2-Aryl-1,4-naphthoquinone-1-oxime Methyl Ethers: Their Cytotoxic Activity

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Preliminary examination for the structure-activity relationship of quinone monooxime derivatives on cytotoxicity against HeLa S3 cell and further trials using eight different cell lines suggested that 2-aryl-6,7-methylenedioxy-1,4-naphthoquinone-1-oxime methyl ethers, carrying 2-methoxy-4,5-methylenedioxyphenyl, 7-methoxy-2-methylbenzofuran-4-yl, and 2-methoxycarbonyl-3,4-dimethoxyphenyl as the 2-aryl substituent, were potential candidates for anti-cancer drugs.

Key words 2-aryl-1,4-naphthoquinone-1-oxime methyl ether; benzophenanthridine; cytotoxicity; structure-activity relationship

Fully-aromatized benzo [c] phenanthridine alkaloids 1 with an isoquinoline core skeleton are structurally classified into O_4 -type (R¹, R², R³, R⁴ or R¹, R², R⁴, R⁵=OR), O₅-type (R¹, R^2 , R^3 , R^4 , $R^6=OR$), and O_6 -type (R^1 , R^2 , R^3 , R^4 , R^6 , R^7 =OR) bases dependent upon the number of oxygen functions in their molecules.¹⁾ They have been attracted much attention due to their valuable pharmacological activities,¹⁻³⁾ especially the anti-tumor activity of O₄-type alkaloids such as nitidine⁴ (1a), fagaronine⁴ (1b), sanguinarine⁵ (1c), and NK109⁶) (1d). For their structure–activity relationship (SAR) on anti-tumor activity, we have established the practical synthetic methods through 2-aryl-1-tetralones as key intermediates under the modified Robinson's route⁷⁾ for the O_4 - and O_5 -type benzo[c]phenanthridine alkaloids. On the other hand, macarpine (1e), a O_6 -type alkaloid, has been prepared by an alternative route through 2-(2-methoxy-4,5-methylenedioxyphenyl)-6,7-methylenedioxy-1,4-naphthoquinone-1oxime methyl ether (2) as a key synthetic precursor⁸ (Fig. 1). In screening for the discovery of potential anti-tumor active seed compounds from natural and unnatural sources, we found that the 2-aryl-1,4-naphthoquinone-1-oxime methyl ether 2 shows strong cytotoxic activity. In this paper we report preliminary results on the SAR of the naphthoquinone monooximes and the related compounds for cytotoxic activ-



Fig. 1. Structures of Benzo[c]phenanthridine Alkaloids 1a-e and Naphthoquinone Monooxime Ether 2

ity against HeLa S3 cell and more precise examination on the potent growth inhibitory effects of the selected 2-aryl-1,4naphthoquinone-1-oxime methyl ethers 2 and 7 using eight cancer cell lines, in which effective inhibition against HeLa and MCF-7 cells was observed.

Results and Discussion

The cytotoxic activity of 2-(2-methoxy-4,5-methylenedioxyphenyl)-6,7-methylenedioxy-1,4-naphthoquinone-1oxime methyl ether (2) was evaluated using HeLa S3 cell by methylene blue staining method and the IC₅₀ was estimated to be less than $0.1 \,\mu$ g/ml using Litchfield and Wilcoxon Thus, the 2-aryl-1,4-naphthoquinone-1-oxime method. methyl ether 2 was decided to be used as a positive control for further SAR examination of the naphthoquinone monooximes and the related compounds and, firstly, substituent variation of skeletal alkoxy functions of 2 was focused on. We had reported the preparation methods of 1,4naphthoquinone-1-oxime methyl ethers by either basic nitrosation [isoamyl nitrite in dimethylformamide (DMF) in the presence of potassium carbonate (K₂CO₃)] of phenol substrate followed by methylation with dimethyl sulfate9) or heating 2-aryl-1-tetralone oxime methyl ethers with 5,6dichloro-2,3-dicycano-1,4-benzoquinone (DDQ) in acetic acid.¹⁰⁾ Three known 2-aryl-1,4-naphthoquinone-1-oxime methyl ethers 3-5 with different alkoxy functions in the naphthoquinone skeleton and/or in the 2-aryl substituent, which had been prepared by application of the latter DDQ oxidation method, were subjected to evaluation of cytotoxic activity using the same protocol for a positive control 2 (Table 1). Displacement of the methylenedioxy moieties either in the naphthoquinone skeleton (see 3) or in the 2-aryl substituent (see 4) with dimethoxy ones led to the reduction of activity (entries 2 and 3 in Table 1). No activity was observed when both methylenedioxy moieties in 2 were converted to methoxy groups like 5 (entry 4 in Table 1). 2-(3,4-Dimethoxyphenyl)-6,7-methylenedioxy-1,4-naphthoquinone-1-oxime methyl ether (6), carrying the same substitution pattern that an anti-tumor active benzo[c]phenanthridine alkaloid, nitidine⁴⁾ (1a), was newly prepared according to the

Table 1. Cytotoxic Activity of 2-Aryl-1,4-naphthoquinone-1-oxime Methyl Ethers 2-8



Entry	Compd.	Ar	\mathbb{R}^1	R ²	Results ^{a)}
1	2 ⁸⁾	2-Methoxy-4,5-methylenedioxyphenyl	CH ₂		(+)
2	3 ¹⁰⁾	2-Methoxy-4,5-methylenedioxyphenyl	Me	Me	(\pm)
3	4 ¹⁰⁾	2,4,5-Trimethoxyphenyl	CH,		(\pm)
4	5 ¹⁰⁾	2,4,5-Trimethoxyphenyl	Me	Me	(-)
5	6	3,4-Dimethoxyphenyl	CH ₂		(\pm)
6	7 ⁹⁾	7-Methoxy-2-methylbenzofuran-4-yl	CH ₂		(+)
7	8 ¹¹⁾	2-Methoxycarbonyl-3,4-dimethoxyphenyl	CH_2		(+)

a) Evaluated by methylene blue staining method using HeLa S3 cell. IC₅₀ (μ g/ml) was estimated using Litchfield and Wilcoxon method and the following symbols are used as criteria: (+)=<0.1 μ g/ml; (±)=0.1-1.0 μ g/ml; (-)=>1.0 μ g/ml.

Table 2.Cytotoxic Activity of Other 1,4-Quinone OximesI.1,4-Naphthoquinone-1-oxime Methyl Ethers 9 and 10



Entry	Compd.	\mathbb{R}^1	R ²	R ³	Results ^{a)}
I-1	9 ⁹⁾	7-Methoxy-2-methylbenzofuran-4-yl	OCH ₂ O	Н	(-)
I-2	10 ⁹⁾	H	H		(±)

II. 1,4-Benzoquinone-1-oximes 11-15

$R^{1} \xrightarrow{O} R^{2}$ $R^{1} \xrightarrow{V_{n}} H$ OH					
Entry	Compd.	\mathbb{R}^1	\mathbb{R}^2	Results ^{a)}	
II-1	11 ⁹⁾	Me	CO ₂ Me	(\pm)	
II-2	12 ⁹⁾	Н	CO_2Me	(-)	
II-3	13 ⁹⁾	Н	CHO	(±)	
II-4	14 ⁹⁾	OMe	CHO	(-)	
II-5	15 ⁹⁾	OMe	Н	(-)	

III. Heterocyclic 1,4-Quinone Oximes 16-19 and 2-Nitroso-1-naphthol 20



Entry	Compd.	Results ^{<i>a</i>})
III-1	2-Methylbenzofuran-4,7-quinone-4-oxime ⁹⁾ 16	(\pm)
III-2	Coumarin-5,8-quinone-5-oxime ⁹⁾ 17	(-)
III-3	Quinoline-5,8-quinone-5-oxime ⁹⁾ 18	(\pm)
III-4	Quinoline-5,8-quinone-5-oxime methyl ether ⁹⁾ 19	(\pm)
III-5	2-Nitroso-1-naphthol ⁹⁾ 20	(\pm)

a) See footnote *a*) in Table 1.

DDQ oxidation method; however, its activity was estimated to be moderate (entry 5 in Table 1). On the other hand, alternative 1,4-naphthoquinone-1-oxime methyl ethers with 7methoxy-2-methylbenzofuran-4-yl⁹⁾ 7 or 2-methoxycarbonyl-3,4-dimethoxyphenyl¹¹⁾ 8 as the 2-aryl pendant, which had been prepared by the former basic nitrosation-methylation method, were interestingly proven to maintain strong cytotoxic activity (entries 6 and 7 in Table 1).

A variety of quinone monooxime derivatives had also been prepared from the corresponding phenolic compounds by application of the basic nitrosation method.⁹⁾ Thus, selected samples in our hands, including 3-(7-methoxy-2-methylbenzofuran-4-yl)-6,7-methylenedioxy-1,4-naphthoquinone-1oxime methyl ether (9), a positional isomer of aryl substituent in the cytotoxic 2-(7-methoxy-2-methylbenzofuran-4-yl) derivative 7, were subjected to evaluation in order to discuss on the skeletal requirement of 2-aryl-1,4-naphthoquinone-1-oxime methyl ether core to cytotoxic activity. The results of 1,4-naphthoquinone-1-oxime ethers, 1,4-benzoquinone-1-oximes, and heterocyclic 1,4-quinone oximes are shown in I, II, and III in Table 2, respectively, and all quinone monooxime derivatives examined were found to be negative.

These experimental facts obtained in Tables 1 and 2 suggest that the presence of 6,7-methylenedioxy function in the 2-aryl-1,4-naphthoquinone-1-oxime skeleton is crucial for the positive cytotoxic activity, in spite of substituent tolerance in the 2-aryl group. It should be noted here that no distinct change in conformation of the 2-aryl-1,4-naphthoquinone-1-oxime skeleton was observed in crystal structures between an active **2** and an inactive pentamethoxy-substituted derivatives **5**.¹²

Next, our attention turned to the structural modification of the oxime moiety in the 2-aryl-1,4-naphthoguinone-1-oxime methyl ether for the examination of the contribution of the ether unit in oxime methyl ether function to positive cytotoxicity. In general, 2-aryl-1,4-naphthoguinone-1-oxime methyl ether derivatives are hard to dissolve in the test solution for evaluation. Thus, introduction of polar functional group to a positive control 2 was targeted for the increase of solubility, and the basic nitrosation method was applied to the preparation of oxime-modified derivatives from 3-(2-methoxy-4,5methylenedioxyphenyl)-6,7-methylenedioxy-1-naphthol 21 (Chart 1). Naphthoquinone oxime derivatives 22 and 23 with acetic acid residue were prepared by alkylation with bromoacetate and by alkaline hydrolysis of the resulting oxime acetate 22, respectively, after the basic nitrosation of 21. The aminoethyl-inserted ether 24 was given by alkylation using dimethylaminoethyl chloride in place of bromoacetate in 22. The known iminoquinone⁸⁾ 25, in addition to the three derivatives 22-24 prepared above, were subjected to the evaluation of cytotoxic activity under the same condition that on a positive control 2 (Table 3). However, all of the four N-modified analogues¹³⁾ 22–25 showed no activity, indicating that the oxime methyl ether function in the 1,4-naphthoquinone-1-oxime skeleton plays an important role for the positive activity.

Finally, based on the above results, cell prolification of a positive control **2** and an alternative potential **7** on a concentration-dependent manner was further examined using eight cancer cell lines, in which expected activity was observed except for H28 cell (Table 4). In particular, these naphtho-



Chart 1. Preparation of *N*-Modified 2-(2-Methoxy-4,5-methylenedioxyphenyl)-6,7-methylenedioxy-1,4-naphthoquinone-1-oxime Ether Derivatives 22—25

Table 3. Cytotoxic Activity of 2-(2-Methoxy-4,5-methylenedioxyphenyl)-6,7-methylenedioxy-1,4-naphthoquinone-1-oxime Methyl Ether (2) and Its *N*-Modified Derivatives **22**—**25**

Entry	Compd.	Results ^{a)}
1	2 ⁸⁾	(+)
2	22	(-)
3	23	(-)
4	24	(-)
5	25 ⁸⁾	(-)

a) See footnote *a*) in Table 1.

Table 4. IC_{50} Values of Two Naphthoquinone Monooximes **2** and **7** in Different Cancer Cell Lines^{*a*})

Call	Derivation —	IC ₅₀ (µм)±S.D.		
Cell		2	7	
HeLa Caki-1 786-O A549 MCF-7 H28 H2052	Ovarian cancer Renal cancer Lung cancer Breast cancer Mesothelioma Mesothelioma	$\begin{array}{c} 0.10 \pm 0.02 \\ 0.60 \pm 0.14 \\ 0.28 \pm 0.07 \\ 0.43 \pm 0.04 \\ 0.20 \pm 0.07 \\ 5.78 \pm 10.50 \\ 0.54 \pm 0.25 \end{array}$	$\begin{array}{c} 0.23 \pm 0.04 \\ 0.91 \pm 0.06 \\ 0.71 \pm 0.09 \\ 1.06 \pm 0.07 \\ 0.51 \pm 0.09 \\ 6.52 \pm 3.39 \\ 0.47 \pm 0.15 \end{array}$	
MSTO-211H	Mesothelioma	0.38 ± 0.25	1.05 ± 0.79	

a) Cells were treated with $\mathbf{2}$ and $\mathbf{7}$ for 48 h. Data are from 3 independent experiments.

quinone oximes effectively attacked HeLa (IC₅₀ 0.10 μ M on **2** and 0.23 μ M on **7**, respectively) and MCF-7 cells (IC₅₀ 0.20 μ M on **2** and 0.51 μ M on **7**, respectively).

In conclusion, it was found that 2-aryl-6,7-methylenedioxy-1,4-naphthoquinone-1-oxime methyl ethers showed cytotoxic activity in a wide range of cancer cell lines, especially HeLa and MCF-7 cells. These facts suggest that the quinone monooxime core could serve as a potential scafford in screening for the discovery of bioactive small molecules because of known antiviral activity.⁹ Mechanistic approaches to the cytotoxicity of these 2-aryl-1,4-naphthoquinone-1oxime methyl ethers, in addition to further modification of the substituent in the 2-aryl moiety focusing on increasing hydrophilicity for extension to *in vivo* trials due to the hard solubility of the 1,4-naphthoquinone-1-oxime ethers to the test solution, are under investigation in our laboratory.

Experimental

General IR spectra were recorded on a JASCO IR-300E spectrophotometer (ATR). High resolution (HR)-FAB-MS and HR-electrospray ionization (ESI)-MS spectra were measured by JEOL JMS-HX 110A and JMS- T100LP, and Thermo Scientific Exactive Bentitop Orbitrap spectrometers, respectively. ¹H- (400 MHz) and ¹³C-NMR (100 MHz) spectra were obtained on JEOL JNM ECP 400 in CDCl₃ unless otherwise stated. For TLC was used SiO₂ 60 F_{254} , 0.25 mm (Merck) and for column chromatography SiO₂ 60 (63—210 mm) (Kanto-Cica) or FL100D SiO₂ (Fuji Silysia Chemical Ltd., Japan).

2-(3,4-Dimethoxyphenyl)-6,7-methylenedioxy-1,4-naphthoquinone-1oxime Methyl Ether (6) According to the reported method for the DDQ oxidation,¹⁰⁾ a mixture of 2-(3,4-dimethoxyphenyl)-6,7-methylenedioxy-1tetralone oxime methyl ether (0.11 g, 3.1×10^{-4} mol), easily derived from the corresponding 1-tetralone¹⁴⁾ by conventional treatment with hydroxylamine methyl ether, and DDQ (0.30 g, 1.3×10^{-3} mol) in benzene (3 ml) containing acetic acid (0.2 ml) was heated at 80 °C for 22 h. After worked up, purification of a crude product by SiO₂ column chromatography (ethyl acetate: hexane=1:3) provided the naphthoquinone-1-oxime ether 6 (0.009 g, 8%)as yellow prisms, mp 248—250 °C. IR cm⁻¹: 1641. ¹H-NMR δ: 3.92 (3H, s, OMe), 3.94 (3H, s, OMe), 4.13 (3H, s, OMe), 6.13 (2H, s, OCH2O), 6.63 (1H, s, C3-H), 6.91 (1H, d, J=8.4 Hz, C5'-H), 7.06 (1H, d, J=1.6 Hz, C2'-H), 7.10 (1H, dd, J=8.4 Hz, 1.6 Hz, C6'-H), 7.69 (1H, s, C5-H), 8.36 (1H, s, C8-H). ¹³C-NMR δ: 55.9, 56.0, 64.5, 102.2, 106.0, 110.2, 110.4, 113.3, 122.8, 124.6, 127.5, 128.7, 129.7, 145.6, 148.1, 149.3, 149.8, 151.2, 151.7, 183.4. HR-FAB-MS m/z: 368.1145 (Calcd for C₂₀H₁₈NO₆: 368.1134).

2-(2-Methoxy-4,5-methylenedioxyphenyl)-6,7-methylenedioxy-1,4naphthoquinone-1-oxime Ethoxycarbonylmethyl Ether (22) According to the reported method for basic nitrosation,^{8,9)} isoamyl nitrite (0.02 ml, d=0.87, 14×10^{-4} mol) was added to a mixture of the naphthol **21** (0.030 g, 8.0×10^{-5} mol) and K₂CO₃ (0.060 g, 4.0×10^{-4} mol) in DMF (0.3 ml) at rt and then the whole was stirred at rt for 1 h. After addition of ethyl bromoacetate (0.012 ml, d=1.51, 11×10^{-4} mol) under ice-cooling, the resulting mixture was stirred at rt for 1 h, quenched with water, and extracted with ethyl acetate. The organic solution was washed with water and brine, dried (K₂CO₃), and evaporated. Purification of the residual red brown oil by preparative TLC (ethyl acetate: hexane=1:2, Rf=0.30-0.38) afforded orange prisms (0.030 g, 76%), mp 125-129 °C, which were triturated with hexane. IR cm⁻¹: 1768, 1639. ¹H-NMR δ : 1.28 (3H, t, J=7.1 Hz, CH₂CH₃), 3.70 (3H, s, OMe), 4.22 (2H, q, J=7.1 Hz, OCH_2CH_3), 4.74 (2H, s, OCH2CO), 5.97 (2H, s, OCH2O), 6.11 (2H, s, OCH2O), 6.54 (2H, s, C5'-, C3-H), 6.72 (1H, s, C2-H), 7.67 (1H, s, C5-H), 8.46 (1H, s, C8-H). ¹³C-NMR δ: 14.1, 56.6, 61.2, 72.7, 94.8, 101.4, 102.1, 106.0, 110.2, 110.6, 118.6, 124.5, 128.7, 129.7, 140.8, 147.0, 148.7, 149.4, 149.6, 151.3, 152.5, 168.6, 183.6. HR-ESI-MS m/z: 476.0927 (Calcd for C23H10NNaO9: 476.0958).

2-(2-Methoxy-4,5-methylenedioxyphenyl)-6,7-methylenedioxy-1,4-naphthoquinone-1-oxime Hydroxycarbonylmethyl Ether (23) A mixture of the naphthoquinone ester **22** (0.025 g, 5.5×10^{-5} mol) and 2 mol/l aq NaOH (0.5 ml) in EtOH (1 ml) was heated at 50 °C for 10 min, quenched with water, and extracted with ethyl acetate. The aqueous solution was acidified with 5% HCl solution and extracted with ethyl acetate. The organic solution was dried (MgSO₄) and evaporated to give red prisms (0.023 g, quant.), mp 202—205 °C, which were triturated with hexane. IR cm⁻¹: 1758, 1622. ¹H-NMR δ : 3.70 (3H, s, OMe), 4.79 (2H, s, OCH₂CO), 5.98 (2H, s, OCH₂O), 6.13 (2H, s, OCH₂O), 6.55 (1H, s, C5'- or C3-H), 6.56 (1H, s, C5'- or C3-H), 6.72 (1H, s, C2-H), 7.68 (1H, s, C5-H), 8.36 (1H, s, C8-H). ¹³C-NMR (CD₃OD) δ : 57.0, 73.1, 95.8, 102.4, 103.6, 105.9, 110.6, 111.1, 119.6, 125.3, 129.4, 129.9, 141.7, 147.4, 149.8, 150.6, 151.0, 152.3, 153.7, 170.0, 183.3. HR-ESI-MS *m*/*z*: 448.0663 (Calcd for C₂₁H₁₅NNaO₉: 448.0645).

2-(2-Methoxy-4,5-methylenedioxyphenyl)-6,7-methylenedioxy-1,4naphthoquinone-1-oxime 2-Dimethylaminoethyl Ether (24) According to the reported method for basic nitrosation,^{8,9)} isoamyl nitrite (0.04 ml, $d=0.87, 3.0\times10^{-4}$ mol) was added to a mixture of the naphthol **21** (0.030 g, 8.9×10^{-5} mol) and K₂CO₃ (0.10 g, 7.2×10^{-4} mol) in DMF (1.6 ml) under ice-cooling and then the whole was stirred at rt for 2 h. After addition of 2dimethylaminoethyl chloride hydrochloride (0.017 g, 1.2×10^{-4} mol) under ice-cooling, the resulting mixture was stirred at rt for 15 h, quenched with ice-water, and extracted with ethyl acetate. The organic solution was washed with water and brine, dried (K₂CO₂), and evaporated. Purification of the residual dark brown gummy product by SiO₂ column chromatography (ethyl acetate: hexane: methanol=10:10:1) afforded yellow prisms (0.023 g, 59%), mp 109—112 °C, which were triturated with hexane. IR cm⁻¹: 1639. ¹H-NMR δ : 2.28 (6H, s, NMe₂), 2.65 (2H, t, J=5.9 Hz, CH₂CH₂N), 3.71 (3H, s, OMe), 4.33 (2H, t, J=5.9 Hz, OCH₂CH₂), 5.98 (2H, s, OCH₂O), 6.11 (2H, s, OCH₂O), 6.52 (1H, s, C3-H), 6.56 (1H, s, C5'-H), 6.72 (1H, s, C2'-H), 7.68 (1H, s, C5-H), 8.40 (1H, s, C8-H). ¹³C-NMR δ : 45.8, 56.8, 58.0,

74.9, 95.0, 101.6, 102.3, 106.2, 110.3, 119.4, 124.8, 128.8, 129.4, 141.0, 145.7, 148.7, 149.4, 150.4, 151.4, 152.5, 183.8. HR-ESI-MS m/z: 439.1526 (Calcd for C₂₃H₂₃N₂O₇: 439.1505).

Preliminary Screening Using HeLa S3 The sample was dissolved in dimethyl sulfoxide (DMSO) and then diluted with saline at the final concentrations of 30, 10, 3, 1, 0.3, 0.1, 0.03, 0.01 µg/ml. HeLa S3 cell containing 1×10^4 cells/ml were inoculated into 96-well microplates with 0.2 ml/well and incubated at 37 °C for 24 h under 5% CO₂ atmosphere. After addition of the test solution (0.1 µl) cells were cultured for 72 h. Following cells were fixed with methanol and cells were stained with 0.05% methyleneblue in 10 mmol/l Tris–HCl (pH 8.5) (0.1 ml/well) at rt for 30 min. After removal of the staining solution using aspirator the residue was washed with water three times and the stained dye was extracted with 3% HCl (0.2 ml/well). The inhibition ratio was estimated by measurement of absorbance of each well using a wave length of 660 nm and IC₅₀ (µg/ml) was calculated by Litchfield and Wilcoxon method.

Cell Cytotoxicity Assay All cell lines were provided by Tomohiro Yano (Graduate School of Life Sciences, Toyo University, Japan), unless otherwise indicated. Caki-1 (human kidney cancer cell line from ATCC) was grown in McCoy's 5A (Wako). HeLa (human ovarian cancer cell line from RIKEN), 786-O (human kidney cancer cell line), and A549 (human lung cancer cell line) were grown in Dulbecco's Modified Eagle's Medium (DMEM) (Wako). MCF-7 (human breast cancer cell line), H2052 (human malignant mesothelioma cell line), and MSTO-211H (human malignant mesothelioma cell line) were grown in RPMI 1640 (Wako). H28 (human malignant mesothelioma cell line) was grown in RPMI 1640 supplemented with 0.01 M N-(2-hydroxyethyl)piperazine-N'-2-(ethanesulfonic acid) (HEPES) buffer solution (GIBCO BRL, Grand Island, NY, U.S.A.), 1 mM sodium pyruvate (GIBCO BRL), and 9.0 g/l glucose. All cells were cultured with 10% fetal bovine serum (FBS) (EQUITECH-BIO, Kerrville, TX, U.S.A.), 1.0 units/ml penicillin (GIBCO BRL), and 2.0 mg/ml streptomycin (GIBCO BRL) in atmosphere at 37 °C with 5% CO₂.

A total of 3.0×10^3 cells/well was seeded in a 96-well plate. After 24 h incubation, an optimum concentration gradient of the 2-aryl-1,4-naphthoquinone-1-oxime methyl ethers **2** or **7** was added to each well, followed by culturing for 48 h. Finally, to assess the sensitivity of the cells to these compounds, cell viability was assessed using the Proliferation Reagent 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) (DOJINDO, Kumamoto, Japan) according to the manufacturer's instructions. Control cells were treated with 0.1% DMSO, which is the vehicle for **2** and **7**. The inhibition ratio was estimated by measurement of absorbance of each well using a wave length of 570 nm and IC₅₀ (µg/ml) was calculated by GraphPad PRISM 5J (Graphpad Software, Inc., San Diego, CA, U.S.A.).

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