

## A Mass-spectrometric Study of Biosynthesis: Conversion of Deutero-*m*-cresol into Patulin

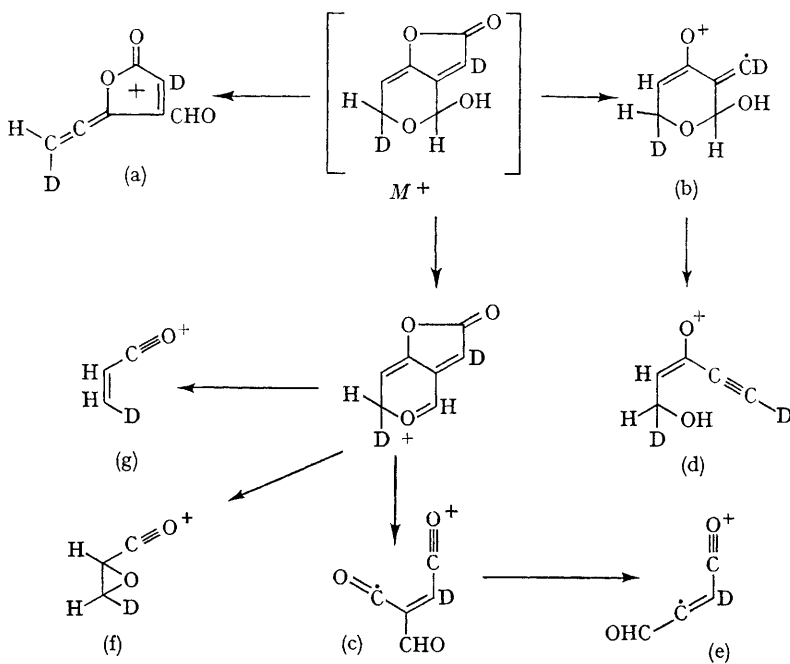
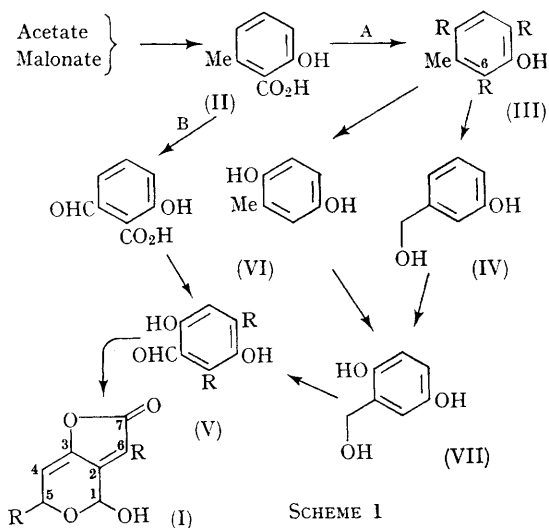
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THE convenient and rapid direct mass-spectrometric mapping of the biosynthesis of indole alkaloids has recently been described.<sup>1</sup> The same technique has now been applied to the biosynthesis of patulin (I) in *Penicillium patulum*. Although conversion of 6-methylsalicylic acid (II) into patulin (I) has been previously demonstrated,<sup>2</sup> the nature of other intermediates has remained obscure. The generally accepted pathway is as shown in Scheme 1, path B.

The discovery of several new phenolic metabolites of *P. patulum*, namely *m*-cresol,<sup>3,4</sup> (III; R = H) *m*-hydroxybenzyl alcohol<sup>5</sup> (IV) and toluquinol<sup>4</sup> (VI), suggested (III; R = H) as the most likely intermediate after 6-methylsalicylic acid (II). When [2,4,6-<sup>2</sup>H<sub>3</sub>]-3-methylphenol (III; R = D) was administered to *P. patulum* under normal culture conditions and the derived patulin examined in the mass spectrometer, the enrichment of the (*M* + 2) peak (156) corresponded to a 30% incorporation of (III; R = D) (after correction for the loss of one deuterium atom). Using a glucose-deficient medium the conversion of (III; R = D)

into (I; R = D) was 57.9% (*M* + 2/*M* = 1.4). The mass spectrum of (I; R = D) disclosed seven principal fragment ions (a)–(g) (Table; Scheme 2)



in accord with the predicted labelling pattern. These assignments are supported by high-resolution mass measurements (Table) and by n.m.r. integration measurements which, for example, show the ratio of the C-5 protons in (I; R = H) to (I; R = D) to be 1.5/1.0.

TABLE  
Relative abundance of ( $M + 2$ ) and ( $M + 1$ ) peaks in deuteropatulin

Mass	Fragment	Ratio	
		( $M + 2$ )/ $M$	( $M + 1$ )/ $M$
156	$M + 2$	1.4	—
138	(a)	1.4	—
128	(b)	1.4	—
111	(c) (111.0061)	—	1.3
99	(d) (99.0392)	1.4	—
83	(e) (83.0115)	—	1.5
72	(f) (72.0197)	—	1.3
56	(g) (56.0241)	—	1.3

Ratios were obtained in each case by comparison with the corresponding ( $M - 2$ ) or ( $M - 1$ ) peaks in unlabelled patulin.

Since deuterium from C-6 in (III; R = D) is retained at C-5 in patulin, ring fission of gentisaldehyde (V), a presumed intermediate, must be followed by a stereospecific reduction. In support of this step the deuteropatulin (I; R = D) was found to be optically active [ $M$ ]<sub>320</sub>  $-122^\circ$ ; [ $M$ ]<sub>300</sub>  $+3800^\circ$ , whereas natural patulin is optically inactive in all accessible regions of the spectrum.

In addition, the toluquinol (VI), gentisyl alcohol (VII), and gentisaldehyde (V) obtained from this source showed a deuterium enrichment of the ( $M + 2$ ) peak corresponding to respective incorporations of 71.0, 65.6, and 66.7% of (III; R = D). It is interesting to note that, in contrast to the observations of Witkop and others,<sup>6</sup> no intramolecular migration of deuterium during the course of the aromatic hydroxylation (III  $\rightarrow$  VI or IV  $\rightarrow$  VII) is evident in our experiments as this would require the presence of an ( $M + 3$ ) peak in the mass spectra of our products.

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<sup>2</sup> S. W. Tanenbaum and E. W. Bassett, *J. Biol. Chem.*, 1959, **234**, 1961; *Biochem. Biophys. Acta*, 1960, **40**, 535.

<sup>3</sup> J. D. Bu'Lock, D. Hamilton, M. A. Hulme, A. J. Powel, H. M. Smalley, D. Shepherd, and G. N. Smith, *Canad. J. Microbiol.*, 1965, **11**, 765.

<sup>4</sup> A. I. Scott and M. Yalpani, unpublished results.

<sup>5</sup> M. C. Rebstock, *Arch. Biochem. Biophys.*, 1964, **104**, 156.

<sup>6</sup> D. Jerina, J. Daly, W. Landis, B. Witkop, and S. Udenfriend, *J. Amer. Chem. Soc.*, 1967, **89**, 3347.

<sup>7</sup> Cf. S. H. Pomeranz, *J. Biol. Chem.*, 1966, **241**, 161 and references cited, wherein mammalian tyrosinase is shown to hydroxylate tyrosine to Dopamine with loss of hydrogen *ortho* to the phenolic hydroxy-group. A similar effect has recently been observed in plants (A. R. Battersby) and in mammalian systems (J. Fishman [private communications]).