub>Ph), 4.49 (1H, d, J=11.0 Hz, CH<sub>2</sub>Ph), 4.58 (1H, d, J=11.7 Hz, CH<sub>2</sub>Ph), 4.68 (1H, d, J=11.7 Hz, CH<sub>2</sub>Ph), 4.79 (1H, d, J=12.4 Hz, CH<sub>2</sub>Ph), 4.85 (1H, d, J=10.4 Hz, CH<sub>2</sub>Ph), 4.87 (1H, d, J=11.0 Hz,  $CH_2Ph$ ), 5.04 (1H, d, J=10.3 Hz,  $CH_2Ph$ ), 5.36 (1H, d, J=3.4 Hz, H-1), 6.80 (2H, d, J=8.9 Hz, Ph), 7.00 (2H, d, J=8.9 Hz, Ph), 7.13—7.39 (20H, m, Ph); <sup>13</sup>C-NMR (CDCl<sub>3</sub>) δ: 2.6 (<u>C</u>H<sub>2</sub>I), 33.0 (CH2CH2I), 67.7 (CH2CH2CH2I), 68.3 (C-6), 70.7 (C-5), 73.3 (CH2), 73.4 (CH2), 75.1 (CH2), 75.8 (CH2), 77.4 (C-4), 79.7 (C-2), 82.0 (C-3), 96.3 (C-1), 115.3 (Ph), 118.1 (Ph), 127.6-128.5 (Ph), 137.8-138.8 (Ph), 150.9 (Ph), 154.0 (Ph); HR-MS (ESI) m/z: 823.2098 (Calcd for C43H45O7I: M<sup>+</sup>+Na, 823.2102).

Heptaxis-(2,3-di-O-benzyl)-6<sup>B,C,E,F,G</sup>-penta-O-benzyl-6<sup>A</sup>-O-(3-O-[4-O-{2,3,4,6-tetra-O-benzyl-a-d-glucopyranoside-1-yl}phenyl]propane-1yl)β-cyclodextrin or Heptaxis-(2,3-di-O-benzyl)-6<sup>B,C,E,E,G</sup>-penta-O-benzyl-6<sup>D</sup>-O-(3-O-[4-O-{2,3,4,6-tetra-O-benzyl-a-dependence}-dependence] yl}phenyl]propane-1-yl)β-cyclodextrin (13) To a solution of 11 (84 mg, 0.1 mmol), 12 (75 mg, 0.026 mmol) in DMF (4 ml) were added KOH (179 mg, 3.19 mmol) and n-Bu<sub>4</sub>NI (4.9 mg, 0.013 mmol). After the reaction mixture was stirred for 64 h, the reaction was then quenched by adding water (10 ml). The mixture was extracted with EtOAc (three times) and the combined organic solvent was dried over anhydrous Na2SO4. The organic solvent was filtered and evaporated under reduced pressure. The crude product was purified by preparative silica-gel TLC (hexane/ethyl acetate=3/1) to afford 13 (64 mg, 69%) as a colorless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.87 (2H, m, CH2CH2OPh), 3.36-5.39 (106H, m, CyD, H-1, H-2, H-3, H-4, H-5, H-6, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>OPh, CH<sub>2</sub>Ph), 6.73 (2H, dd, J=3.4 Hz, J=8.9 Hz, Ph), 6.96 (2H, dd, J=1.4 Hz, J=8.9 Hz, Ph), 7.05-7.38 (115H, m, Ph); <sup>13</sup>C-NMR  $(CDCl_3)$   $\delta$ : 14.2, 21.0, 29.5, 60.3—82.0, 96.3, 98.3—98.7, 115.1, 118.1, 126.9—128.4, 137.8—139.3, 150.7, 154.2; MALDI-TOF-MS m/z: 3540.6 (Calcd for C<sub>218</sub>H<sub>228</sub>O<sub>42</sub>: M<sup>+</sup>+Na, 3540.6).

**6<sup>A</sup>-O-(3-O-[4-O-{α-D-Glucopyranoside-1-yl}phenyl]propane-1-yl)β**cyclodextrin (5) To a solution of **13** (56 mg, 0.015 mmol) in DMF (5 ml) was added palladium hydroxide (76 mg, 0.49 mmol). Hydrogen was bubbled through the solution for 6 h. After the solvent was filtered and evaporated under reduced pressure, the crude product was isolated by adsorption on LH 20 followed by eluting with methanol to afford **5** (22 mg, 98%) as white crystals. mp: 284.1—286.0 °C;  $[\alpha]_D^{23} + 97^\circ$  (c=1.0, CH<sub>3</sub>OH); <sup>1</sup>H-NMR (D<sub>2</sub>O)  $\delta$ : 1.88 (2H, m, CH<sub>2</sub>CH<sub>2</sub>OPh), 3.43—4.08 (52H, m, CyD-2, CyD-3, CyD-4, CyD-5, CyD-6, H-2, H-3, H-4, H-5, H-6, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>OPh), 4.90—5.06 (7H, m, CyD-1), 5.56 (1H, d, J=2.1 Hz, H-1), 6.82 (2H, d, J=8.2 Hz, Ph), 7.12 (2H, d, J=8.9 Hz, Ph); <sup>3</sup>C-NMR (D<sub>2</sub>O)  $\delta$ : 29.4, 60.2—74.5, 81.2—82.9, 98.2, 101.4—102.7, 115.9—118.3, 151.3, 154.7; MALDI-TOF-MS *m/z*: 1469.2 (Calcd for C<sub>57</sub>H<sub>90</sub>O<sub>42</sub>: M<sup>+</sup>+Na, 1469.5).

**4-Allyloxyphenyl 2-Acetamido-3,4,6-tri-O-benzyl-2-deoxy-\alpha-D-glucopyranoside (15)** Yb(OTf)<sub>3</sub> (635 mg, 1 mmol) was added to a solution of **14** (653 mg, 1.2 mmol), 4-allyloxyphenol (153 mg, 1 mmol) and BF<sub>3</sub>·OEt<sub>2</sub> (3.9  $\mu$ l, 0.03 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (8 ml) at 0 °C. The resulting mixture was stirred for 24 h at room temperature. The reaction was then quenched by the addition of a sat. NaHCO<sub>3</sub> solution (5 ml). The reaction mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub>, and the organic layer was washed with water and a sat. NaCl solution. After the organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, the solvent was evaporated under reduced pressure. The crude product was purified by preparative silica-gel TLC (ethyl acetate/hexane=2/1) to give 15 (389 mg, 61%) as white crystals. mp: 195.0—197.0 °C;  $[\alpha]_D^{23} + 159^\circ$  (*c*=1.0, CHCl<sub>3</sub>); <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.58 (3H, s, CH<sub>3</sub>), 3.63 (1H, dd, J=2.0 Hz, J=11.0 Hz, H<sub>a</sub>-6), 3.76 (1H, dd, J=3.4 Hz, J=11.0 Hz, H<sub>b</sub>-6), 3.34—3.90 (2H, m, H-3, H-4), 3.92-3.94 (1H, m, H-5), 4.38 (1H, dt, J=2.7 Hz, J=8.9 Hz, H-2), 4.45—4.47 (3H, m,  $CH_2CH=CH_2$ ,  $CH_aH_bPh$ ), 4.57 (1H, d, J=10.3 Hz, CH<sub>2</sub>Ph), 4.61 (1H, d, J=11.7 Hz, CH<sub>2</sub>Ph), 4.70 (1H, d, J=12.3 Hz, CH<sub>2</sub>Ph), 4.83 (1H, d, J=10.4 Hz, CH<sub>2</sub>Ph), 4.90 (1H, d, J=11.7 Hz, CH<sub>2</sub>Ph), 5.27-5.31 (2H, m,  $CH=C\underline{H}_{a}H_{b}$ , NH), 5.34 (1H, dd, J=2.1 Hz, J=17.9 Hz, CH=CH<sub>a</sub><u>H</u><sub>b</sub>), 5.41 (1H, d, J=3.4 Hz, H-1), 6.03 (1H, m, C<u>H</u>=CH<sub>2</sub>), 6.81 (2H, d, J=9.0 Hz, Ph), 6.90 (2H, d, J=8.9 Hz, Ph), 7.20-7.37 (15H, m, Ph); <sup>13</sup>C-NMR (CDCl<sub>3</sub>)  $\delta$ : 23.4 (CH<sub>3</sub>), 52.5 (C-2), 68.4 (C-6), 69.3 (<u>CH</u><sub>2</sub>CH=CH<sub>2</sub>), 71.6 (C-5), 73.4 (<u>CH</u><sub>2</sub>Ph), 74.8 (<u>C</u>H<sub>2</sub>Ph), 75.1 (<u>C</u>H<sub>2</sub>Ph), 78.3 (C-4), 79.9 (C-3), 92.2 (C-1), 115.6 (Ph), 117.6 (CH=CH<sub>2</sub>), 117.9 (Ph), 127.6—128.7 (Ph), 133.3 (CH=CH<sub>2</sub>), 134.1—166.8 (Ph), 169.8 (C=O); HR-MS (ESI) m/z: 646.2787 (Calcd for  $C_{38}H_{41}NO_7$ : M<sup>+</sup>+Na, 646.2775).

4-O-(Carboxymethyl)phenyl 2-Acetamido-3,4,6-tri-O-benzyl-2-deoxy- $\alpha$ -**p-glucopyranoside** (16) Ozone was bubbled through a stirred solution of 15 (389 mg, 0.62 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (15 ml) at -78 °C for 2 h. After triphenylphosphine (497 mg, 1.9 mmol) was added at -78 °C and the reaction temperature was raised to room temperature, the reaction mixture was stirred for 2 h. The solvent was then evaporated under reduced pressure. To a solution of the crude product in t-butylalcohol (10 ml)-H<sub>2</sub>O (2 ml) was added NaClO<sub>2</sub> (561 mg, 6.2 mmol), NaH<sub>2</sub>PO<sub>4</sub> (116 mg, 0.74 mmol) and 2methyl-2-butene (305  $\mu$ l, 2.9 mmol). After the reaction mixture was stirred for 15 h, the reaction was quenched by adding 2 M HCl (1 ml) and water (5 ml). After the reaction mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> (three times), the combined organic solvent was dried over anhydrous Na2SO4. The organic solvent was filtered and evaporated under reduced pressure. The crude product was purified by preparative silica-gel TLC (chloroform/methanol= 5/1) to afford 16 (281 mg, 70%) as white crystals. mp: 165.0-165.9 °C;  $[\alpha]_{D}^{23}$  +79° (c=1.0, CHCl<sub>3</sub>); <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.90 (3H, s, CH<sub>3</sub>), 3.63 (1H, d, J=10.9 Hz, H<sub>a</sub>-6), 3.77 (1H, dd, J=4.2 Hz, J=11.0 Hz, H<sub>b</sub>-6), 3.83 (3H, m, H-4, CH<sub>2</sub>COOH), 3.94-3.98 (2H, m, H-3, H-5), 4.37 (1H, s, NH), 4.38 (1H, dt, J=3.4 Hz, J=11.0 Hz, H-2), 4.46 (1H, d, J=12.4 Hz, CH<sub>2</sub>Ph), 4.54 (1H, d, J=10.3 Hz, CH<sub>2</sub>Ph), 4.61 (1H, d, J=12.4 Hz, CH<sub>2</sub>Ph), 4.76 (1H, d, J=11.0 Hz, CH<sub>2</sub>Ph), 4.81 (1H, d, J=11.0 Hz, CH<sub>2</sub>Ph), 4.89 (1H, d, J=11.0 Hz, CH<sub>2</sub>Ph), 5.36 (1H, d, J=4.2 Hz, H-1), 6.96 (2H, d, J=9.0 Hz, Ph), 7.18 (2H, d, J=7.6 Hz, Ph), 7.29–7.39 (15H, m, Ph); <sup>13</sup>C-NMR (CDCl<sub>3</sub>) *δ*: 22.5 (CH<sub>3</sub>), 52.6 (C-2), 67.4 (C-6), 71.3 (C-5), 73.2 (<u>C</u>H<sub>2</sub>Ph), 74.8 (CH<sub>2</sub>Ph), 74.9 (CH<sub>2</sub>Ph), 78.0 (CH<sub>2</sub>COOH), 78.1 (C-4), 79.9 (C-3), 97.2 (C-1), 115.9 (Ph), 117.9 (Ph), 127.5-128.3 (Ph), 133.0-138.2 (Ph), 150.9 (Ph), 180.0 (C=O); HR-MS (ESI) m/z: 664.2508 (Calcd for C<sub>37</sub>H<sub>30</sub>NO<sub>0</sub>: M<sup>+</sup>+Na, 664.2517).

4-O-(2-Acetamido-2-deoxy-a-D-glucopyranoside-1-yl)phenyl-N-(6<sup>A</sup>deoxy- $\beta$ -cyclodextrin- $6^{A}$ -yl)acetamide (6) To a solution of 16 (41 mg, 0.062 mmol) in DMF (1.5 ml) were added dimethylphosphinothioyl chloride (13 mg, 0.097 mmol) and DIEA (21 µl, 0.12 mmol). After the reaction mixture was stirred for 40 min, 17 (84 mg, 0.074 mmol) in DMF (1.5 ml) was added. After the reaction mixture was stirred for 24 h, the solvent was evaporated under reduced pressure. The resulting reaction mixture was washed with diethyl ether (eight times) and dissolved in DMF (3 ml). Palladium hydroxide (29 mg, 0.18 mmol) was added to the solution and hydrogen was bubbled through it for 24 h. After the solvent was filtered and evaporated under reduced pressure, the crude product was isolated by adsorption on HP 20 (DIAION) followed by eluting with methanol to afford 6 (52 mg, 56%) as white crystals. mp: 281.0–283.0 °C; <sup>1</sup>H-NMR (D<sub>2</sub>O) δ: 1.92 (3H, s, CH<sub>2</sub>), 3.21-3.93 (51H, m, CyD, H-2, H-3, H-4, H-5, H-6, CH<sub>2</sub>C(O)NH), 4.65-4.96 (7H, m, CyD), 5.59 (1H, d,J=3.4 Hz, H-1), 6.63 (2H, d, J=8.9 Hz, Ph), 7.01 (2H, d, J=9.0 Hz, Ph); MALDI-TOF-MS m/z: 1510.1 (Calcd for C<sub>58</sub>H<sub>90</sub>N<sub>2</sub>O<sub>42</sub>: M<sup>+</sup>+Na, 1509.5).

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