## A New Amide, Piperchabamide F, and Two New Phenylpropanoid Glycosides, Piperchabaosides A and B, from the Fruit of *Piper chaba*

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A new amide, piperchabamide F (1), and two new phenylpropanoid glycosides, piperchabaosides A (2) and B (3), were isolated from 80% aqueous acetone extract from fruit of *Piper chaba*. Their stereostructures were elucidated on the basis of chemical and physicochemical evidence.

Key words Piper chaba; piperchabamide; piperchabaoside; Piperaceae; Thai natural medicine

During the course of our studies on Thai natural medicines.<sup>1-13</sup> we found that the 80% aqueous acetone extract from the fruit of Piper chaba HUNTER (syn. P. retrofractum VAHL., Piperaceae) was found to show protective effects on ethanol- or indomethacin-induced gastric lesions in rats,<sup>1)</sup> inhibitory effects on the increase in serum aspartate aminotransaminase (sAST) and alanine aminotransaminase (sALT) levels induced by D-galactosamine (D-GalN)/lipopolysaccharide (LPS)-induced liver injury in mice and on cell death induced by D-GalN/tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) in primary cultured mouse hepatocytes,<sup>6,12)</sup> and promoting effect on adipogenesis of 3T3-L1 cells.<sup>13)</sup> As a continuing study on the constituents from the fruit of *P. chaba*, we additionally isolated a new amide constituent named piperchabamide F (1) and two new phenylpropanoid glycosides, piperchabaosides A (2) and B (3), together with three known sesquiterpenes (4-6) and a known phenylpropanoid glycoside (7). This paper deals with the isolation and stereostructure elucidation of three new compounds (1-3) from the fruit of P. chaba.

The 80% aqueous acetone extract from the fruit of *P. chaba* (19.7%, purchased in Thailand) was partitioned between EtOAc–H<sub>2</sub>O (1:1, v/v) to give EtOAc-soluble fraction (9.7%) and an aqueous phase.<sup>1)</sup> The aqueous phase was further extracted with *n*-BuOH to give *n*-BuOH-soluble fraction (2.1%) and H<sub>2</sub>O-soluble fraction (7.0%). The EtOAc-soluble fraction was subjected to normal- and reversed-phase silica gel column chromatographies and finally HPLC to furnish **1** (0.0007%) together with three sesquiterpenes, 1-hydroxybisabola-2,10-dien-4-one<sup>14)</sup> (**4**, 0.0006%), 1,4-dihydroxybisabola2,10-diene<sup>15)</sup> (5, 0.0007%), and 3,4-dihydroxybisabola-1,10-diene<sup>15-17)</sup> (6, 0.0013%). From the *n*-BuOH-soluble fraction, 2 (0.012%) and 3 (0.0019%) were purified together with a phenylpropanoid glycoside,  $rosin^{18}$  (7, 0.0007%) using normal- and reversed-phase silica gel chromatographies and finally HPLC.

Structure of Piperchabamide F (1) Piperchabamide F (1) was obtained as colorless oil with positive optical rotation  $(\alpha_{\rm D}^{27} + 7.1^{\circ} \text{ in CHCl}_3)$ . The IR spectrum of 1 showed absorption bands at 2924, 1626, 1561, 1509, 1491, and 1258  $cm^{-1}$  ascribable to methylene, unsaturated amide group, and aromatic ring. The electron ionization (EI)-MS of 1 showed a molecular ion peak at m/z 343 (M<sup>+</sup>), and the molecular formula was determined as C21H29NO3 by high-resolution EI-MS measurement. The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of 1 (CDCl<sub>3</sub>, Table 1) showed signals assignable to two methyls  $[\delta 0.91 (3H, dd, J=6.6, 7.4 Hz, 4'-H_2), 0.92 (3H, d, J=$ 6.6 Hz, 5'-H<sub>3</sub>)], six methylenes [ $\delta$  1.16, 1.40 (1H each, both m, 3'-H<sub>2</sub>), 1.49, 2.19 (4H each, both m, 5, 6, 4, 7-H<sub>2</sub>), 3.13, 3.27 (1H each, both m, 1'-H<sub>2</sub>)], a methine [ $\delta$  1.59 (1H, m, 2'-H)], a 3,4-methylenedioxyphenyl group [ $\delta$  5.92 (2H, br s, 12-H<sub>2</sub>), 6.72 (2H, br s, 14, 15-H), 6.88 (1H, br s, 11-H)], two trans-olefinic proton pairs [ $\delta$  5.75 (1H, dd, J=1.2, 15.4 Hz, 2-H), 6.00 (1H, dt, J=16.5, 6.7 Hz, 8-H), 6.29 (1H, d, J= 16.5 Hz, 9-H), 6.81 (1H, dt, J=15.4, 7.1 Hz, 3-H)], and a conjugated amide group [ $\delta_{\rm C}$  165.8 (C-1)]. The planar structure of 1 was constructed on the basis of various NMR experiments.<sup>19)</sup> Namely, the <sup>1</sup>H-<sup>1</sup>H correlation spectroscopy (COSY) experiments on 1 indicated the presence of three partials written in bold lines, while in the heteronuclear multiple



bond connectivity (HMBC) experiments, long range correlations were observed between the following proton and carbon pairs: 2-H and 1-C; 8, 9-H and 10-C; 1'-H<sub>2</sub> and 1-C (Fig. 1). Finally, acid hydrolysis of **1** with  $6 \le HCl^{12,20}$  liberated (*R*)-2-methylbutylamine,<sup>21</sup> which was identified by HPLC analysis using refractive index and optical rotation detectors. On the basis of above-mentioned evidence, the stereostructure of **1** was elucidated as shown.

Structures of Piperchabaosides A (2) and B (3) Piperchabaoside A (2) was isolated as a white powder with negative optical rotation ( $[\alpha]_D^{23} - 26.5^\circ$  in MeOH). In the positive-ion fast atom bombardment (FAB)-MS of 2, the quasimolecular ion peaks were observed at m/z 481  $(M+Na)^+$  and m/z 457  $(M-H)^-$ , and the molecular formula C<sub>21</sub>H<sub>30</sub>O<sub>11</sub> was determined by high resolution FAB-MS measurement. The IR spectrum of 2 showed absorption bands at 3426, 1089, and 1030 cm<sup>-1</sup> ascribable to hydroxyl and ether functions. Acid hydrolysis of 2 with 1.0 M HCl liberated D-glucose, which was identified by HPLC analysis.<sup>22-26)</sup> The <sup>1</sup>H- and <sup>13</sup>C-NMR (pyridine-d<sub>5</sub>, Table 2) spectra of 2 showed signals assignable to a methylene bearing an oxygen function [ $\delta$  4.43, 4.68 (1H each, both dd, J=5.8, 13.1 Hz, 9-H<sub>2</sub>)], a *trans*-olefin pair [ $\delta$  6.45 (1H, dt, J=16.2, 5.8 Hz, 8-H), 6.76 (1H, d, J=16.2 Hz, 7-H)], and a monosubstituted benzene ring [ $\delta$  7.22 (1H, t, J=7.9 Hz, 4-H), 7.30 (2H, dd, J=7.3, 7.9 Hz, 3,5-H), 7.39 (2H, d, J=7.3 Hz, 2,6-H)] together with two  $\beta$ -D-glucopyranosyl parts [ $\delta$  4.88 (1H, d, J=7.6 Hz, 1'-H), 5.16 (1H, d, J=7.9 Hz, 1"-H)], which were superimposable on those of rosin (7), except for the signals due to an additional  $\beta$ -D-glucopyranosyl unit. Finally, the connectivities of the glycosyl linkages in 2 were elucidated on the basis of HMBC experiment, which showed long-range correlations between the following proton and carbon pairs as shown in Fig. 1: the inner-Glc-1-proton (1'-H) and the 9-carbon ( $\delta_{\rm C}$  69.8), and the *terminal*-Glc-1-proton (1"-H) and the *inner*-Glc-4-carbon ( $\delta_{\rm C}$  81.2, 4'-C). Consequently, piperchabaoside A was determined to be trans-cinnamyl alcohol  $O-\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 4)- $\beta$ -D-glucopyranoside (2).

Piperchabaoside B (3) was also isolated as a white powder with negative optical rotation ( $[\alpha]_{D}^{26}$  – 18.0°, MeOH). Its molecular formula C<sub>27</sub>H<sub>38</sub>O<sub>15</sub> was determined from the positiveand negative-ion FAB-MS and by high resolution FAB-MS measurements. The IR spectrum of 3 showed absorption bands at 3401, 1736, 1719, 1075, and 1040  $\text{cm}^{-1}$  assignable to hydroxyl, ester carbonyl, carboxyl, and ether functions. Treatment of 3 with 1.0% sodium methoxide (NaOMe)-MeOH provided 2 together with (S)-3-hydroxy-3-methylglutaric acid monomethyl ester<sup>27)</sup> (3a). The proton and carbon signals in the <sup>1</sup>H- and <sup>13</sup>C-NMR (pyridine- $d_5$ , Table 2) spectra of 3 were similar to those of 2, except for the signals due to an acyl part [ $\delta$  1.74 (3H, s, 6<sup>'''</sup>-H<sub>3</sub>), 3.13 (4H, m, 2<sup>'''</sup>,4<sup>'''</sup>-H<sub>2</sub>)]. The connectivities of the acyl moiety and glycosyl linkages in 3 were elucidated on the basis of HMBC experiment, which showed long-range correlations were observed between following proton and carbon pairs as shown in Fig. 1: the *inner*-Glc-1-proton [ $\delta$  4.86 (1H, d, J=7.6 Hz, 1'-H)] and the 9-carbon ( $\delta_{\rm C}$  69.9), the *terminal*-Glc-1-proton [ $\delta$  5.10 (1H, d, J=7.9 Hz, 1"-H)] and the inner-Glc-4-carbon ( $\delta_{\rm C}$ 81.6, 4'-C), and the *inner*-Glc-6-protons [ $\delta$  4.49 (1H, dd, J= 6.1, 12.8 Hz), 5.15 (1H, br d, J=ca. 13 Hz), 6'-H<sub>2</sub>] and the



Fig. 1.  ${}^{1}H{}^{-1}H$  COSY and HMBC Correlations of 1—3

Table 1. <sup>1</sup>H- and <sup>13</sup>C-NMR Data of Piperchabamide F (1)

Position	1		
	$\delta_{_{ m H}}(J{ m Hz})$	$\delta_{ m c}$	
1		165.8	
2	5.75 (dd, 1.2, 15.4)	123.5	
3	6.81 (dt, 15.4, 7.1)	144.2	
4	2.19 (2H, m)	31.8	
5	1.49 (2H, m)	$27.7^{a}$	
6	1.49 (2H, m)	$28.9^{a)}$	
7	2.19 (2H, m)	32.6	
8	6.00 (dt, 16.5, 6.7)	128.6	
9	6.29 (d, 16.5)	129.4	
10		132.1	
11	6.88 (br s)	105.2	
12		147.7	
13		146.3	
14	6.72 (br s)	108.0	
15	6.72 (br s)	120.0	
16	5.92 (2H, brs)	100.8	
1'	3.13, 3.27 (both m)	45.0	
2'	1.59 (m)	34.9	
3'	1.16, 1.40 (both m)	27.0	
4'	0.91 (3H, dd, 6.6, 7.4)	17.2	
5'	0.92 (3H, d, 6.6)	11.3	

Measured in CDCl<sub>3</sub>. a) May be interchangeable.

acyl ester carbonyl carbon ( $\delta_{\rm C}$  171.6, 1‴-C). Thus, the structure of piperchabaoside B was constructed as *trans*-cinnamyl alcohol *O*- $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 4)-6-*O*-(*S*)-3-hydroxy-3-methylglutaroyl- $\beta$ -D-glucopyranoside (**3**).

## Experimental

The following instruments were used to obtain physical data: specific rotations, Horiba SEPA-300 digital polarimeter (l=5 cm); UV spectra, Shimazdu UV-1600 spectrometer; IR spectra, Shimadzu FTIR-8100 spectrometer; <sup>1</sup>H-NMR spectra, JEOL JNM-LA500 (500 MHz) and EX-270 (270 MHz) spectrometers; <sup>13</sup>C-NMR spectra, JEOL JNM-LA500 (125 MHz) and EX-270 (68 MHz) spectrometers with tetramethylsilane as an internal standard; EI-MS and high-resolution EI-MS, JEOL JMS-GCMATE mass spectrometer; FAB-MS and high resolution FAB-MS, JEOL JMS-SX 102A mass spectrometer; HPLC detector, Shimadzu RID-10A refractive index, Shimadzu SPD-10A UV–VIS, and Shodex OR-2 optical rotation detectors. HPLC column, Cosmosil 5C<sub>18</sub>-MS-II (Nacalai Tesque Inc., 250×4.6 mm

Position –	2		3	
	$\delta_{ m H}(J{ m Hz})$	$\delta_{ m c}$	$\delta_{ m H}$ (J Hz)	$\delta_{ m c}$
1		137.4		137.4
2,6	7.39 (2H, d, 7.3)	126.9	7.38 (2H, d, 7.9)	126.9
3,5	7.30 (2H, dd, 7.3, 7.9)	129.0	7.28 (2H, dd, 7.3, 7.9)	128.9
4	7.22 (t, 7.9)	127.9	7.21 (t, 7.3)	127.9
7	6.76 (d, 16.2)	132.2	6.77 (d, 16.2)	132.4
8	6.45 (dt, 16.2, 5.8)	126.8	6.44 (dt, 16.2, 6.1)	126.9
9	4.43, 4.68 (both dd, 5.8, 13.1)	69.8	4.45, 4.72 (both dd, 6.1, 12.8)	69.9
1'	4.88 (d, 7.6)	103.6	4.86 (d, 7.6)	103.5
2'	4.09 (m)	74.8	4.09 (m)	74.5
3'	4.24 (m)	76.8	4.21 (m)	76.6
4'	4.31 (m)	81.2	4.18 (m)	81.6
5'	3.87 (m)	76.5	4.03 (m)	73.5
6'	4.44, 4.52 (both m)	62.1	4.49 (dd. 6.1, 12.8)	64.1
			5.15 (br d. ca. 13)	
1″	5.16 (d. 7.9)	104.9	5.10 (d. 7.9)	105.2
2″	4.06 (m)	74.8	4.03 (m)	74.9
3"	3.99 (m)	78.4	4.18 (m)	78.4
4″	4.15 (dd, 8.3, 8.5)	71.6	4.09 (m)	71.9
5″	4.19 (m)	78.2	4.08 (m)	78.5
6"	4.29, 4.49 (both m)	62.5	4.24 (m)	62.8
			4.54 (br d. ca. 12)	
1‴				171.6
2‴			3.13 (2H, m)	46.8
3‴				70.1
4‴			3 13 (2H m)	46.5
5‴			5.15 (211, 111)	174.9
6‴			1.74 (3H, s)	28.3

Table 2. <sup>1</sup>H- and <sup>13</sup>C-NMR Data of Piperchabaosides A (2) and B (3)

Measured in pyridine- $d_5$ .

i.d.) and (250×20 mm i.d.) columns were used for analytical and preparative purposes, respectively.

The following experimental conditions were used for chromatography: normal-phase silica gel column chromatography (CC), silica gel 60N (Kanto Chemical Co., Ltd., 63—210 mesh, spherical, neutral); reversed-phase silica gel CC, Chromatorex ODS DM1020T (Fuji Silysia Chemical, Ltd., 100—200 mesh); normal-phase TLC, pre-coated TLC plates with silica gel 60F<sub>254</sub> (Merck, 0.25 mm); reversed-phase TLC, pre-coated TLC plates with silica gel RP-18 F<sub>254S</sub> (Merck, 0.25 mm), detection was achieved by spraying with 1% Ce(SO<sub>4</sub>)<sub>2</sub>–10% aqueous H<sub>2</sub>SO<sub>4</sub>, followed by heating.

Plant Material This item was described in a previous report.<sup>1)</sup>

Extraction and Isolation The dried fruit of P. chaba (4.0 kg) was finely cut and extracted four times with 80% (v/v) aqueous acetone at room temperature for 1 d. Evaporation of the solvent under reduced pressure provided an 80% aqueous acetone extract (788 g, 19.7% from the dried fruit). The 80% aqueous acetone extract (463 g) was partitioned in an EtOAc-H<sub>2</sub>O (1: 1, v/v) mixture. The aqueous phase was further extracted with *n*-BuOH. Removal of the solvent under reduced pressure yielded EtOAc-, n-BuOH-, and H<sub>2</sub>O-soluble fractions [227 g (9.7%), 50 g (2.1%), and 164 g (7.0%)], respectively. Fractions 4-2 (0.21 g), 5-2 (0.62 g), and 6-2 (0.07 g) were obtained from the EtOAc-soluble fraction (118 g) as described previously.<sup>1,12</sup>) The fraction 4-2 (0.21 g) was subjected to preparative HPLC [CH3CN-H2O (50:50, v/v)] to give 1-hydroxybisabola-2,10-dien-4-one (4, 7 mg, 0.0006%) together with piperchabamide A (27 mg) and N-isobutyl-(2E,4E)-deca-2,4dienamide (27 mg). The fraction 5-2 (0.62 g) was subjected to HPLC [CH<sub>3</sub>CN-H<sub>2</sub>O (70:30, v/v)] to give piperchabamide F (1, 9 mg, 0.0007%) together with piperchabamide D (20 mg), pipercide (=retrofractamide B, 100 mg), and guineensine (292 mg). The fraction 6-2 (0.07 g) was further purified by HPLC [CH3CN-H2O (45:55, v/v)] to give 1,4-dihydroxybisabola-2,10-diene (5, 9 mg, 0.0007%) and 3,4-dihydroxybisabola-1,10-diene (6, 16 mg, 0.0013%).

The *n*-BuOH-soluble fraction (40.5 g) was subjected to normal-phase silica gel CC [1.5 kg, CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O ( $30:3:1 \rightarrow 15:3:1 \rightarrow 10:3:1$ , lower layer $\rightarrow 6:4:1$ , v/v/v) $\rightarrow$ MeOH] to give six fractions [Fr. 1 (3.3 g), Fr. 2 (2.4 g), Fr. 3 (4.3 g), Fr. 4 (1.7 g), Fr. 5 (0.8 g), Fr. 6 (25.7 g)]. The fraction 2

(2.4 g) was subjected to reversed-phase silica gel CC [120 g, MeOH–H<sub>2</sub>O (20:80–40:60–80:20, v/v)–MeOH] to give three fractions [Fr. 2-1 (2.05 g), 2-2 (0.20 g), 2-3 (0.07 g)]. The fraction 2-2 (0.20 g) was further purified by HPLC [MeOH–H<sub>2</sub>O (20:80, v/v)] to give rosin (7, 13 mg, 0.0007%). The fraction 4 (1.7 g) was subjected to reversed-phase silica gel CC [50 g, MeOH–H<sub>2</sub>O (20:80–40:60, v/v)–MeOH] to give three fractions [Fr. 4-1 (0.67 g), 4-2 (0.57 g), 4-3 (0.17 g)]. The fraction 4-2 (0.57 g) was further purified by HPLC [MeOH–H<sub>2</sub>O (10:90, v/v)] to give piperchabasosides A (**2**, 54 mg, 0.0028%) and B (**3**, 36 mg, 0.0019%). The fraction 5 (0.8 g) was purified by HPLC [MeOH–H<sub>2</sub>O (10:90, v/v)] to give **2** (177 mg, 0.0029%).

Piperchabamide F (1): Colorless oil,  $[α]_D^{25} + 7.1^\circ$  (*c*=0.27, CHCl<sub>3</sub>). Highresolution EI-MS: Calcd for C<sub>21</sub>H<sub>29</sub>NO<sub>3</sub> (M<sup>+</sup>) 343.2147; Found 343.2152. UV [ $λ_{max}$  (log ε), EtOH]: 260 (4.1) nm. IR (KBr, cm<sup>-1</sup>): 2924, 1626, 1561, 1509, 1491, 1258, 1043, 965, 924. <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$ : given in Table 1. <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)  $\delta_C$ : given in Table 1. EI-MS (%): *m/z* 343 (M<sup>+</sup>, 41), 135 (100).

Piperchabaoside A (2): A white powder,  $[\alpha]_D^{23} - 26.5^\circ$  (c=2.40, MeOH). High-resolution positive-ion FAB-MS: Calcd for  $C_{21}H_{30}O_{11}Na$  (M+Na)<sup>+</sup> 481.1686; Found 481.1678. UV  $[\lambda_{max} (\log \varepsilon), MeOH]$ : 251 (4.5) nm. IR (KBr, cm<sup>-1</sup>): 3426, 1089, 1030. <sup>1</sup>H-NMR (500 MHz, pyridine- $d_5$ )  $\delta$ : given in Table 2. <sup>13</sup>C-NMR (125 MHz, pyridine- $d_5$ )  $\delta_C$ : given in Table 2. Positive-ion FAB-MS m/z: 481 (M+Na)<sup>+</sup>. Negative-ion FAB-MS m/z: 457 (M-H)<sup>-</sup>.

Piperchabaoside B (3): A white powder,  $[\alpha]_D^{26} - 18.0^\circ$  (c=3.21, MeOH). High-resolution positive-ion FAB-MS: Calcd for  $C_{27}H_{38}O_{15}Na$  (M+Na)<sup>+</sup> 625.2108; Found 625.2115. UV [ $\lambda_{max}$  (log  $\varepsilon$ ), MeOH]: 251 (4.3) nm. IR (KBr, cm<sup>-1</sup>): 3401, 1736, 1719, 1075, 1040. <sup>1</sup>H-NMR (500 MHz, pyridine- $d_5$ )  $\delta$ : given in Table 2. <sup>13</sup>C-NMR (125 MHz, pyridine- $d_5$ )  $\delta_C$ : given in Table 2. Positive-ion FAB-MS m/z: 625 (M+Na)<sup>+</sup>. Negative-ion FAB-MS m/z: 601 (M-H)<sup>-</sup>.

Acid Hydrolysis of Piperchabamide F (1) A solution of 1 (5 mg) in 6 M HCl (0.5 ml) was heated at 110 °C for over night in a sealed tube. After cooling, the reaction mixture was basified (pH 10) with aq. NaOH and then the mixture was extracted with *n*-hexane. The *n*-hexane phase was evaporated *in vacuo* gave a residue, which was subjected to HPLC analysis [column: Cosmosil 5C<sub>18</sub>-MS-II, 250×4.6 mm i.d.; mobile phase: MeOH–H<sub>2</sub>O (50: 50, v/v); detection: RI and OR; flow rate: 1.0 ml/min] to identify (*R*)-2-

methylbutylamine [ $t_R$  9.1 min (positive)].

Acid Hydrolysis of Piperchabaoside A (2) A solution of 2 (3.0 mg) in 1 M HCl (1.0 ml) was heated at 80 °C for 3 h. After cooling, the reaction mixture was neutralized with Amberlite IRA-400 (OH<sup>-</sup> form) and then the resin was removed by filtration. Removal of the solvent from the filtrate under reduced pressure, the residue was separated by Sep-Pak C18 cartridge column (H<sub>2</sub>O→MeOH). The H<sub>2</sub>O-eluted fraction was subjected to HPLC analysis under the following conditions: HPLC column, Kaseisorb LC NH<sub>2</sub>-60-5, 250×4.6 mm i.d. (Tokyo Kasei Co., Ltd., Tokyo, Japan); detection, optical rotation [Shodex OR-2 (Showa Denko Co., Ltd., Tokyo, Japan); mobile phase, CH<sub>3</sub>CN-H<sub>2</sub>O (85 : 15, v/v); flow rate 0.8 ml/min]. Identification of Dglucose (i) present in the aqueous layer was carried out by comparison of their retention times and optical rotations with those of authentic samples.  $t_{\rm R}$ : (i) 13.9 min (positive optical rotation).

**Deacylation of Piperchabaoside B (3)** A solution of **3** (20.2 mg) in 1.0% sodium methoxide (NaOMe)–MeOH (3.0 ml) was stirred at room temperature for 3 h. The reaction mixture was neutralized with Dowex HCR-W2 (H<sup>+</sup> form) and the resin was removed by filtration. Evaporation of the solvent from the filtrate under reduced pressure gave a residue, which was purified by HPLC [Cosmosil 5C<sub>18</sub>-MS-II, MeOH–H<sub>2</sub>O (40:60, v/v)] to furnish **2** (12.0 mg) and (*S*)-3-hydroxy-3-methylglutaric acid monomethyl ester<sup>27</sup>) (**3a**, 1.6 mg).

Acknowledgements T. M. was supported by a Grant-in Aid for Scientific Research from 'High-tech Research Center' Project for Private Universities: matching fund subsidy from The Ministry of Education, Culture, Sports, Science and Technology (MEXT) of Japan, 2007—2011, a Grant-in Aid for Scientific Research from MEXT, and Takeda Science Foundation. M. Y. and H. M. were supported by the 21st COE Program, Academic Frontier Project, and a Grant-in Aid for Scientific Research from MEXT.

## **References and Notes**

- Morikawa T., Matsuda H., Yamaguchi I., Pongpiriyadacha Y., Yoshikawa M., *Planta Med.*, **70**, 152–159 (2004).
- Matsuda H., Morikawa T., Xu F., Ninomiya K., Yoshikawa M., *Planta Med.*, **70**, 1201–1209 (2004).
- Morikawa T., Ando S., Matsuda H., Kataoka S., Muraoka O., Yoshikawa M., Chem. Pharm. Bull., 53, 625–630 (2005).
- Morikawa T., Xu F., Matsuda H., Yoshikawa M., Chem. Pharm. Bull., 54, 1530—1534 (2006).
- Yoshikawa M., Xu F., Morikawa T., Pongpiriyadacha Y., Nakamura S., Asao Y., Kumahara A., Matsuda H., *Chem. Pharm. Bull.*, 55, 308– 316 (2007).
- Matsuda H., Ninomiya K., Morikawa T., Yasuda D., Yamaguchi I., Yoshikawa M., *Bioorg. Med. Chem. Lett.*, 18, 2038–2042 (2008).
- Yoshikawa M., Morikawa T., Funakoshi K., Ochi M., Pongpiriyadacha Y., Matsuda H., *Heterocycles*, 75, 1639–1650 (2008).

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- Morikawa T., Funakoshi K., Ninomiya K., Yasuda D., Miyagawa K., Matsuda H., Yoshikawa M., Chem. Pharm. Bull., 56, 956–962 (2008).
   Area Y. Marihama T. Yia Y. Okamata M. Hamas M. Matarda H.
- Asao Y., Morikawa T., Xie Y., Okamoto M., Hamao M., Matsuda H., Muraoka O., Yuan D., Yoshikawa M., *Chem. Pharm. Bull.*, **57**, 198– 203 (2009).
- Matsuda H., Asao Y., Nakamura S., Hamao M., Sugimoto S., Hongo M., Pongpiriyadacha Y., Yoshikawa M., *Chem. Pharm. Bull.*, 57, 487–494 (2009).
- Morikawa T., Xie Y., Asao Y., Okamoto M., Yamashita C., Muraoka O., Matsuda H., Pongpiriyadacha Y., Yuan D., Yoshikawa M., *Phyto-chemistry*, **70**, 1166—1172 (2009).
- Matsuda H., Ninomiya K., Morikawa T., Yasuda D., Yamaguchi I., Yoshikawa M., *Bioorg. Med. Chem.*, 17 (2009), in press.
- Zhang H., Matsuda H., Nakamura S., Yoshikawa M., *Bioorg. Med. Chem. Lett.*, 18, 3272–3277 (2008).
- Castro V., Tamayo-Castillo G., Jakupovic J., *Phytochemistry*, 28, 2415—2418 (1989).
- 15) Sy L.-K., Brown G. D., Phytochemistry, 45, 537-544 (1997).
- 16) Zdero C., Bohlmann F., Scott R., Phytochemistry, 26, 1999—2006 (1987).
- Ohshiro M., Kuroyanagi M., Ueno A., *Phytochemistry*, **29**, 2201– 2205 (1990).
- 18) Zapesochnaya G. G., Kurkin V. A., Chem. Nat. Compd., 18, 685–688 (1982).
- 19) The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of 1—3 were assigned with the aid of distortionless enhancement by polarization transfer (DEPT), homocorrelation spectroscopy (<sup>1</sup>H–<sup>1</sup>H COSY), heteronuclear multiple-quantum coherence (HMQC), and HMBC experiments.
- Matsumoto T., Nishimura K., Takeya K., Chem. Pharm. Bull., 50, 857–860 (2002).
- 21) Bazylak G., Analyst, 117, 1429-1433 (1992).
- 22) Morikawa T., Li X., Nishida E., Ito Y., Matsuda H., Nakamura S., Muraoka O., Yoshikawa M., J. Nat. Prod., 71, 828–835 (2008).
- Yoshikawa M., Li X., Nishida E., Nakamura S., Matsuda H., Muraoka O., Morikawa T., *Chem. Pharm. Bull.*, 56, 559–568 (2008).
- 24) Morikawa T., Wang L.-B., Nakamura S., Ninomiya K., Yokoyama E., Matsuda H., Muraoka O., Wu L.-J., Yoshikawa M., *Chem. Pharm. Bull.*, 57, 361–367 (2009).
- Wang L.-B., Morikawa T., Nakamura S., Ninomiya K., Matsuda H., Muraoka O., Wu L.-J., Yoshikawa M., *Heterocycles*, 78, 1235–1242 (2009).
- 26) Morikawa T., Wang L.-B., Ninomiya K., Nakamura S., Matsuda H., Muraoka O., Wu L.-J., Yoshikawa M., *Chem. Pharm. Bull.*, **57**, 853– 859 (2009).
- 27) Kawashima K., Mimaki Y., Sashida Y., Phytochemistry, 32, 1267– 1272 (1993).