

## Securinega Alkaloids from *Flueggea leucopyra*

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Six new Securinega alkaloids (**1**, **3**, **5**, **7**, **9**, **10**) together with four known ones were isolated from the twigs and leaves of *Flueggea leucopyra*. The structures of new compounds were established on the basis of the spectroscopic methods including UV, IR, HR-electrospray ionization (ESI)-MS, 1D and 2D NMR, and the absolute configurations of these new alkaloids were assigned by the modified Mosher's method and the circular dichroism (CD) spectra.

**Key words** *Flueggea leucopyra*; Euphorbiaceae; Securinega alkaloid; absolute configuration

The Securinega alkaloids are a group of polycyclic compounds isolated from the plants of Securinega and Phyllanthus genera (Euphorbiaceae).<sup>1</sup> Previous phytochemical investigations had led to the isolation of a number of Securinega alkaloids,<sup>1–8</sup> which exhibited antimalarial, antibacterial and antitumor activities.<sup>8–11</sup> Among the Securinega alkaloids, securinine was a major alkaloid obtained from the plant *Securinega suffruticosa*,<sup>12</sup> and clinically applied to treat sequela of poliomyelitis and aplastic anemia.<sup>13</sup> Pharmacology investigations indicated that securinine was a stereospecific GABA<sub>A</sub> receptor antagonist with a significant central nervous system (CNS) activity.<sup>14,15</sup> In searching for bioactive alkaloids from the Euphorbiaceae plants, we had isolated some chemical constituents from *Securinega suffruticosa* and *Flueggea virosa*.<sup>3,4</sup> *Flueggea leucopyra* was a shrub which only distributed in Sichuan and Yunnan Provinces of China. Recently, our phytochemical study of *F. leucopyra* had resulted in the isolation of six new Securinega alkaloids together with four known ones (Fig. 1). This paper reports the isolation and structural elucidation of the new alkaloids (**1**, **3**, **5**, **7**, **9**, **10**) from the twigs and leaves of *F. leucopyra*.

### Results and Discussion

The air-dried twigs and leaves of *F. leucopyra* were extracted with 95% EtOH. The solution was evaporated *in vacuo* to get a residue. The residue was suspended in H<sub>2</sub>O and then adjusted to pH 6 with 5% HCl. The acidic suspension was partitioned with CHCl<sub>3</sub> to remove the neutral component. The aqueous layer was basified with NH<sub>3</sub>·H<sub>2</sub>O and extracted with CHCl<sub>3</sub> to obtain a residue. Repeated column chromatography of the residue afforded six new compounds (**1**, **3**, **5**, **7**, **9**, **10**) and four known compounds (**2**, **4**, **6**, **8**). The known compounds were identified by comparison with the

literature data as securitinine (**2**),<sup>5</sup> secu'amamine B (**4**),<sup>8</sup> secu'amamine C (**6**),<sup>8</sup> and fluggeainol (**8**),<sup>6,7</sup> respectively.

**1** was obtained as yellow oil, and its molecular formula was determined as C<sub>13</sub>H<sub>15</sub>NO<sub>3</sub> on the basis of HR-electrospray ionization (ESI)-MS at *m/z*: 234.1127 [M+H]<sup>+</sup> (Calcd for C<sub>13</sub>H<sub>16</sub>NO<sub>3</sub>, 234.1125). The IR spectrum of **1** implied the presence of hydroxyl group (3436 cm<sup>-1</sup>) and α,β-unsaturated γ-lactone ring (1746, 1632 cm<sup>-1</sup>). An α,β-unsaturated γ-lactone ring at δ<sub>C</sub> 174.0, 168.8, 110.7 and 92.9, a double bond at δ<sub>C</sub> 124.3 and 150.1, and an oxygenated methine at δ<sub>C</sub> 64.9 were observed in the <sup>13</sup>C-NMR spectrum. Accordingly, the <sup>1</sup>H-NMR spectrum showed three olefinic protons at δ<sub>H</sub> 5.71 (1H, br s), 6.64 (1H, d, *J*=9.2 Hz) and 6.77 (1H, dd, *J*=9.2, 5.2 Hz), and an oxygenated proton at δ<sub>H</sub> 4.19 (1H, m) (Table 1). All the data indicated that **1** was a securinine type alkaloid with a hydroxyl group.<sup>5</sup>

Comparison of <sup>1</sup>H- and <sup>13</sup>C-NMR data of **1** (Table 1) with those of securitinine<sup>5</sup> (**2**) revealed that the signals of the two compounds were very similar, except the absence of the methoxyl group in **1** as well as the upfield shift of C-4 from δ<sub>C</sub> 72.8 in **2** to δ<sub>C</sub> 64.9 in **1**, suggesting that **1** had a hydroxyl group at C-4 position. This was further confirmed by the heteronuclear multiple bond connectivity (HMBC) correlations between H-4 (δ<sub>H</sub> 4.19) and C-2 (δ<sub>C</sub> 57.0) and C-6 (δ<sub>C</sub> 43.7) (Fig. 2). The relative configuration of **1** was assigned as shown in Fig. 3 by the nuclear Overhauser effect spectroscopy (NOESY) correlations between H-2 and H-6a, between H-4 and H-6b, as well as between H-2 and H-8a. The circular dichroism (CD) spectrum of **1** showed Cotton effects at λ<sub>max</sub> 216 nm (Δε -3.34) and 312 nm (Δε -17.37), which were similar to those of securitinine (**2**) [λ<sub>max</sub> 227 nm (Δε -1.13) and 302 nm (Δε -19.36)]. Furthermore, the absolute configuration of **1** was confirmed by application of the modified Mosher's method.<sup>16</sup> Differences of proton chemical shift (Δδ values, δ<sub>S</sub>-δ<sub>R</sub>) between (*S*)-α-methoxy-α-(trifluoromethyl)phenylacetic acid (MTPA) ester (**1a**) and (*R*)-MTPA ester (**1b**) (Fig. 4) indicated the presence of *S* configuration at C-4 in **1**. Hence, the absolute configuration of **1** was assigned as 2*S*, 4*S*, 7*S*, and 9*S*. **1** was identified as 4α-hydroxyallosecurinine.

**3** was isolated as yellow oil, and its HR-ESI-MS showed an [M+H]<sup>+</sup> ion peak at *m/z*: 266.1387 (Calcd for C<sub>14</sub>H<sub>20</sub>NO<sub>4</sub>, 266.1387) for the molecular formula of C<sub>14</sub>H<sub>19</sub>NO<sub>4</sub>. The presence of α,β-unsaturated γ-lactone ring

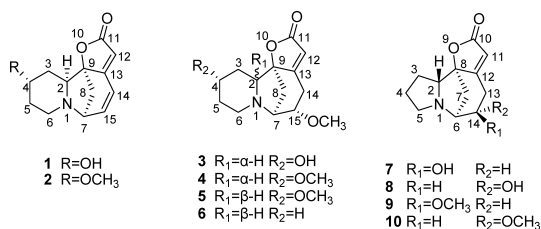


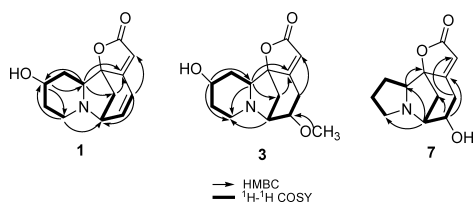
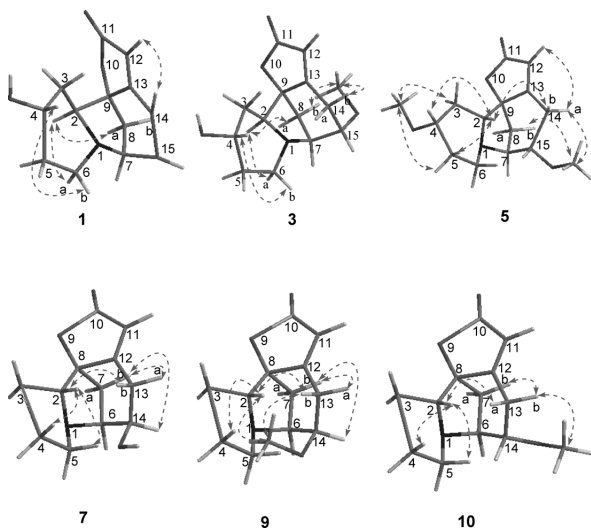
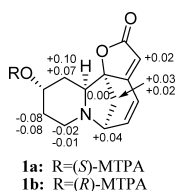
Fig. 1. Chemical Structures of Compounds **1**–**10**

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Table 1. NMR Data of **1**, **3** and **5** (CDCl<sub>3</sub>, *J* in Hz)

Position	<b>1</b>		<b>3</b>		<b>5</b>	
	$\delta_C$	$\delta_H$	$\delta_C$	$\delta_H$	$\delta_C$	$\delta_H$
2	57.0	3.95 (dd, 12.8, 4.0)	61.5	3.51 <sup>a)</sup>	60.2	2.96 <sup>a)</sup>
3	32.5	1.39 <sup>a)</sup> 1.35 <sup>a)</sup>	34.0	1.62 <sup>a)</sup>	33.8	2.14 (m)
4	64.9	4.19 (m)	66.0	1.06 (m)	77.4	1.28 <sup>a)</sup>
5	34.4	2.18 (m)	35.5	4.14 (t, 2.8)	32.1	3.23 <sup>a)</sup>
6	43.7	1.53 (m)	47.5	1.69 <sup>a)</sup>	48.9	1.80 (m)
		2.78 (m)		1.61 <sup>a)</sup>		1.27 <sup>a)</sup>
7	60.1	2.56 (m)	60.1	3.04 (m)	62.9	3.06 <sup>a)</sup>
		3.89 (brt, 4.8)		2.79 <sup>a)</sup>		2.95 <sup>a)</sup>
8	44.1	2.67 (dd, 10.0, 4.0)	36.2	3.35 (brt, 5.2)	33.6	3.33 <sup>a)</sup>
		1.91 (d, 10.0)		2.34 (dd, 10.8, 4.6)		2.51 (dd, 11.2, 4.8)
9	92.9	—	91.0	—	92.3	—
11	174.0	—	174.1	—	174.6	—
12	110.7	5.71 (brs)	114.6	5.68 (d, 2.4)	112.6	5.64 (brs)
13	168.8	—	172.2	—	174.1	—
14	124.3	6.64 (d, 9.2)	32.1	2.95 (m)	30.6	2.84 <sup>a)</sup>
		—		2.74 <sup>a)</sup>		2.82 <sup>a)</sup>
15	150.1	6.77 (dd, 9.2, 5.2)	81.0	3.55 <sup>a)</sup>	81.0	3.56 (d, 2.8)
4-OCH <sub>3</sub>	—	—	—	—	56.6	3.35 (s)
15-OCH <sub>3</sub>	—	—	58.1	3.27 (s)	58.2	3.30 (s)

a) Overlapped signals were reported without designating multiplicity.

Fig. 2. Key HMBC and <sup>1</sup>H-<sup>1</sup>H COSY Correlations of **1**, **3** and **7**Fig. 3. Key NOESY Correlations of **1**, **3**, **5**, **7**, **9** and **10**Fig. 4.  $\Delta\delta$  Values ( $\delta_S - \delta_R$ ) of the MTPA Esters **1a** and **1b**

was suggested by its IR spectrum (1756, 1643 cm<sup>-1</sup>). Fourteen carbon signals including one methyl, five methylenes, five methines, and three quaternary carbons were showed by the <sup>13</sup>C and distortionless enhancement by polarization transfer (DEPT) NMR spectra. An  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone ring ( $\delta_C$  174.1, 172.2, 114.6, 91.0), two oxygenated methines ( $\delta_C$  66.0, 81.0) and a methoxyl group ( $\delta_C$  58.1) were displayed in the <sup>13</sup>C-NMR spectrum. In the <sup>1</sup>H-NMR spectrum, an olefinic proton ( $\delta_H$  5.68, d, *J*=2.4 Hz), two oxygenated protons [ $\delta_H$  3.55 (m) and 4.14 (t, *J*=2.8 Hz)] and a methoxy singlet ( $\delta_H$  3.27, s) were observed. The above data indicated that **3** was a dihydrosecurinine type alkaloid with a methoxyl group and a hydroxyl group.<sup>8)</sup>

Comparison of NMR data of **3** (Table 1) with those of secu'amamine B<sup>8)</sup> (**4**) showed that the NMR signals of the two compounds were similar, except that **3** only had a methoxyl group as well as the <sup>13</sup>C-NMR value at C-4 shifted from  $\delta_C$  76.2 in **4** to  $\delta_C$  66.0 in **3**, suggesting that **3** had a methoxyl group at C-15 position and a hydroxyl group at C-4 position. It was further confirmed by the HMBC correlations between OCH<sub>3</sub> ( $\delta_H$  3.27) and C-15 ( $\delta_C$  81.0) (Fig. 2). The NOESY correlations between H-2 and H-6a, between H-6b and H-4, between H-2 and H-8a, between H-8b and 15-OMe, as well as between 15-OMe and H-14b (Fig. 3) established the relative configuration of **3**. The Cotton effects at  $\lambda_{max}$  226 nm ( $\Delta\epsilon$ +2.87) and 277 nm ( $\Delta\epsilon$ +0.76) in CD spectrum of **3** were similar to those of **4**.<sup>8)</sup> Therefore, the absolute configuration of **3** was assigned as 2*S*, 4*S*, 7*S*, 9*S*, and 15*S*. The structure of **3** was elucidated as 4 $\alpha$ -hydroxy-15 $\alpha$ -methoxy-14,15-dihydroallosecurinine.

**5** showed an [M+H]<sup>+</sup> ion peak at *m/z*: 280.1540 (Calcd for C<sub>15</sub>H<sub>22</sub>NO<sub>4</sub>, 280.1543) in the HR-ESI-MS spectrum, consistent with a molecular formula of C<sub>15</sub>H<sub>21</sub>NO<sub>4</sub>. The IR spectrum indicated the presence of  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone ring (1758, 1644 cm<sup>-1</sup>). The <sup>1</sup>H- and <sup>13</sup>C-NMR data of **5** (Table 1) were very similar to those of secu'amamine B (**4**),<sup>8)</sup> suggesting that **5** was a dihydrosecurinine type alkaloid with

Table 2. NMR Data of **7**, **9** and **10** (CDCl<sub>3</sub>, *J* in Hz)

Position	<b>7</b>		<b>9</b>		<b>10</b>	
	$\delta_C$	$\delta_H$	$\delta_C$	$\delta_H$	$\delta_C$	$\delta_H$
2	65.6	3.14 <sup>a)</sup>	66.7	3.22 <sup>a)</sup>	66.9	3.15 (dd, 8.8, 6.4)
3	28.9	1.89 <sup>a)</sup>	29.1	1.91 <sup>a)</sup>	30.3	1.90 <sup>a)</sup>
4	26.6	1.81 <sup>a)</sup>	26.5	1.75 <sup>a)</sup>	27.9	1.71 <sup>a)</sup>
		1.97 <sup>a)</sup>		1.96 <sup>a)</sup>		1.95 <sup>a)</sup>
5	57.2	1.79 <sup>a)</sup>	57.6	1.79 <sup>a)</sup>	58.6	1.71 <sup>a)</sup>
		3.36 (m)		3.47 (m)		3.36 (m)
6	66.0	2.63 (m)	63.3	2.62 <sup>a)</sup>	64.7	2.62 (m)
7	29.5	3.10 <sup>a)</sup>	31.5	3.24 <sup>a)</sup>	31.2	3.22 (m)
		2.33 (dd, 11.6, 6.4)		2.51 (dd, 11.6, 6.4)		2.27 (dd, 11.6, 6.4)
8	91.8	1.96 (d, 6.4)	91.4	1.31 (d, 6.4)	92.5	1.81 (d, 6.4)
10	172.9	—	172.8	—	173.9	—
11	111.1	5.66 (d, 2.0)	110.6	5.64 (d, 2.4)	112.3	5.59 (d, 2.0)
12	172.9	—	171.5	—	173.9	—
13	32.2	2.93 <sup>a)</sup>	29.4	3.22 <sup>a)</sup>	30.1	2.88 (d, 12.8)
		2.80 <sup>a)</sup>		2.64 <sup>a)</sup>		2.77 (m)
14	69.1	4.24 (t, 4.4)	79.8	3.34 (dd, 6.4, 2.0)	79.1	3.67 (dd, 4.8, 4.4)
14-OCH <sub>3</sub>			56.6	3.41 (s)	58.1	3.26 (s)

a) Overlapped signals were reported without designating multiplicity.

two methoxyl groups. The HMBC spectrum showed the correlations between OCH<sub>3</sub> ( $\delta_H$  3.35) and C-4 ( $\delta_C$  77.4) as well as between OCH<sub>3</sub> ( $\delta_H$  3.30) and C-15 ( $\delta_C$  81.0), indicating that two methoxyl groups were located at C-4 and C-15 positions, respectively. The NOE correlation between H-2 and H-14b (Fig. 3) indicated that the relative configuration at C-2 of **5** was opposite to those of **3** and **4**. The absolute configuration of **5** was determined as 2*R*, 4*S*, 7*S*, 9*S*, and 15*S* by comparison of the CD spectrum of **5** with that of secu'amamine C (**6**).<sup>8)</sup> Therefore, the structure of **5** was elucidated as 4 $\alpha$ -methoxy-15 $\alpha$ -methoxy-14,15-dihydrosecurinine.

**7** was obtained as yellow oil. The molecular formula of **7** was determined as C<sub>12</sub>H<sub>15</sub>NO<sub>3</sub> according to the HR-ESI-MS at *m/z*: 222.1126 [M+H]<sup>+</sup> (Calcd for C<sub>12</sub>H<sub>16</sub>NO<sub>3</sub>, 222.1125). The IR spectrum showed the presence of hydroxyl group (3442 cm<sup>-1</sup>) and  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone ring (1755, 1631 cm<sup>-1</sup>). The <sup>1</sup>H-NMR spectrum showed an olefinic proton at  $\delta_H$  5.66 (d, *J*=2.0 Hz) and an oxygenated proton at  $\delta_H$  4.24 (t, *J*=4.4 Hz). The <sup>13</sup>C-NMR spectrum displayed twelve carbon signals including five methylenes, four methines and three quaternary carbons. Furthermore, in the <sup>13</sup>C-NMR spectrum, an  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone ring at  $\delta_C$  172.9, 172.9, 111.1 and 91.8 as well as an oxygenated methine group at  $\delta_C$  69.1 were observed. The above data were very similar to those of flugeainol (**8**),<sup>6,7)</sup> indicating that **7** was a dihydronorsecurinine type alkaloid with a hydroxyl group. The position of hydroxyl group was assigned by the HMBC correlations between H-14 ( $\delta_H$  4.24) and C-6 ( $\delta_C$  66.0), C-7 ( $\delta_C$  29.5) and C-12 ( $\delta_C$  172.9) (Fig. 2). The relative configuration of **7** was assigned by the NOE correlations between H-2 and H-13b, between H-7b and H-13a, as well as between H-7b and H-14 (Fig. 3). The Cotton effects of **7** [ $\lambda_{max}$  219 nm ( $\Delta\epsilon$ -2.76), 238 nm ( $\Delta\epsilon$ +1.74), and 265 nm ( $\Delta\epsilon$ -0.85)] were also similar to those of flugeainol (**8**) [ $\lambda_{max}$  218 nm ( $\Delta\epsilon$ -1.37), 231 nm ( $\Delta\epsilon$ +2.67), and 261 nm ( $\Delta\epsilon$ -0.27)]. Hence, the absolute configuration of **7** was assigned as 2*R*, 6*S*, 8*S*, and 14*R*. **7** was identified as 14 $\beta$ -hydroxy-13,14-dihydronorsecurinine.

**9** was isolated as yellow oil. Its molecular formula was determined as C<sub>13</sub>H<sub>17</sub>NO<sub>3</sub> by the HR-ESI-MS at *m/z*: 236.1286 [M+H]<sup>+</sup> (Calcd for C<sub>13</sub>H<sub>18</sub>NO<sub>3</sub>, 236.1281). The IR spectrum of **9** implied the presence of  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone ring (1755, 1649 cm<sup>-1</sup>). Except for an additional methoxyl group ( $\delta_H$  3.41/ $\delta_C$  56.6), the <sup>1</sup>H- and <sup>13</sup>C-NMR data of **9** were similar to those of **7** (Table 2), suggesting **9** was a dihydronorsecurinine type alkaloid with a methoxyl group. The HMBC correlations between OCH<sub>3</sub> ( $\delta_H$  3.41) and C-14 ( $\delta_C$  79.8) indicated that the methoxyl group was attached to C-14 position. The relative configuration of **9** was assigned by NOESY correlations (Fig. 3). The CD spectrum [ $\lambda_{max}$  208 nm ( $\Delta\epsilon$ -2.56), 234 nm ( $\Delta\epsilon$ +2.25), and 284 nm ( $\Delta\epsilon$ -0.62)] of **9** was similar to those of **7** and **8**, suggesting the absolute configuration of **9** was 2*R*, 6*S*, 8*S*, and 14*R*. Thus, the structure of **9** was elucidated as 14 $\beta$ -methoxy-13,14-dihydronorsecurinine.

**10** was obtained as yellow oil. The molecular formula of **10** was established as C<sub>13</sub>H<sub>17</sub>NO<sub>3</sub> by the HR-ESI-MS at *m/z*: 258.1093 [M+Na]<sup>+</sup> (Calcd for C<sub>13</sub>H<sub>17</sub>NO<sub>3</sub>Na, 258.1101), which had the same molecular formula as **9**. The IR spectrum showed the presence of  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone ring (1756, 1642 cm<sup>-1</sup>). The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of **10** were also very similar to those of **9**, suggesting that **10** was the isomeride of **9**. In the NOESY spectrum of **10**, the correlations between H-7b and H-13b, as well as between H-13b and 14-OMe (Fig. 3) were observed, indicating that the configuration at C-14 of **10** was opposite to that of **9**. The absolute configuration of **10** was finally determined as 2*R*, 6*S*, 8*S*, and 14*S* by comparing the CD spectrum of **10** with those of **7**, **8** and **9**. Therefore, the structure of **10** was elucidated as 14 $\alpha$ -methoxy-13,14-dihydronorsecurinine.

#### Experimental

**General Experimental Procedures** Optical rotations were determined on a JASCO P-1020 polarimeter. UV spectra were obtained on a JASCO V-550 UV/VIS spectrophotometer. CD spectra were measured on a JASCO J-720 spectrometer. IR spectra were obtained on a JASCO FT/IR-480 plus spectrometer. <sup>1</sup>H-, <sup>13</sup>C- and 2D-NMR spectra were determined on a Bruker-



AV-400 spectrometer in  $\text{CDCl}_3$ . ESI-MS spectra were run on a HP-1100 HPLC/EST spectrometer. HR-ESI-MS spectra were obtained on a Biosystems Mariner™ 5140 spectrometer. For column chromatography, silica gel (200–300 mesh, Qingdao Marine Chemical Factory, P. R. China), Sephadex LH-20 (Pharmacia) and ODS (YMC) were used. TLC analyses were carried out using precoated silica gel GF254 plates (Qingdao Marine Chemical Factory, P. R. China). HPLC separations were performed on a COSMOSIL  $\text{C}_{18}$  preparative column (5  $\mu\text{m}$ ,  $20 \times 250 \text{ mm}$ ).

**Plant Material** The twigs and leaves of *F. leucopyra* were collected in Nujiang country, Yunnan province of China, in September of 2006. The plant was authenticated by Prof. Guang-xiong Zhou of Jinan University. A voucher specimen (No. 060914) was deposited in the Institute of Traditional Chinese Medicine and Natural Products, Jinan University, Guangzhou, P. R. China.

**Extraction and Isolation** The air-dried twigs and leaves of *F. leucopyra* (8.5 kg) were extracted with 95% EtOH and the solution was evaporated *in vacuo* to get a residue (1045 g). The residue was dissolved in  $\text{H}_2\text{O}$  to form a suspension and then adjusted to pH 6 with 5% HCl. The acidic suspension was partitioned with  $\text{CHCl}_3$  to remove the neutral components. The aqueous phase was basified with 2%  $\text{NH}_3 \cdot \text{H}_2\text{O}$  to pH 8 and then extracted with  $\text{CHCl}_3$  to obtain a total alkaloid part (42 g), which was subjected to silica gel column chromatography (CC) ( $\text{CHCl}_3$ – $\text{CH}_3\text{OH}$ , 100:0→0:100) to give 10 fractions (1–10). Fraction 3 (3.86 g) was subjected to silica gel CC (*n*-hexane–EtOAc, 10:1→1:1) to afford 8 subfractions (3-1–3-8). Subfraction 3-4 (346 mg) was purified by HPLC [ $\text{CH}_3\text{OH}$ – $\text{H}_2\text{O}$ –( $\text{CH}_3\text{CH}_2$ ) $_3\text{N}$ , 85:15:0.02] to afford **8** (12 mg), **9** (11 mg) and **10** (9 mg). Fraction 4 (2.76 g) was purified by Sephadex LH-20 column ( $\text{CH}_3\text{OH}$ ) and silica gel CC ( $\text{CHCl}_3$ – $\text{CH}_3\text{OH}$ , 100:0→80:20) to yield **4** (21 mg) and **5** (12 mg). Fraction 5 (3.34 g) was subjected to reversed-phase silica gel (ODS) CC ( $\text{CH}_3\text{OH}$ – $\text{H}_2\text{O}$ , 60:40→90:10) to afford 9 subfractions (5-1–5-9). Subfraction 5-3 (569 mg) was purified by preparative HPLC [ $\text{CH}_3\text{OH}$ – $\text{H}_2\text{O}$ –( $\text{CH}_3\text{CH}_2$ ) $_3\text{N}$ , 70:30:0.02] to obtain **1** (22 mg), **3** (19 mg) and **7** (15 mg), respectively. Subfraction 5-4 (369 mg) was purified by preparative HPLC [ $\text{CH}_3\text{OH}$ – $\text{H}_2\text{O}$ –( $\text{CH}_3\text{CH}_2$ ) $_3\text{N}$ , 75:25:0.02] to obtain **2** (22 mg) and **6** (19 mg), respectively.

**4 $\alpha$ -Hydroxyallosecurinine (1):** Yellow oil;  $[\alpha]_{\text{D}}^{20}$  –102 ( $c=0.07$ , MeOH); UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 218 (1.65), 257 (1.97) nm; CD (MeOH)  $\lambda_{\text{max}}$  ( $\Delta\epsilon$ ) 216 (–3.34), 312 (–17.37) nm; IR (KBr)  $\nu_{\text{max}}$ : 3436, 2948, 1746, 1632, 1464, 1286  $\text{cm}^{-1}$ ;  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data see Table 1; ESI-MS  $m/z$ : 234  $[\text{M}+\text{H}]^+$ ; HR-ESI-MS  $m/z$ : 234.1127  $[\text{M}+\text{H}]^+$  (Calcd for  $\text{C}_{13}\text{H}_{16}\text{NO}_3$ , 234.1125).

**4 $\alpha$ -Hydroxy-15 $\alpha$ -methoxy-14,15-dihydroallosecurinine (3):** Yellow oil;  $[\alpha]_{\text{D}}^{20}$  –32 ( $c=0.12$ , MeOH); UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 212 (1.85) nm; CD (MeOH)  $\lambda_{\text{max}}$  ( $\Delta\epsilon$ ) 226 (+2.87), 277 (+0.76) nm; IR (KBr)  $\nu_{\text{max}}$ : 3447, 2936, 1756, 1643, 1438, 1268  $\text{cm}^{-1}$ ;  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data see Table 1; ESI-MS  $m/z$ : 266  $[\text{M}+\text{H}]^+$ ; HR-ESI-MS  $m/z$ : 266.1387  $[\text{M}+\text{H}]^+$  (Calcd for  $\text{C}_{14}\text{H}_{20}\text{NO}_4$ , 266.1387).

**4 $\alpha$ -Methoxy-15 $\alpha$ -methoxy-14,15-dihydrosecurinine (5):** Yellow oil;  $[\alpha]_{\text{D}}^{20}$  +37 ( $c=0.11$ , MeOH); UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 220 (1.89) nm; CD (MeOH)  $\lambda_{\text{max}}$  ( $\Delta\epsilon$ ) 218 (–1.39), 240 (+2.62), 279 (+0.77), 334 (–0.50) nm; IR (KBr)  $\nu_{\text{max}}$ : 3035, 2928, 1758, 1644, 1453, 1204  $\text{cm}^{-1}$ ;  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data see Table 1; ESI-MS  $m/z$ : 280  $[\text{M}+\text{H}]^+$ ; HR-ESI-MS  $m/z$ : 280.1540  $[\text{M}+\text{H}]^+$  (Calcd for  $\text{C}_{15}\text{H}_{22}\text{NO}_4$ , 280.1543).

**14 $\beta$ -Hydroxy-13,14-dihydronorsecurinine (7):** Yellow oil;  $[\alpha]_{\text{D}}^{20}$  –45 ( $c=0.13$ , MeOH); UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 216 (2.15) nm; CD (MeOH)  $\lambda_{\text{max}}$  ( $\Delta\epsilon$ ) 219 (–2.76), 238 (+1.74), 265 (–0.85) nm; IR (KBr)  $\nu_{\text{max}}$ : 3442, 2929, 1755, 1631, 1460, 1274  $\text{cm}^{-1}$ ;  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR see Table 2; ESI-MS  $m/z$ : 222  $[\text{M}+\text{H}]^+$ ; HR-ESI-MS  $m/z$ : 222.1126  $[\text{M}+\text{H}]^+$  (Calcd for  $\text{C}_{12}\text{H}_{16}\text{NO}_3$ , 222.1125).

**14 $\beta$ -Methoxy-13,14-dihydronorsecurinine (9):** Yellow oil;  $[\alpha]_{\text{D}}^{20}$  –44 ( $c=0.16$ , MeOH); UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 216 (2.21) nm; CD (MeOH)  $\lambda_{\text{max}}$  ( $\Delta\epsilon$ ) 208 (–2.56), 234 (+2.25), and 284 (–0.62) nm; IR (KBr)  $\nu_{\text{max}}$ : 3135, 2924, 1755, 1649, 1451, 1383  $\text{cm}^{-1}$ ;  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data see Table 2; ESI-MS  $m/z$ : 236  $[\text{M}+\text{H}]^+$ ; HR-ESI-MS  $m/z$ : 236.1286  $[\text{M}+\text{H}]^+$  (Calcd for  $\text{C}_{13}\text{H}_{18}\text{NO}_3$ , 236.1281).

**14 $\alpha$ -Methoxy-13,14-dihydronorsecurinine (10):** Yellow oil;  $[\alpha]_{\text{D}}^{20}$  +27 ( $c=0.10$ , MeOH); UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 217 (2.05) nm; CD (MeOH)  $\lambda_{\text{max}}$  ( $\Delta\epsilon$ ) 213 (–1.71), 235 (+2.69), 263 (–1.75) nm; IR (KBr)  $\nu_{\text{max}}$ : 2936, 1756, 1642, 1460, 1189  $\text{cm}^{-1}$ ;  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data, see Table 2; ESI-MS  $m/z$ : 258  $[\text{M}+\text{Na}]^+$ ; HR-ESI-MS  $m/z$ : 258.1093  $[\text{M}+\text{Na}]^+$  (Calcd for

$\text{C}_{13}\text{H}_{17}\text{NO}_3\text{Na}$ , 258.1101).

**Preparation of MTPA Esters of 1** **1** (4.5 mg) was dissolved in 0.5 ml of dried pyridine and treated with (*R*)-(–)- $\alpha$ -methoxy- $\alpha$ -(trifluoromethyl) phenylacetyl chloride (10  $\mu\text{l}$ ). The reaction product was stirred at room temperature overnight and then dried *in vacuo*. The reaction mixture was poured into water (5 ml) and extracted with EtOAc (5 ml). The EtOAc extract was purified by silica gel column chromatography [*n*-hexane–EtOAc (70:30)] to yield (*S*)-MTPA ester (**1a**, 3.2 mg). (*R*)-MTPA ester (**1b**, 2.7 mg) was obtained using the same method by treatment of **1** (4.3 mg) with (*S*)-(+)– $\alpha$ -methoxy- $\alpha$ -(trifluoromethyl) phenylacetyl chloride.

(*S*)-MTPA Ester (**1a**):  $^1\text{H}$ -NMR ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 1.63 (1H, m, H-3b), 1.96 (1H, d,  $J=14.8$  Hz, H-3b), 2.10 (1H, m, H-5b), 2.20 (1H, d,  $J=11.6$  Hz, H-8b), 2.57 (1H, dd,  $J=16.0, 9.6$  Hz, H-5a), 2.81 (1H, dd,  $J=14.0, 12.8$  Hz, H-6b), 3.12 (1H, br d,  $J=11.6$  Hz, H-8a), 3.55 (1H, overlapped, H-6a), 3.60 (3H, s, OMe of MTPA), 4.12 (1H, dd,  $J=14.0, 3.2$  Hz, H-7), 4.81 (1H, br s, H-2), 5.39 (1H, m, H-4), 6.06 (1H, br s, H-12), 6.70 (1H, dd,  $J=8.8, 6.4$  Hz, H-15), 7.01 (1H, d,  $J=8.8$  Hz, H-14), 7.27 and 7.45 (phenyl protons of MTPA); ESI-MS  $m/z$ : 450  $[\text{M}+\text{H}]^+$ ; HR-ESI-MS  $m/z$ : 450.1519  $[\text{M}+\text{H}]^+$  (Calcd for  $\text{C}_{23}\text{H}_{23}\text{F}_3\text{NO}_5$ , 450.1523).

(*R*)-MTPA Ester (**1b**):  $^1\text{H}$ -NMR ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 1.56 (1H, m, H-3b), 1.86 (1H, dd,  $J=14.8, 2.0$  Hz, H-3b), 2.18 (1H, overlapped, H-5b), 2.18 (1H, d,  $J=11.6$  Hz, H-8b), 2.65 (1H, dd,  $J=16.0, 9.6$  Hz, H-5a), 2.82 (1H, dd,  $J=14.0, 12.8$  Hz, H-6b), 3.09 (1H, dd,  $J=11.6, 4.4$  Hz, H-8a), 3.57 (1H, overlapped, H-6a), 3.60 (3H, s, OMe of MTPA), 4.08 (1H, dd,  $J=14.0, 3.6$  Hz, H-7), 4.81 (1H, t,  $J=4.8$  Hz, H-2), 5.35 (1H, m, H-4), 6.04 (1H, br s, H-12), 6.70 (1H, dd,  $J=8.8, 6.4$  Hz, H-15), 7.01 (1H, d,  $J=9.0$  Hz, H-14), 7.27 and 7.45 (phenyl protons of MTPA). ESI-MS  $m/z$ : 450  $[\text{M}+\text{H}]^+$ ; HR-ESI-MS  $m/z$ : 450.1529  $[\text{M}+\text{H}]^+$  (Calcd for  $\text{C}_{23}\text{H}_{23}\text{F}_3\text{NO}_5$ , 450.1523).

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