Biosynthesis of the Cinchona Alkaloids: Late Stages of the Pathway

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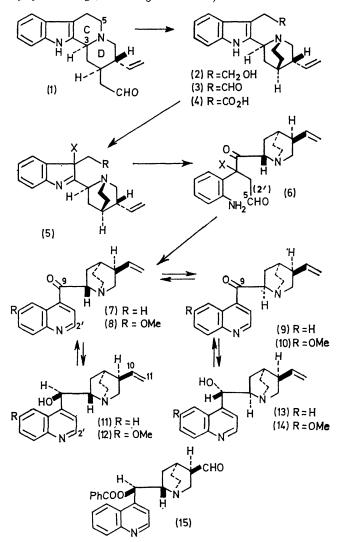
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Summary A combination of evidence from isolations and from tracer experiments supports the sequence $(1) \rightarrow (7)$ and $(9) \rightleftharpoons (11)$ and (13) (and similarly for the methoxylated series) as the pathway to the *Cinchona* alkaloids: the results also point to cinchonaminal (3) as being the key intermediate between (1) and (7).

THE preceding communication¹ traced the biosynthetic pathway to the quinoline alkaloids of *Cinchona* as far as corynantheal (1). Ideas²⁻⁴ regarding the subsequent steps have made use of cinchonamine (2) or a close relative as an intermediate and our working hypothesis was the sequence $(1) \rightarrow (3) \rightarrow (5) \rightarrow (6) \rightarrow (7)$. The ketone (7) could then lead, as illustrated, to the main quinoline bases (11-14). Experiments which support this scheme are now outlined.

An understanding of the processes whereby ring c of corynantheal (1) is cleaved requires knowledge of the oxidation level at C-5 of (1) as it passes along the biosynthetic pathway to become C-2' of the quinoline bases [see e.g. (11)]. This was examined by preparing $[1-^{3}H_{2}]$ tryptamine from indolvlacetonitrile using borotritiidecobalt chloride.⁵ When the product was administered to Cinchona ledgeriana shoots in admixture with [1-14C]tryptamine, the results under Expt. 1 (Table) were obtained. Cinchonidinone (7), prepared by oxidation⁶ of cinchonidine (11), was added to the work-up of Expt. 1 in addition to the three known bases (11-13). The isolated ketone was purified to constant activity and then was reduced with borohydride to a mixture of cinchonidine (11) and cinchonine (13); the latter was purified to constant activity. The results from Expt. 1 show (i) that cinchonidinone (7) is a natural product, (ii) within experimental error, 50% of the tritium at C-1 of tryptamine [corresponding to C-5 of corynantheal (1)] is lost during the formation of the four quinoline systems and it follows that oxidative attack at C-5 of (1) is a stereospecific process, and (iii) a carboxylic acid such as (4) cannot be a biosynthetic intermediate.

The methoxylated ketones (8 and 10) would also be expected to be present in *Cinchona* plants and this was tested by adding quinidinone (10) to the alkaloids from



Expt. 1 described in the preceding communication. Re-isolation and rigorous purification afforded active quinidinone and its radiochemical purity was checked by reduction as yield formaldehyde from C-11 which carried 105% of the original molar activity. Labelling was thus proved to be entirely at the expected site.

Tracer experiments on Cinchona ledgeriana

Expt.		Incorporations $(\%)$			
no.	Precursor	Cinchonidinone (7)	Cinchonine (13)	Cinchonidine(11)	Quinine(12)
1.	[1-³H₂,1-¹⁴C]Tryptamine Ratio ³ H : ¹⁴ C 4·70	0·47; Ratio 2·28 (48% retention ³ H)	0·20; Ratio 2·33 (50% retention ³ H)	0.12; Ratio 2.45 (52% retention ³ H)	0·33; Ratio 2·48 (53% retention ³ H)
2.	[11- ³ H ₂]Cinchonidinone(7)	, ,,	0.14	0.03	0.002
3. 4.	[11- ⁸ H ₂]Cinchonidine(11) [Ar- ³ H]Cinchonamine(2)	0.06	0.001	0.0008	0.0001

described above to give quinine (12) and quinidine (14). The latter was isolated and finally purified as its phenylcarbamate derivative; its constant activity corresponded to 0.002% incorporation which establishes the presence of quinidinone (10) in the plants.

[11-3H₂]Cinchonidinone (7) was prepared for incorporation experiments as follows. Treatment of O-benzovlcinchonidine [cf. (11)] with osmium tetroxide-periodate yielded the aldehyde (15), m.p. 189-191° which was converted by [methylene-³H₂]triphenylphosphine methide into O-benzoyl-[11-3H2]cinchonidine. Specificity of labelling was confirmed by cleaving the vinyl group as before from a diluted aliquot of this product to give formaldehyde which carried 98% of the activity of the starting material. The ester group of the high activity O-benzoyl-[11-3H2 cinchonidine was hydrolysed, and oxidation then afforded $[11-^{3}H_{2}]$ cinchonidinone (7). Expt. 2 showed that this precursor was incorporated into the main Cinchona alkaloids. The isolated cinchonine (13), as its phenylcarbamyl derivative, was degraded with osmium tetroxide-periodate to

The reversibility of the step $(7) \rightarrow (11)$ was studied by feeding [11-3H2] cinchonidine (11) to C. ledgeriana shoots and isolating cinchonidinone (7), added as carrier. The product was purified by the same reductive sequence used above and the result (Expt. 3) showed that reversal does occur [*i.e.*, $(11) \rightarrow (7)$].

Finally, cinchonamine (2) was assessed as a possible precursor of the Cinchona alkaloids by Ar-labelling (using [³H]trifluoroacetic acid) and administration of the product to the plants. The incorporations (Expt. 4), though not zero, are so low that it is unlikely that cinchonamine (2) is on the direct pathway. This result taken in conjunction with Expt. 1 makes it probable that cinchonaminal (3) is the key intermediate between corynantheal (1) and the 9-ketoquinoline bases (7-10).

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