## **Four New Ursane-Type Triterpenes, Olibanumols K, L, M, and N, from Traditional Egyptian Medicine Olibanum, the Gum-Resin of** *Boswellia carterii*

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**Four new ursane-type triterpenes, olibanumols K (1), L (2), M (3), and N (4), were isolated from traditional** Egyptian medicine olibanum, the exuded gum-resin from *Boswellia carterii* BIRDW. Their structures were eluci**dated on the basis of chemical and physicochemical evidence.**

**Key words** olibanumol; olibanum; *Boswellia carterii*; triterpene; Burseraceae

During the course of our studies on bioactive constituents from Egyptian and Yemeni natural medicines, $1-12$  we found that 80% aqueous acetone extract of exuded gum-resin of *Boswellia carterii*, so called olibanum (frankincense), showed anti-inflammatory effect by monitoring the inhibitory activity of nitric oxide (NO) production induced by lipopolysaccharide (LPS) in mouse peritoneal macrophages.<sup>10</sup>) From the extract, three monoterpenes, olibanumols A—C, a diterpene, olibanumol D, and six triterpenes, olibanumols E—J, were isolated together with five monoterpenes and two triterpenes, epilupeol  $(5)$  and isofouquierol  $(21)$ .<sup>10,13)</sup> Among them, several constituents including olibanumols A, D, E, and H exhibited the inhibitory effect on the NO produc- $\mu$ <sub>10</sub>) As a continuing study on this natural medicine, we further isolated four new ursane-type triterpenes, olibanumols K (**1**), L (**2**), M (**3**), and N (**4**), together with 19 known triterpenes. This paper deals with the isolation and structure elucidation of these newly isolated triterpenes (**1**—**4**).14)

The gum-resin from *B. carterii* collected in Yemen and purchased at Cairo of Egypt was extracted with 80% aqueous acetone at room temperature to give an aqueous acetone extract  $(78.9\%$  from the natural medicine), as was reported.<sup>10)</sup> By the intensive chromatographies on the aqueous acetone extract, **1** (0.0013%), **2** (0.043%), **3** (0.0043%), and **4** (0.0035%) have been newly isolated together with seven lupane-type triterpenes, epilupeol acetate<sup>15)</sup>  $(6, 2.87\%)$ , lup-20(30)-ene-3 $\alpha$ ,29-diol<sup>16)</sup> (7, 0.0050%), glochidiol<sup>17)</sup> (8, 0.0017%), lupeol<sup>18)</sup> (9, 0.72%), lup-20(29)-ene-2 $\alpha$ , 3 $\beta$ -diol<sup>19)</sup> (**10**, 0.0026%),  $3\beta$ -acetoxylup-20(29)-en-11 $\beta$ -ol<sup>20)</sup> (**11**, 0.043 %), and lupenone<sup>21,22)</sup> (12, 0.33%), five ursane-type triterpenes, urs-9(11),12-dien-3 $\beta$ -ol<sup>23,24)</sup> (13, 0.0012%), neoilexonol<sup>25)</sup> (**14**, 0.039%), neoilexonol acetate<sup>26,27)</sup> (**15**, 0.27%), urs-12-ene-3 $\beta$ ,11 $\alpha$ -diol<sup>28)</sup> (16, 0.0041%), and urs-12-ene- $3\alpha$ ,11 $\alpha$ -diol<sup>28)</sup> (17, 0.015%), and five dammarane-type and two nordammarane-type triterpenes, dammarenediol  $II^{29,30}$ (**18**, 0.089%), dammarenediol II acetate30) (**19**, 0.45%), 3-*O*acetyl-3b,20*S*,24-trihydroxydammar-25-ene31) (**20**, 0.0022%), isofouquierol acetate<sup>31,32)</sup> (22, 0.066%), ocotillol acetate<sup>33)</sup> (**23**, 0.0044%),  $3\beta$ -hydroxymansumbin-13(17)-en-16-one<sup>34)</sup>  $(24, 0.0065\%)$ , and mansumbinol<sup>35)</sup>  $(25, 0.0071\%)$ .

**Structure of Olibanumol K (1)** Olibanumol K (**1**) was obtained as a white powder with positive optical rotation  $([\alpha]_D^{27} + 16.2$  in MeOH). The IR spectrum of 1 showed absorption bands at  $1725 \text{ cm}^{-1}$  ascribable to an ester car-



Chart 1

bonyl function. In the positive-ion fast atom bombardment (FAB)-MS of **1**, a quasimolecular ion peak was observed at  $m/z$  477 (M+Na)<sup>+</sup>, and high-resolution positive-ion FAB-MS analysis revealed the molecular formula of **1** to be  $C_{31}H_{50}O_2$ . The <sup>1</sup>H- (CDCl<sub>3</sub>) and <sup>13</sup>C-NMR (Table 1) spectra of 1, which were assigned by various NMR experiments,<sup>36)</sup> showed signals assignable to eight methyls  $\delta$  0.80, 0.89, 0.92, 0.92, 1.11, 1.13 (3H each, all s, 28, 23, 24, 25, 27, 26- H<sub>3</sub>), 0.90 (3H, d, J=7.4 Hz, 29-H<sub>3</sub>), 0.92 (3H, d, J=7.0 Hz, 30-H<sub>3</sub>)], a methine bearing an oxygen function [ $\delta$  4.77 (1H, br s, 3-H)], an olefin  $\lceil \delta \rceil$  5.13 (1H, dd-like, 12-H)], and a formyl group  $\delta$  8.13 (1H, s, HCO–)] together with nine methylenes, five methines, and six quaternary carbons. The ursane-type triterpene structure of **1** was constructed on the basis of the  ${}^{1}H-{}^{1}H$  correlation spectroscopy ( ${}^{1}H-{}^{1}H$  COSY) and heteronuclear multiple bond correlation (HMBC) experiments as shown in Fig. 1. Thus,  ${}^{1}H-{}^{1}H$  COSY experiment indicated the presence of partial structures in bold lines. In the HMBC experiment on **1**, long-range correlations were observed between the following protons and carbons: 3-H and the formyl ester carbonyl carbon ( $\delta_c$  160.7); 23-H<sub>3</sub> and 3-5, 24-C; 24-H<sub>3</sub> and 3-5, 23-C; 25-H<sub>3</sub> and 1, 5, 9, 10-C; 26-H<sub>3</sub> and 7—9, 14-C; 27-H<sub>3</sub> and 8, 13—15-C; 28-H<sub>3</sub> and 16—18, 22-C; the formyl proton and 3-C. Finally, alkaline hydrolysis of **1** with 10% aqueous potassium hydroxide (KOH)–50% aqueous 1,4-dioxane (1:1, v/v) liberated 3-epi- $\alpha$ -amyrin<sup>37)</sup> (**1a**), so that the stereostructure of olibanumol K was determined to be 3-epi- $\alpha$ -amyrin formate (1).

**Structures of Olibanumols L (2), M (3), and N (4)** Olibanumol L (**2**) was also obtained as a white powder with positive optical rotation ( $[\alpha]_D^{28}+14.5$  in MeOH) and showed absorption bands at 3432 and  $1739 \text{ cm}^{-1}$  due to hydroxyl and



Chart 2

Table 1. <sup>13</sup>C-NMR Data for Olibanumols K (1), L (2), M (3), and N (4)

Position	$\mathbf{1}$	$\overline{2}$	3	$\overline{4}$
$\mathbf{1}$	33.8	35.7	33.6	34.9
$\overline{2}$	23.0	22.8	25.5	22.9
3	78.5	78.0	75.9	78.1
$\overline{4}$	36.5	36.5	37.5	36.5
5	50.0	49.9	48.8	49.8
6	18.2	18.1	18.3	18.1
7	32.8	33.3	33.6	33.1
8	40.2	43.4	43.5	43.0
$\mathbf{Q}$	47.5	55.5	51.8	52.6
10	36.9	38.0	38.1	38.0
11	23.3	68.1	76.6	76.7
12	124.3	128.4	124.0	123.5
13	139.5	142.7	143.4	143.3
14	42.2	42.1	42.1	42.1
15	28.2	27.9	27.9	27.9
16	26.7	26.7	26.7	26.8
17	33.8	33.5	33.4	33.5
18	59.1	58.0	58.5	58.4
19	39.7	39.4	39.5	39.5
20	39.7	39.3	39.3	39.3
21	31.3	31.1	31.1	31.1
22	41.6	41.2	41.4	41.4
23	28.8	28.6	28.6	28.1
24	22.0	22.0	22.4	22.0
25	15.6	16.6	16.8	16.9
26	17.0	18.1	18.2	18.2
27	23.5	23.2	22.6	22.6
28	28.1	28.7	28.7	28.7
29	17.6	17.5	17.4	17.3
30	21.5	21.3	21.3	21.4
$HCO-$	160.7			
$CH3CO-$		170.5		170.6
$CH_3CO-$		21.4		21.3
$CH3O-$			54.2	54.7

Measured in CDCl<sub>3</sub>.



Fig. 1. <sup>1</sup> H–<sup>1</sup> H COSY and HMBC Experiments of **1** and **2**

ester carbonyl functions in the IR spectrum. The molecular formula  $C_{32}H_{52}O_3$  was determined from the positive-ion FAB-MS  $[m/z 507 (M+Na)^+]$  and by high-resolution MS measurement. The  ${}^{1}$ H- (CDCl<sub>3</sub>) and  ${}^{13}$ C-NMR (Table 1) spectra of 2 indicated the presence of eight methyls  $\lceil \delta 0.81 \rceil$ , 0.86, 0.93, 1.19 (3H each, all s, 28, 24, 23, 27-H<sub>3</sub>), 1.11 (6H, s, 25, 26-H<sub>3</sub>), 0.86, 0.88 (3H each, both d, *J*=7.6 Hz, 29, 30-H<sub>3</sub>)], an acetyl [ $\delta$  2.03 (3H, s, CH<sub>3</sub>CO–)], two methines bearing an oxygen function  $\delta$  4.27 (1H, dd-like, 11-H), 4.61 (1H, br s, 3-H)], and an olefin  $\lbrack \delta$  5.18 (1H, br s, 12-H)] together with eight methylenes, five methines, and seven quaternary carbons. As shown in Fig. 1, the  $\mathrm{^{1}H-^{1}H}$  COSY experiment on **2** indicated the presence of partial structures written in bold lines. In the HMBC experiment on **2**, long-range correlations were observed between the following protons and carbons: 3-H and the acetyl carbonyl carbon ( $\delta_c$  170.5); 23-H<sub>3</sub> and 3-5, 24-C; 24-H<sub>3</sub> and 3-5, 23-C; 25-H<sub>3</sub> and 1, 5, 9, 10-C; 26-H<sub>3</sub> and 7-9, 14-C; 27-H<sub>3</sub> and 8, 13-15-C; 28-H<sub>3</sub> and  $16-18$ , 22-C. Thus, the connectivities of quaternary carbons (4, 8, 10, 13, 14, 17-C) and position of the acetyl group in **2** were clarified and its urs-12-ene-3,11-diol structure was elucidated. Finally, deacetylation of **2** with 0.14% sodium methoxide (NaOMe)–MeOH yielded urs-12-ene- $3\alpha$ ,11 $\alpha$ -diol (17). Consequently, the stereostructure of olibanumol L was determined to be 3-*O*-acetyl-urs-12-ene- $3\alpha$ , 11 $\alpha$ -diol (2).

Olibanumol M (**3**), a white powder with positive optical rotation ( $[\alpha]_D^{27}$ +21.0 in MeOH), showed absorption bands at 3475 and  $1735 \text{ cm}^{-1}$  due to hydroxyl and olefin functions in the IR spectrum. The positive-ion FAB-MS spectrum of **3** showed a quasimolecular ion peak at  $m/z$  479 (M+Na)<sup>+</sup>, and the molecular formula was determined to be  $C_{31}H_{52}O_2$  by high-resolution MS measurement. The  ${}^{1}H$ - (CDCl<sub>3</sub>) and  ${}^{13}C$ -NMR (Table 1) spectra of **3** indicated the presence of an urs-12-ene-3,11-diol part  $\lceil \delta \ 0.80, \ 0.87, \ 0.97, \ 1.08, \ 1.11, \ 1.16 \rceil$ (3H each, all s, 28, 24, 23, 25, 26, 27-H3), 0.86, 0.88 (3H each, both d, *J*=7.6 Hz, 29, 30-H<sub>3</sub>), 3.39 (1H, br s, 3-H), 3.85  $(1H, dd, J=3.9, 9.2 Hz, 11-H), 5.33 (1H, d-like, 12-H)$ ] together with a methoxyl group  $\lceil \delta \cdot 3.26 \rceil$  (3H, s, CH<sub>3</sub>O–)]. On the other hand, olibanumol N (4),  $C_{33}H_{54}O_3$ , was also obtained as a white powder with positive optical rotation  $([\alpha]_D^{22} + 1.1$  in MeOH). The <sup>1</sup>H- (CDCl<sub>3</sub>) and <sup>13</sup>C-NMR (Table 1) spectra of **4** were similar to those of **2**, except for the signals due to the 11-methoxyl group  $\delta$  3.28 (3H, s,  $CH_3O-$ ]. Methylation of 2 with methyl iodide ( $CH_3I$ ) and potassium carbonate  $(K_2CO_3)$  yielded **4**, whereas acetylation of  $3$  with Ac<sub>2</sub>O/pyridine furnished  $4$ . On the basis of this evidence, the stereostructures of olibanumols M and N were determined to be  $11\alpha$ -methyurs-12-ene-3 $\alpha$ ,11 $\alpha$ -diol (3) and its acetate (**4**).

## **Experimental**

The following instruments were used to obtain spectral and physical data: specific rotations, Horiba SEPA-300 digital polarimeter  $(l=5 \text{ cm})$ ; IR spectra, Shimadzu FTIR-8100 spectrometer; <sup>1</sup>H-NMR spectra, JEOL JNM-LA500 (500 MHz) and EX-270 (270 MHz) spectrometers; <sup>13</sup>C-NMR spectra, JEOL JNM-LA500 (125 MHz) and EX-270 (68 MHz) spectrometers with tetramethylsilane as an internal standard; FAB-MS and high resolution FAB-MS, JEOL JMS-SX 102A mass spectrometer; HPLC detector, Shimadzu RID-6A refractive index and SPD-10A UV–VIS detectors; HPLC column, YMC-Pack ODS-A and YMC-Pack SIL (YMC Co., Ltd., Kyoto, Japan)  $(250 \text{ mm} \times 4.6 \text{ mm} \text{ i.d.})$  and  $(250 \text{ mm} \times 20 \text{ mm} \text{ i.d.})$  columns were used for analytical and preparative purposes, respectively.

The following experimental conditions were used for chromatography: normal-phase silica gel column chromatography (CC), silica gel 60N (Kanto Chemical Co., Ltd., 63—210 mesh, spherical, neutral); reversed-phase silica gel CC, Chromatorex ODS DM1020T (Fuji Silysia Chemical, Ltd., 100— 200 mesh); normal-phase TLC, pre-coated TLC plates with silica gel  $60F_{254}$ (Merck, 0.25 mm); reversed-phase TLC, pre-coated TLC plates with silica gel RP-18 F<sub>254S</sub> (Merck, 0.25 mm); reversed-phase HPTLC, pre-coated TLC plates with silica gel RP-18 WF<sub>254S</sub> (Merck, 0.25 mm), detection was achieved by spraying with  $1\%$  Ce(SO<sub>4</sub>)<sub>2</sub>–10% aqueous H<sub>2</sub>SO<sub>4</sub>, followed by heating.

## **Plant Material** This item was described in a previous report.<sup>10)</sup>

**Extraction and Isolation** The gum-resin from *B. carterii* (2.7 kg) was extracted three times with 80% aqueous acetone at room temperature for 24 h. Evaporation of the solvent under reduced pressure provided an aqueous acetone extract (2131 g, 78.9%). The aqueous acetone extract (360.0 g) was subjected to normal-phase silica gel CC [3.0 kg, *n*-hexane–EtOAc (30 :  $1\rightarrow 20$  :  $1\rightarrow 10$  :  $1\rightarrow 1$  :  $1$ , v/v) $\rightarrow$ MeOH] to give 11 fractions [Fr. 1 (6.3 g), Fr. 2 (11.2 g), Fr. 3 (30.5 g), Fr. 4 (181.4 g), Fr. 5 (12.7 g), Fr. 6 (9.3 g), Fr. 7 (18.2 g), Fr. 8 (14.6 g), Fr. 9 (7.1 g), Fr. 10 (5.9 g), and Fr. 11 (15.6 g)], as reported previously.<sup>10)</sup> The fraction 2 (11.2 g) was subjected to normal-phase silica gel CC [345 g, *n*-hexane→*n*-hexane–EtOAc (100 : 1→80 : 1→30 : 1→ 20 : 1→10 : 1, v/v)→EtOAc] and HPLC [YMC-Pack SIL, UV detector (254 nm), *n*-hexane–EtOAc (300 : 1, v/v)] to give olibanumol K (**1**, 5.8 mg, 0.0013%) together with olibanumol  $F^{13}$  (37.8 mg, 0.0083%). The fraction 3 (3.0 g) was subjected to normal-phase silica gel CC [150 g, *n*-hexane–EtOAc  $(30:1, v/v) \rightarrow \text{MeOH}$  to give epilupeol acetate  $(6, 1288.9 \text{ mg}, 2.87%)$  and lupenone (**12**, 148.2 mg, 0.33%). The fraction 5 (12.7 g) subjected by reversed-phase silica gel CC [381 g, MeOH–H<sub>2</sub>O  $(70:30\rightarrow80:20\rightarrow90:10,$ v/v)→MeOH] and HPLC [YMC-Pack ODS-A, RI detector, MeOH-H<sub>2</sub>O (90 : 10, v/v)] to furnish lupeol (**9**, 3289.2 mg, 0.72%) together with olibanumol  $D^{13}$  (22.8 mg, 0.0050%). The fraction 6 (3.0 g) was subjected by reversed-phase silica gel CC [150 g, MeOH–H2O (70 : 30→80 : 20→90 : 10, v/v)→MeOH] and HPLC [YMC-Pack ODS-A, RI detector, MeOH-H<sub>2</sub>O (90 : 10 or 95 : 5, v/v)] to give neoilexonol acetate (**15**, 338.2 mg, 0.23%) and dammarenediol II acetate (**19**, 661.8 mg, 0.45%). The fraction 7 (9.0 g) was subjected to normal-phase silica gel CC [270 g, *n*-hexane–EtOAc (10 : 1  $\rightarrow$  $5:1\rightarrow1:1$ , v/v) $\rightarrow$ MeOH] to give six fractions [Fr. 7-1 (95.0 mg), Fr. 7-2 (369.7 mg), Fr. 7-3 (2.00 g), Fr. 7-4 (694.3 mg), Fr. 7-5 (3.50 g), and Fr. 7-6  $(2.10 \text{ g})$ ], as reported previously.<sup>13)</sup> The fraction 7-3 (2.0 g) was purified by HPLC [YMC-Pack ODS-A, RI detector, MeOH-H<sub>2</sub>O (95:5, v/v)] to furnish  $3\beta$ -acetoxylup-20(29)-en-11 $\beta$ -ol (11, 97.0 mg, 0.043%) and mansumbinol (**25**, 16.0 mg, 0.0071%). The fraction 7-4 (694.3 mg) was purified by HPLC [YMC-Pack ODS-A, RI detector, MeOH-H<sub>2</sub>O (95:5, v/v)] to furnish olibanumol L  $(2, 97.0 \text{ mg}, 0.043%)$  together with olibanumol  $G^{13}$  (11.3 mg, 0.0050%). The fraction 7-5 (3.5 g) was subjected by reversed-phase silica gel CC [175 g, MeOH–H<sub>2</sub>O (70 : 30→80 : 20→90 : 10, v/v)→MeOH] and HPLC [YMC-Pack ODS-A, RI detector, MeOH-H<sub>2</sub>O (90 : 10 or 95 : 5,  $v/v$ )] to give olibanumol N (**4**, 7.9 mg, 0.0035%), glochidiol (**8**, 3.8 mg, 0.0017%), neoilexonol (**14**, 88.0 mg, 0.039%), and **15** (97.0 mg, 0.043%). The fraction 7-6 (2.1 g) was purified by HPLC [YMC-Pack ODS-A, RI detector, MeOH–1% aqueous AcOH  $(95:5, v/v)$ ] to give dammarenodiol II  $(18,$ 200.8 mg, 0.089%) and ocotillol acetate (**23**, 9.9 mg, 0.0044%). The fraction 9 (7.1 g) subjected to reversed-phase silica gel CC [200 g, MeOH–H2O (70 : 30→90 : 10, v/v)→MeOH] and HPLC [YMC-Pack ODS-A, RI detector, MeOH–H<sub>2</sub>O (95 : 5, v/v)] to give lup-20(30)-ene-3 $\alpha$ , 29-diol (7, 22.8 mg, 0.0050%), lup-20(29)-ene-2 $\alpha$ ,3 $\beta$ -diol (10, 11.8 mg, 0.0026%), urs-9(11), 12-dien-3 $\beta$ -ol (13, 5.5 mg, 0.0012%), urs-12-ene-3 $\beta$ ,11 $\alpha$ -diol (16, 18.7 mg, 0.0041%), urs-12-ene-3 $\alpha$ ,11 $\alpha$ -diol (17, 68.3 mg, 0.015%), and 3-*O*-acetyl-3b,20*S*,24,trihydroxydammar-25-ene (**20**, 10.0 mg, 0.0022%) together with olibanumol  $E^{(13)}$  (13.2 mg, 0.0029%). The fraction 10 (5.9 g) was subjected by reversed-phase silica gel CC [180 g, MeOH–H<sub>2</sub>O (70 : 30 $\rightarrow$ 80 : 20 $\rightarrow$ 90:10,  $v/v$ ) $\rightarrow$ MeOH] and HPLC [MeOH–H<sub>2</sub>O (95:5,  $v/v$ )] to furnish olibanumol M (**3**, 19.7 mg, 0.0043%), isofouquierol acetate (**22**, 301.9 mg, 0.066%), and  $3\beta$ -hydroxymansumbin-13(17)-en-16-one (24, 29.7 mg, 0.0065%).

Olibanumol K (1): A white powder,  $[\alpha]_D^{27} + 16.2$  ( $c = 0.30$ , MeOH). Highresolution positive-ion FAB-MS: Calcd for  $C_{31}H_{50}O_2Na$   $(M+Na)^+$ 477.3709; Found 477.3703. IR (KBr, cm<sup>-1</sup>): 2940, 1725, 1457, 1379. <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$ : 0.80, 0.89, 0.92, 0.92, 1.11, 1.13 (3H each, all s, 28, 23, 24, 25, 27, 26-H<sub>3</sub>), 0.90 (3H, d, J=7.4 Hz, 29-H<sub>3</sub>), 0.92 (3H, d, J= 7.0 Hz, 30-H3), 4.77 (3H, br s, 3-H), 5.13 (1H, dd-like, 12-H), 8.13 (1H, s, HCO–). <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)  $\delta_c$ : given in Table 1. Positive-ion FAB-MS  $m/z$ : 477  $(M+Na)^+$ .

Olibanumol L (2): A white powder,  $[\alpha]_D^{28} + 14.5$  (*c*=1.00, MeOH). Highresolution positive-ion FAB-MS: Calcd for  $C_{32}H_{52}O_3Na$   $(M+Na)^+$ 507.3805; Found 507.3814. IR (KBr, cm<sup>-1</sup>): 3432, 2962, 1739, 1456, 1370. <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$ : 0.81, 0.86, 0.93, 1.19 (3H each, all s, 28, 24, 23, 27-H<sub>3</sub>), 1.11 (6H, s, 25, 26-H<sub>3</sub>), 0.86, 0.88 (3H each, both d, J=7.6 Hz, 29, 30-H<sub>3</sub>), 2.03 (3H, s, CH<sub>3</sub>CO–), 4.27 (1H, dd-like, 11-H), 4.61 (1H, br s, 3-H), 5.18 (1H, br s, 12-H). <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)  $\delta_c$ : given in Table 1. Positive-ion FAB-MS  $m/z$ : 507  $(M+Na)^+$ .

Olibanumol M (3): A white powder,  $[\alpha]_D^{27} + 21.0$  (*c*=1.00, MeOH). Highresolution positive-ion FAB-MS: Calcd for  $C_{31}H_{52}O_2Na$   $(M+Na)^+$ 479.3814; Found 479.3805. IR (KBr, cm<sup>-1</sup>): 3475, 2926, 1735, 1456, 1389. <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$ : 0.80, 0.87, 0.97, 1.08, 1.11, 1.16 (3H each, all s, 28, 24, 23, 25, 26, 27-H<sub>3</sub>), 0.86, 0.88 (3H each, both d,  $J=7.6$  Hz, 29, 30-H<sub>3</sub>), 3.26 (3H, s, CH<sub>3</sub>O-), 3.39 (1H, br s, 3-H), 3.85 (1H, dd, J=3.9, 9.2 Hz, 11-H), 5.33 (1H, d-like, 12-H). <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)  $\delta_c$ : given in Table 1. Positive-ion FAB-MS  $m/z$ : 479 (M+Na)<sup>+</sup>.

Olibanumol N (4): A white powder,  $[\alpha]_D^{22}$  +1.1 (*c*=1.00, MeOH). Highresolution positive-ion FAB-MS: Calcd for  $C_{33}H_{54}O_3$ Na  $(M+Na)^+$  521.3814; Found 521.3805. IR (KBr, cm<sup>-1</sup>): 2926, 1732, 1458, 1374. <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$ : 0.81, 0.86, 0.93, 1.04, 1.08, 1.19 (3H each, all s, 28, 24, 23, 25, 26, 27-H<sub>3</sub>), 0.86, 0.88 (3H each, both d,  $J=7.6$  Hz, 29, 30-H<sub>3</sub>), 2.03 (3H, s, CH<sub>3</sub>CO–), 3.28 (3H, s, CH<sub>3</sub>O–), 3.77 (1H, dd-like, 11-H), 4.61 (1H, br s, 3-H), 5.39 (1H, d-like, 12-H). <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)  $\delta_c$ : given in Table 1. Positive-ion FAB-MS  $m/z$ : 521  $(M+Na)^+$ .

**Deformylation of 1** A solution of 1 (3.0 mg) in 10% aqueous potassium hydroxide (KOH)–50% aqueous 1,4-dioxane (1 : 1, v/v, 2.0 ml) was stirred at 37 °C for 1 h. The reaction mixture was neutralized with Dowex HCR W2  $(H<sup>+</sup>$  form), and the resins were removed by filtration. After removal of the

solvent under reduced pressure, the residue was separated by normal-phase silica gel CC  $[1.0 g, n$ -hexane–EtOAc  $(3:1, v/v)$ ] to give 3-epi- $\alpha$ -amyrin (**1a**, 0.8 mg), which was identified by comparison of its physical and spectral data ( $[\alpha]_D$ , <sup>1</sup>H-NMR, and MS) with reported values.<sup>37)</sup>

**Deacetylation of 2** A solution of 2 (10.0 mg) in 0.14% sodium methoxide (NaOMe)–MeOH (2.0 ml) was stirred at room temperature for 3 h. The reaction mixture was neutralized with Dowex HCR W2  $(H<sup>+</sup>$  form), and the resins were removed by filtration. After removal of the solvent under reduced pressure, the residue was separated by normal-phase silica gel CC [ $1.0 \text{ g}$ , *n*-hexane–EtOAc  $(3:1, \text{ v/v})$ ] to give urs-12-ene-3 $\alpha$ ,11 $\alpha$ -diol (17, 7.5 mg), which was identified by comparison of its physical and spectral data  $([\alpha]_{D}$ , <sup>1</sup>H-NMR, and MS) with reported values.<sup>28)</sup>

**Methylation of 2** To a solution of **2** (10.0 mg) in dry dimethylformamide (DMF, 2.0 ml) was added methyl iodide (CH<sub>3</sub>I, 15  $\mu$ l) and potassium carbonate (K<sub>2</sub>CO<sub>3</sub>, 66.4 mg), and the mixture was stirred at 80 °C for 3 h. The reaction mixture was poured into brine and extracted with EtOAc. The extract was successively washed with saturated aqueous  $NaHCO<sub>3</sub>$  and brine, and filtrated. After removal of the solvent under reduced pressure, the residue was purified by normal-phase silica gel CC [1.0 g, *n*-hexane–EtOAc (3 : 1, v/v)] to give **4** (7.0 mg).

**Acetylation of 3** To a solution of **3** (5.0 mg) in dry pyridine (2.0 ml) was added acetic anhydride (1.0 ml), and the mixture was stirred at room temperature for 3 h. The reaction mixture was poured into ice-water and extracted with EtOAc. The extract was successively washed with 5% aqueous HCl, saturated aqueous NaHCO<sub>3</sub>, and brine, and filtrated. After removal of the solvent under reduced pressure, the residue was purified by normal-phase silica gel CC [1.0 g, *n*-hexane–EtOAc (2 : 1, v/v)] to give **4** (3.0 mg).

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