A Novel 3-Arylcoumarin and Three New 2-Arylbenzofurans from *Mucuna birdwoodiana*

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A novel coumarin (1, mucodianin A), three new 2-arylbenzofurans (2—4, mucodianins B—D) along with four known ones were isolated from the vine stems of *Mucuna birdwoodiana*. Their structures were elucidated on basis of spectral analysis. This is the first report of 7-quinonylcoumarin (1) as stable form in natural products.

Key words Mucuna birdwoodiana; Leguminosae; 3-arylcoumarin; 2-arylbenzofuran

Mucuna birdwoodiana TUTCH. (Leguminosae), distributed in southern China, has long been used as a traditional Chinese medicine named "ji-xue-teng." The vine stems of this plant are effective in promoting blood circulation or relieving stasis and have been used to treat pain or numbness of the wrists, knees or other joints and irregular menstruation.¹⁾ Our continuing study on searching for the bioactive constituents from the plant "ji-xue-teng" led to the isolation of a novel coumarin (1, mucodianin A), three new 2-arylbenzofurans (2—4, mucodianins B—D) and four known ones, 6demethylvignafuran (5), eryvarin L (6), isopterofuran (7), lespeflorin F_1 (8) from the 50% EtOH extract of *M. birdwoodiana*. We herein describe the isolation and structural elucidation of these compounds.

Results and Discussion

Compound **1** was obtained as a fluorescein solid. The negative HR-electrospray ionization (ESI)-MS exhibited the quasi-molecular ion peak $[M-H]^-$ at m/z 343.0827 (Calcd for C₁₈H₁₅O₇, 343.0823), suggesting the molecular formula C₁₈H₁₆O₇ of compound **1**. The UV spectrum showed the absorption maxima at 338, 260, 203 nm and the fluorescent nature suggested that it might be a 3-arylcoumarin derivative.^{2,3} The ¹H-NMR spectral data (Table 1) of **1** indicated the presence of three methoxyl signals at $\delta_{\rm H}$ 3.67, 3.72, 3.75 and five aromatic protons, appearing as one singlet at $\delta_{\rm H}$ 7.43; two pairs of doublets at $\delta_{\rm H}$ 6.84 and 6.23 (each 1H, J=8.8 Hz), $\delta_{\rm H}$ 6.78 and 6.59 (each 1H, J=8.4 Hz). The ¹³C-NMR spectrum (Table 1) displayed three methoxyl signals and fifteen nucleus signals. In the heteronuclear multiple bond correlations (HMBC) experiment (Table 1), the proton signal $\delta_{\rm H}$ 6.84 (1H, d, J=8.8 Hz) coupling to the typical carbon signal for coumarin (C-4) at $\delta_{\rm C}$ 143.2 must be assigned as H-5. Based on the correlations of H-5 to $\delta_{\rm C}$ 168.7 and H-4 to $\delta_{\rm C}$ 161.4, the signals $\delta_{\rm C}$ 168.7 and $\delta_{\rm C}$ 161.4 could be assigned as C-7 and C-2 respectively, which were opposite to the data of hydranget,⁴⁾ implying that **1** should be in the form of 7-quinonylcoumarin rather than in its normal form (1a). The methoxy at $\delta_{\rm H}$ 3.72 was located at C-8 on the basis of the HMBC spectrum ($\delta_{\rm H}$ 3.72 to C-8 at $\delta_{\rm C}$ 168.7). HMBC correlations of H-5' to C-1' and C-3'; H-6' to C-2' and C-3; methoxy at $\delta_{\rm H}$ 3.67 to C-2'; methoxy at $\delta_{\rm H}$ 3.75 to C-3', suggested 2',3'-dimethoxyl, 4'-hydroxyl groups substitued in ring C. Based on the above spectral evidence, the structure of 1 was determined as shown in Fig. 1. 1 and 1a which have never been discovered before are two tautomeric forms of the compound, named mucodianin A. Theoretically, structure 1a is more stable than 1. However, 1 was isolated in our experi-

Table 1. ¹H- and ¹³C-NMR Spectral Data and ²J and ³ J_{H-C} Correlations of Compound 1 (in DMSO)

No.	$\delta_{ m H}$	$\delta_{ m C}$	^{2}J	³ J
2		161.4		
3		110.3		
4	7.43 (s)	143.2	104.7 (C-4a)	161.4 (C-2), 148.8 (C-8a), 123.7 (C-5), 121.6 (C-1')
4a		104.7		
5	6.84 (d, 8.8)	123.7	104.7 (C-4a)	168.7 (C-7), 148.8 (C-8a), 143.2 (C-4)
6	6.23 (d, 8.8)	118.9		135.6 (C-8), 104.7 (C-4a)
7		168.7		
8		135.6		
8a		148.8		
1'		121.6		
2'		151.6		
3'		140.7		
4'		150.9		
5'	6.59 (d, 8.4)	111.2	150.9 (C-4')	121.6 (C-1'), 140.7 (C-3')
6'	6.78 (d, 8.4)	125.3	· · · ·	110.3 (C-3), 151.6 (C-2')
8-OMe	3.72 (s)	58.7		135.6 (C-8)
2'-OMe	3.67 (s)	60.2		151.6 (C-2')
3'-OMe	3.75 (s)	59.9		140.7 (C-3')

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ment and was stable in dimethyl sulfoxide (DMSO). It was reported that 7-hydroxycoumarin could be converted to 7auinonvlcoumarin in aquenous solutions, which mainly was caused by the transfer of a proton between two sites in the molecule by the participation of water moleculues.⁵⁾ The reason of why 1 was stable in DMSO may be the structure itself or external factors.

The UV spectra with maxima at 324 and 338 nm and the characteristic aromatic proton signal ($\delta_{\rm H}$ 6.98–7.04, br s) in the ¹H-NMR spectra indicated that compounds 2-8 all were 2-arvlbenzofuran derivatives.⁶⁾

Compound 2 was obtained as a yellow solid with the molecular formula C₁₆H₁₄O₅, deduced from the HR-ESI-MS $(C_{16}H_{14}O_5Na \ m/z \ 309.0739 \ [M+Na]^+, Calcd for \ 309.0733).$ The ¹H-NMR spectrum (Table 2) displayed two broad proton singlets at $\delta_{\rm H}$ 6.98 and 6.94, assignable to the two aromatic protons H-3 and H-7 of 2-arylbenzofuran. One ABX coupling system at $\delta_{\rm H}$ 6.53 (d, J=2.4 Hz), 6.46 (dd, J=8.7, 2.4 Hz), and 7.63 (d, J=8.7 Hz) similar to the ¹H-NMR data of 5,7) was assigned to the 2'-methoxyl, 4'-hydroxyl-2phenyl moiety (ring C). This was also confirmed by the HMBC spectrum (Fig. 2). Another methoxy at $\delta_{\rm H}$ 3.79 must be positioned at C-5 and one aromatic proton singlet at $\delta_{\rm H}$ 7.07 assigned as H-4 determined by the key nuclear Overhauser effect (NOE) correlations between H-4 at $\delta_{
m H}$ 7.07 and 5-OMe at $\delta_{\rm H}$ 3.79, H-3 at $\delta_{\rm H}$ 6.98. The ¹³C-NMR (Table 2) and HMBC (Fig. 2) experiments confirmed this structure of 2 and assigned all carbon resonances. Thus, it was determined to be 2-(4-hydroxy-2-methoxylphenyl)-5-methoxyl-

1a

Fig. 1. Chemical Structures of Compounds 1 and 1a

benzofuran-6-ol, named mucodianin B.

Compound 3 was isolated as a pale yellow solid. The molecular formula was established as C₁₇H₁₆O₆ by HR-ESI-MS, which displayed a quasi-molecular ion peak at m/z 339.0843 [M+Na]⁺, Calcd for 339.0839. The ¹H-NMR spectrum (Table 2) showed the presence of one proton singlet at $\delta_{\rm H}$ 7.10 (H-4), two broad proton singlets at $\delta_{\rm H}$ 6.96 (H-7) and 7.03 (H-3), one methoxyl group at $\delta_{\rm H}$ 3.79. Its similatity to that of 2 indicated that 5-methoxyl, 6-hydroxyl groups substituted in ring A. The AB coupling system at $\delta_{\rm H}$ 6.71 (d, J=8.7 Hz) and 7.38 (d, J=8.7 Hz) in ¹H-NMR spectrum (Table 2) indicated that there were three substituents in ring C. Except two methoxys at $\delta_{\rm H}$ 3.79 and 3.85, it was deduced that there should be a hydroxyl group according to the molecular formula. In the NOE spectrum, the methoxy at $\delta_{\rm H}$ 3.85 correlating to H-3 meant it should be at C-2'. Due to one methoxy resonate in the range 60-63 ppm in the ¹³C-NMR spectrum, which required the presence of both orthopositions substituted,⁸⁾ the methoxy at $\delta_{\rm H}$ 3.79 was determined to be at C-3' and the hydroxy was assigned to be at C-4'. The structure was further confiremed by the HMBC spectrum (Fig. 3) and also by comparison with the data of ring C of 7.7) Therefore, the structure of 3 was determined as 2-(4hydroxy-2,3-dimethoxylphenyl)-5-methoxylbenzofuran-6-ol and named mucodianin C.

Compound 4, a brown solid, had the molecular formula of C₁₆H₁₄O₆ established by HR-ESI-MS (m/z 303.0877 [M+H]⁺, Calcd for 303.0863). Its ¹H-NMR spectral data (Table 2) were reminiscent of those of 3, only missing the signal of 3'-methoxy at $\delta_{\rm H}$ 3.79 and the hydroxyl group substituted, so the AB type proton signals upfield shifted to $\delta_{\rm H}$

Fig. 2. Chemical Structure and Key HMBC Correlations of Compound 2

Table 2. ¹H-NMR (300 MHz) and ¹³C-NMR Spectral Data of Compounds 2-4 in DMSO

No.	2		3		4	
	$\delta_{_{ m H}}$	$\delta_{\mathrm{C}}{}^{a)}$	$\delta_{ m H}$	$\delta_{\mathrm{C}}{}^{\scriptscriptstyle b)}$	$\delta_{ m H}$	$\delta_{ m c}{}^{a)}$
2		150.8		150.3		145.4
3	6.98 (br s)	103.6	7.03 (br s)	103.5	7.02 (br s)	103.1
3a		120.5		120.4		120.5
4	7.07 (s)	103.0	7.10 (s)	102.9	7.10 (s)	103.0
5		145.3		145.4		145.4
6		144.9		145.2		145.0
7	6.94 (br s)	97.8	6.96 (br s)	97.8	6.95 (br s)	97.8
7a		147.8		148.0		148.0
1'		110.4		115.5		115.4
2'		157.0		150.4		150.9
3'	6.53 (d, 2.4)	99.4		141.3		138.9
4'		158.6		151.3		147.0
5'	6.46 (dd, 8.7, 2.4)	107.6	6.71 (d, 8.7)	112.3	6.64 (d, 8.7)	111.3
6'	7.63 (d, 8.7)	126.6	7.38 (d, 8.7)	120.6	7.12 (d, 8.7)	115.8
5-OMe	3.79 (s)	56.1	3.79 (s)	56.1	3.79 (s)	56.1
2'-OMe	3.88 (s)	55.4	3.85 (s)	59.9	3.76 (s)	59.0
3'-OMe			3.79 (s)	60.2		

a) Recorded in 100 MHz, b) recorded in 125 MHz.





Fig. 3. Chemical Structures and Key HMBC Correlations of Compounds ${\bf 3} \text{ and } {\bf 4}$

6.64 (d, J=8.7 Hz) and 7.12 (d, J=8.7 Hz). The key NOEs (H-3/2'-OMe), the molecular formula and the HMBC spectral data (Fig. 3) also supported the structure of 4. Thus, it was characterized as 2-(3,4-dihydroxy-2-methoxylphenyl)-5-methoxylbenzofuran-6-ol and named mucodianin D.

Phytochemical study of this plant also resulted in the isolation of four known 2-arylbenzofurans: 6-demethylvignafuran (5),⁷⁾ eryvarin L (6),⁹⁾ isopterofuran (7),⁷⁾ and lespeflorin F_1 (8),¹⁰⁾ which were characterized by comparison of the respective spectral data (¹H-, ¹³C-NMR, MS) with those found in the literatures.

2-Arylbenzofuran derivatives isolated from leguminous plants were considered to be of isoflavonoid origin,²⁾ the cooccurrence of compounds 1 and 8 also confirmed this hypothesis.

Experimental

General Experimental Procedures Melting points were determined by XT Digital Melting-Point Apparatus with Microscope and uncorrected. UV spectra were obtained by JASCO U-650 spectrophotometer. IR spectra were run on IMPACT 400 spectrometer. HR-ESI-MS were performed on Autospec Ultima-TOF mass spectrometer and ESI-MS on Agilent 1100 LC/MSD Trap-SL mass spectrometer. ¹H-NMR (500 MHz), ¹³C-NMR (125 MHz), NOE difference, heteronuclear single quantum coherence (HSQC) and HMBC spectra were run on INOVA-500 spectrometer with TMS as internal standard, ¹H-NMR (400 MHz), ¹³C-NMR (100 MHz), NOE difference, HSQC and HMBC spectra were recorded on Mercury-400 spectrometer, and ¹H-NMR (300 MHz) spectrum on Mercury-300 spectrometer. Sephadex LH-20 (Pharmacia), Macroporous resin D101 (26-60 mesh, tianjin, China) and Polyamide resin (30-50 mesh, zhejiang, China) were used for Column Chromatography. HPLC separations were performed on preparation YMC-Pack ODS-A column (10 μ m, 250×20 mm i.d.) equipped with Shimadzu SPD-6A UV spectrophotometric detector and Thermo constametric pumping system.

Plant Material The vine stem of *M. birdwoodiana* was collected in the County of Jin Xiu, Guangxi Province, China, in August 2006, and identified by Prof. Shou-yang Liu, Guangxi Traditional Chinese Medical College. A voucher specimen (No. S2263) has been deposited in the herbarium of Institute of Materia Medica, Chinese Academy of Medical Sciences & Peking Union Medical College.

Extraction and Isolation Crushed dry vine stem of *M. birdwoodiana* (10 kg) was extracted with 50% EtOH (3×101) under reflux to yield a crude extract (1600 g). The extract was passed over a D101 macroporous resin column chromatography (CC) (8 kg) eluting with a gradient of aqueous EtOH (0, 30, 60, 75, 100%, v/v) to yield five fractions (Fr. 1—5). Fr. 3 (145 g) was further fractionated on a Polyamide resin CC (2.5 kg) eluting with a gradient of aqueous EtOH (0, 15, 30, 50, 70, 100%, v/v) to give six fractions (Frs. 3.1—3.6). Fr. 3.4 (16.3 g) was separated by reversed-phase silica gel (300 g) CC and eluted with 30—70% aqueous MeOH to provide nine fractions (Frs. 3.4.1—3.4.9). Fr. 3.4.4 (624 mg) was applied to repeated ODS, Sephadex

LH-20 CC, and preparative HPLC (43% aqueous MeOH, 4 ml/min) to give 4 (4.5 mg, $t_{\rm R}$ =45.3 min) and 1 (5.2 mg, $t_{\rm R}$ =87.0 min). Fr. 3.4.5 (390 mg) was purified by Sephadex LH-20 CC and preparative HPLC (48% aqueous MeOH, 4 ml/min) to yield compounds 2 (6.2 mg, $t_{\rm R}$ =75.2 min) and 7 (2.3 mg, $t_{\rm R}$ =94.0 min). Fr. 3.4.6 (1.0 g) was further purified by repeated ODS CC and preparative HPLC (50% aqueous MeOH, 4 ml/min) to give compounds 3 (3.6 mg, $t_{\rm R}$ =64.2 min) and 8 (5.3 mg, $t_{\rm R}$ =72.0 min). Fr. 4 (10.6 g) was subjected to silica gel CC (160—200 mesh, 400 g), eluted with CHCl₃-MeOH (100:0—50:1—20:1—10:1—8:1—4:1—1:1, v/v) to provide eight fractions. Fr. 4.2 (810 mg) was purified by Sephadex LH-20 CC and preparative HPLC (47% aqueous MeOH, 4 ml/min) to yield compound 5 (7.2 mg, $t_{\rm R}$ =84.0 min).

Mucodianin A (1): Fluorescein solid (MeOH). mp 116.2—117.5 °C. UV (MeOH) λ_{max} (log ε): 338 (3.81), 260 (3.76), 203 (4.33) nm. IR v_{max} : 3372, 2918, 2850, 1706, 1589, 1503, 1465, 1415, 1362, 1312, 1069, 983, 827 cm⁻¹. HR-ESI-MS *m/z*: 343.0827 [M–H]⁻ (Calcd for C₁₈H₁₅O₇, 343.0823). ¹H-NMR (300 MHz) and ¹³C-NMR (100 MHz) in DMSO: see Table 1.

Mucodianin B (2): Yellow solid (MeOH). mp 155.1—156.6 °C. UV (MeOH) λ_{max} (log ε): 342 (4.52), 327 (4.47), 282 (4.18), 208 (4.68) nm. IR v_{max} : 3167, 2928, 2850, 1716 (w), 1612, 1488 1460, 1429, 1330, 1308, 1202, 1025, 960, 837 cm⁻¹. HR-ESI-MS *m/z*: 309.0739 [M+Na]⁺ (Calcd for C₁₆H₁₄O₅Na, 309.0733). ¹H-NMR (300 MHz) and ¹³C-NMR (100 MHz) in DMSO: see Table 2.

Mucodianin C (3): Pale yellow solid (MeOH). mp 125.4—126.5 °C. UV (MeOH) λ_{max} (log ε): 338 (4.33), 324 (4.34), 293 (4.08), 284 (4.07), 211 (4.41) nm. IR v_{max} : 3394, 2917, 2850, 1715(w), 1605, 1468, 1420, 1329, 1203, 1030, 961 cm⁻¹. HR-ESI-MS *m/z*: 339.0843 [M+Na]⁺ (Calcd for C₁₇H₁₆O₆Na, 339.0839). ¹H-NMR (300 MHz) and ¹³C-NMR (100 MHz) in DMSO: see Table 2.

Mucodianin D (4): Brown solid (MeOH). mp 140.6—142.0 °C. UV (MeOH) λ_{max} (log ε): 338 (4.18), 324 (4.22), 293 (3.97), 285 (3.95), 212 (4.23) nm. IR v_{max} : 3352, 2925, 2851, 1609, 1467, 1424, 1329, 1205, 1124, 1024, 961, 932 cm⁻¹. HR-ESI-MS *m/z*: 303.0877 [M+H]⁺ (Calcd for C₁₆H₁₅O₆, 303.0863). ¹H-NMR (300 MHz) and ¹³C-NMR (125 MHz) in DMSO: see Table 2.

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