

The Transfer of the Double Bond from 8(9) to 7(8) in Cholesterol Biosynthesis

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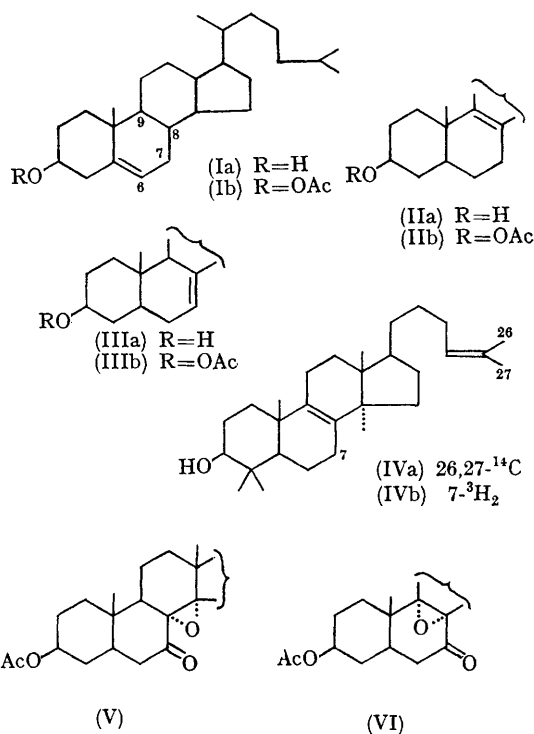
RECENT evidence from several laboratories suggests that the transfer of the double bond from 8(9) to 5(6) in cholesterol biosynthesis occurs through the combination of three reactions.¹⁻⁴ An initial migration of the double bond from 8(9) to 7(8) which is then followed by an oxidative dehydrogenation^{2,3} to give a 5,7-diene system; and finally, a NADPH-linked reduction of the 7(8) bond to furnish the 5(6) double bond of cholesterol.⁴ Here we report observations pertinent to the migration of the double bond from 8(9) (IIa) to 7(8) (IIIa). Our main objective was to differentiate between two main mechanisms:

(i) an intramolecular hydrogen transfer mechanism in which one of the C-7 hydrogens is transferred to C-9,

(ii) an "addition-elimination" mechanism in which one of the C-7 protons is lost and one from water is incorporated at C-9 during the biosynthesis. Evidence proves that the migration of the double bond from 8(9) to 7(8) occurs through an "addition-elimination" mechanism.

For our mechanistic studies we needed a ¹⁴C-labelled lanosterol and lanosterol labelled specifically with tritium at C-7. [26,27-¹⁴C]Lanosterol was synthesised by the method described previously⁵ and [7-³H₂]lanosterol was prepared as follows: dry chloroform (10 ml.) was saturated with tritiated HCl gas for 10-15 min. Lanosteryl acetate dibromide (250 mg.) was then added and allowed to equilibrate at room temperature for 4 hr. The resulting mixture of tritiated Δ⁸- and Δ⁷-dibromide acetates was debrominated (Zn/benzene-methanol) and separated on a silica gel-silver nitrate plate. The [7-³H₂]lanosteryl acetate thus prepared incorporates radioactivity at both the C-7 hydrogens.

When a mixture of [26,27-¹⁴C]- and [7-³H₂]-lanosterol (IVa and IVb) (T/C = 5.6) was incubated with a 10,000 g. supernatant of rat liver



homogenates under aerobic conditions,⁴ the biosynthetic cholesterol (Ia; T/C = 1.9) which was isolated as the dibromide had lost 66% of the tritium activity. That the remaining activity

was still located at C-7 of cholesterol was shown by the conversion of the biosynthetic cholesteryl acetate (Ib) into 3 β -acetoxycholest-5-en-7-one which resulted in the removal of 82% of the remaining activity (T/C = 0.34). This experiment therefore proves that in the migration of the double bond from 8(9) to 7(8) during biosynthesis, one of the hydrogens at C-7 is lost and the other remains undisturbed. (The loss of 66% of the activity instead of 50% shows the uneven incorporation of label at C-7 α and C-7 β during the chemical synthesis.)[†]

In order to locate the origin of the C-9 hydrogen during the biosynthesis, non-radioactive cholest-8(9)-enol (IIa) was incubated for 2 hr. with rat liver microsomes⁶ under anaerobic conditions in the presence of tritiated water (counts/min./mg. atom of hydrogen = 3.4×10^8). This resulted

in the incorporation of 10^6 counts/min. in the biosynthetic cholest-7-enol.

After the addition of carrier, biosynthetic 3 β -acetoxycholest-7-ene (IIIb; counts/min./mmole = 6×10^4) was oxidised with chromium trioxide-acetic acid⁷ to give a mixture of 3 β -acetoxy-8,9-epoxycholestan-7-one (VI; counts/min./mmole = 1150) and 3 β -acetoxy-8,14-epoxycholestan-7-one (V; counts/min./mmole = 5.8×10^4). The complete retention of activity in (V) and its complete removal in (VI) unambiguously proves that the C-9 hydrogen of the biosynthetic cholest-7-enol (IIIa) contained all the radioactivity.

We therefore conclude that the biological transfer of the 8(9)-double bond to 7(8) occurs through an "addition-elimination" mechanism.

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[†] The distribution of radioactivity between the two hydrogen atoms at C-7 of lanosterol may vary from experiment to experiment, because of the difficulty involved in maintaining a known concentration of hydrogen chloride in the reaction mixture. Also one sample of labelled lanosterol prepared by the equilibration method contained 15–20% of the activity at positions other than C-7.

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