Evaluation of the Thermodynamic Parameters for the Interconversion of the Conformational Isomers of Two N-Acyl-prolines by Nuclear Magnetic Resonance Spectroscopy

By H. L. MAIA, K. G. ORRELL, and H. N. RYDON* (Department of Chemistry, The University, Exeter EX4 4QD)

Summary Study of the ¹H n.m.r. spectra of the N-acylprolines (I) and (II) in deuteriochloroform over a range of temperature has enabled the thermodynamic parameters for the interconversion of such conformers by rotation about the N-CO bond to be evaluated for the first time.

It is generally accepted, following Pauling, Corey, and Branson,¹ that peptide bonds in acyclic peptides of α -aminoacids have the planar *trans*-conformation. This is not so for imino-acids, in which the interconversion of the *cis*- and *trans*-rotamers is sufficiently slow for their independent existence to be detected by n.m.r. spectroscopy. In the past two years such rotamers have been detected in this way in peptides of proline,² N-methylalanine³ and sarcosine,⁴ while very recently the thermodynamic parameters for the interconversion of the rotamers of N-methylalanine peptides have been determined.⁵ We now report a similar determination of the thermodynamic parameters for rotamer interconversion in the biologically important proline series.

The ¹H n.m.r. spectra of N-benzyloxycarbonyl-L-proline t-butyl ester (I) and N-benzyloxycarbonyl-O-t-butyl-Lseryl-L-proline t-butyl ester (II) in deuteriochloroform show by their complexity the presence of the *cis*- and *trans*rotamers, (a) and (b), which undergo interconversion, at a rate slow on the n.m.r. time scale, by rotation about the



[I; $R = Ph \cdot CH_2O -$; II; $R = Ph \cdot CH_2 \cdot O \cdot CO \cdot NH \cdot CH(CH_2OBu^t) -$]

N-CO bond, which is of the urethane type in (I) and of the peptide type in (II). The signals given by the t-butyl ester group in these compounds (τ 8.62 for I and 8.57 for II) are intense and sharp, with negligible spin-spin splitting; the proportions and rates of interconversion of the rotamers can be accurately determined by observing the change of line shape of these signals with tempertaure. The coalescence temperatures were 45° for (I) and 67.5° for (II).

The procedure used involved computing line shapes for the t-butyl signals for a range of proton mean life-times, using a computer programme⁶ based on the equations of Gutowsky and Holm,⁷ and fitting these to the experimental curves obtained between 25 and 90°. The line shapes were described in terms of four parameters, viz. line width at half height (above the coalescence temperature), line width at half height for the stronger signal (below the coalescence temperature), peak to valley ratio, and chemical shift difference. The sensitivity of these parameters to changes in line shape varied appreciably and the curve fitting procedure made due allowance for this. This procedure is free from the assumptions made in other, more approximate, methods⁸ and we believe that the thermodynamic parameters given below are among the most accurate so far reported for the interconversion of rotamers in amides and peptides.

The fraction of the most abundant rotamer (0.56 for I; 0.63 for II) remained constant, within experimental error, over the temperature range investigated; the more abundant rotamer is assigned the *trans*-conformation (b) on the basis of general considerations and the results of Madison and Schellman²c and Liberek *et al.*⁴ The Arrhenius plots for the rates of interconversion of the rotamers were good straight lines, the slopes and intercepts of which were determined by the method of least squares.

The thermodynamic parameters so obtained for the conversion of the *cis*- into the *trans*-rotamers of (I) and (II)

TABLE

Equilibrium constants and thermodynamic parameters for conversion of cis- into trans rotamers in deuteriochloroform at 25°

	K L	$\Delta G^0 \qquad \Delta H^0$	ΔS^{0}	ΔG ‡	ΔH ‡	ΔS^{\ddagger}
$(Ia) \rightleftharpoons (Ib) \dots \dots I$ $(IIa) \rightleftharpoons (IIb) \dots I$	kcal n l 27 -0 l 70 -0	$\begin{array}{ccc} nol^{-1} & kcal mol\\ 14 & 0 00\\ 31 & 0 00 \end{array}$	$ \begin{array}{cc} -1 & \text{ cal } \mathrm{K}^{-1} \ \mathrm{mol}^{-1} \\ & +0 \ 47 \\ & +1 \ 05 \end{array} $	$^{-1}$ kcal mol ⁻¹ +17 29 +19.18	kcal mol ⁻¹ +15 87 +21 01	$\begin{array}{c} cal \ \mathrm{K^{-1} \ mol^{-1}} \\ -4 \ 77 \\ +6 \ 14 \end{array}$

are given in the Table, they are very similar to those found by Portnova et al ⁵ for N-methylalanine peptides, save that the entropy of activation for the urethane (I) is negative The energy barrier (21 kcal mol^{-1}) we have observed in the interconversion of the rotamers for the peptide (II) is very similar to the barriers $(20-23 \text{ kcal mol}^{-1})$ measured in other ways⁹ for the interconversion of individual cis- and trans-proline residues in the very much slower isomerisation of polyproline-I

It is noteworthy that the energy barrier for the interconversion of the rotamers of the urethane (I) is 5 kcal mol^{-1} less than that for the interconversion of the rotamers of the peptide (II) Since rotamer interconversion involves overcoming the delocalisation which results in the planarity of the amide bond this implies that the degree of delocalisation, and hence the contribution of the charge-separated structure $+N=CRO^{-}$, is less in the urethane than in the peptide It is a well known fact, of great importance in peptide synthesis, that urethanes form oxazolones much less readily than do peptides Determann and his colleagues¹⁰ have ascribed this to the lesser nucleophilicity of the carbon oxygen, and greater double-bond character of the carbonyl group, in urethanes than in peptides, our results give quantitative support for this view

We thank the University of Luanda for study leave granted to one of us (H L M)

(Received, July 5th, 1971, Com 1130)

¹ L Pauling, R B Corey and H R Branson Proc Nat Acad Sci USA, 1951, 37 205 ² (a) R Garner and W B Watkins, Chem Comm 1969, 386, (b) C M Deber, F A Bovey, J E Carver, and E R Blout, J Amer. Chem Soc 1970 92 6191, (c) V Madison and J Schellman Biopolymers 1970, 9, 511, (d) H Okabayashi and T Isemura, Bull Chem Soc Japan 1970, 43, 359 ⁸ M Goodman and N S Choi, Peptides Proc Ninth European Peptide Symp, 1968, 1

⁴ B Liberek K Steporowska and E Jereczek Chem and Ind, 1970 1263 ⁵ S L Portnova, V F Bystrov, T A Balashova, V T Ivanov, and Yu A Ovchinnokov, Izvest Akad Nauk S S S R., Ser Khim., 1970, 825

⁶ T Nakagawa Bull Chem Soc Japan 1966 39, 1006

¹ I Nakagawa Bau Chem Soc Japan 1900 39, 1000
⁷ H S Gutowsky and C H Holm, J Chem Phys, 1956, 25 1228
⁸ Cf A Allerhand, H S Gutowsky J Jonas and R A Menzer J Amer Chem Soc, 1966, 88, 3185
⁹ A R Downie and A A Randall, Trans Faraday Soc, 1959, 55, 2132, I Steinberg, W F Harrington, A Berger, M Sela, and E Katchalski, J Amer Chem Soc, 1960, 82 5263, D A Torchia and F A Bovey, Macromolecules, 1971, 4, 246
¹⁰ H Determann, J Heuer, P Pfaender, and M L Reinertz, Annalen, 1966, 694, 190, H Determann, Peptides Proc Eighth European

Symp , 1966, 73