

^{19}F Magnetic Resonance Spectroscopic Investigation of the Binding of 2-Deoxy-2-trifluoroacetamido- α -D-glucose to Lysozyme

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Summary The ^{19}F n.m.r. signal of 2-deoxy-2-trifluoroacetamido- α -D-glucose, but not the β -anomer, shows an upfield shift in the presence of lysozyme.

It has been shown that when both anomers of 2-acetamido-2-deoxy-D-glucose are bound to lysozyme there are upfield shifts in the ^1H n.m.r. signals of the *N*-acetyl groups.¹⁻³ We find that there is a similar shift of the fluorine signal of 2-deoxy-2-trifluoroacetamido- α -D-glucose† but that certain

significant differences suggest that this compound is bound differently to 2-acetamido-2-deoxy- α -D-glucose.

Mutarotated 2-deoxy-2-trifluoroacetamido-D-glucose has a similar inhibiting effect to 2-acetamido-2-deoxy-D-glucose on the rate of hydrolysis of the cell walls of *Micrococcus lysodeikticus* catalysed by lysozyme (Hens' egg white lysozyme from Boehringer). Thus under standard conditions this reaction was 24% inhibited in the presence of 2.5×10^{-2} M-2-acetamido-2-deoxy-D-glucose and 20%

† M.p. 199—202°, $[\alpha]_{\text{D}}^{20} = 38.0 \rightarrow 13.6^\circ$ (*c*, 1.0 in H_2O).

inhibited in the presence of 1.8×10^{-2} M 2-deoxy-2-trifluoroacetamido-D-glucose. An aqueous mutarotated solution of 2-deoxy-2-trifluoroacetamido-D-glucose has ^{19}F -signals at 339.6 and 312.0 Hz \ddagger downfield from external trifluoroacetic acid arising from the α - and β -anomers respectively. The signal of the α -anomer moves upfield in the presence of lysozyme (3.9×10^{-3} M) and the extent of this shift depends on its concentration. The signal of the β -anomer moves downfield *ca.* 4 Hz and this shift is concentration independent. When tetra-*N*-acetylchitotetraose is added at a concentration 1.2 times that of the lysozyme the ^{19}F signals of the α - and β -anomers appear 0.7 and 3.9 Hz downfield from their original position in the absence of enzyme. These downfield shifts presumably arise from a medium or bulk-susceptibility effect since downfield shifts were also found in the presence of chymotrypsin. The upfield shift of the α -anomer which is suppressed by tetra-*N*-acetylchitotetraose must therefore arise from binding in the active site.

The variation of the shift of 2-deoxy-2-trifluoroacetamido- α -D-glucose with its concentration was analysed by the method of Sykes³ and values of 78 Hz for the differences in chemical shift between unbound and bound 2-deoxy-2-trifluoroacetamido- α -D-glucose and 9.1×10^{-3} mol l⁻¹ for the binding constant were calculated. The analogous values for 2-acetamido-2-deoxy- α -D-glucose determined

under identical conditions were 87 Hz and 2.04×10^{-2} mol l⁻¹. The similarity of the shift differences suggest that although 2-deoxy-2-trifluoroacetamido- α -D-glucose is bound in the active site it is situated differently from 2-acetamido-2-deoxy- α -D-glucose since if they were situated identically and experience identical changes in magnetic environment a much larger chemical shift should be found with the fluoro-compound. This conclusion is supported by the observation that methyl 2-deoxy-2-trifluoroacetamido- α -D-glucoside \S shows only a concentration-independent downfield shift in the presence of lysozyme whereas methyl 2-acetamido-2-deoxy- α -D-glucoside shows an upfield shift similar to that shown by the free sugar.^{1,2} Presumably the situation in which the *N*-trifluoroacetyl compound is bound leads to an unfavourable interaction with the enzyme when there is a methoxy-group at C-1.

Kent and Dwek have recently reported an investigation of the binding of 2-deoxy-2-fluoroacetamido- α -D-glucose to lysozyme using ^{19}F n.m.r. spectroscopy. It is interesting that there is a downfield shift in the presence of the enzyme in contrast to our results with 2-deoxy-2-trifluoroacetamido- α -D-glucose.⁵

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\ddagger All measurements were carried out at 94.1 MHz. The signals were assigned on the basis of the ^1H and ^{19}F n.m.r. spectra of the mutarotated and non-mutarotated *N*-trifluoroacetyl- α -D-glucosamine.

\S M.p. 192–193°; $[\alpha]_D^{25} = +84.6^\circ$ (*c.* 0.8 in H₂O).

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