

## The Biosynthesis of Gliotoxin; Possible Involvement of a Phenylalanine Epoxide

By N. JOHNS and G. W. KIRBY\*

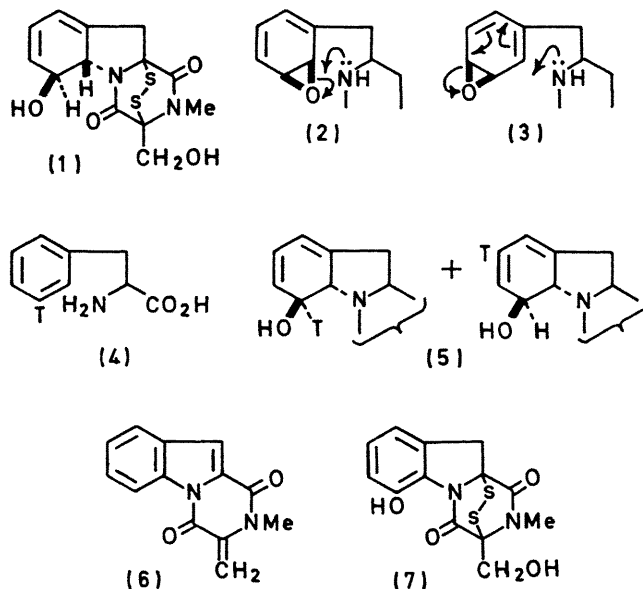
(Chemistry Department, University of Technology, Loughborough, Leicestershire LE11 3TU)

**Summary** Phenylalanine is incorporated, by *Trichoderma viride*, into gliotoxin *ca.* 100 times more efficiently than is *m*-tyrosine: incorporation of DL-[3-<sup>3</sup>H]phenylalanine occurs without loss or migration of tritium suggesting the involvement of an arene oxide intermediate.

PHENYLALANINE is well established<sup>1-3</sup> as a biosynthetic precursor for the fungal toxin, gliotoxin (1). Also, Winstead and Suhadolnik<sup>2</sup> reported that generally tritiated *m*-tyrosine was incorporated considerably more efficiently (30.8 and 44.3%) than phenylalanine (14.2 and 17.6%) into this metabolite. The biological hydroxylation of phenylalanine to give *m*-tyrosine might, in principle, involve an arene oxide<sup>4</sup> intermediate [(2) or (3)] having the appropriate functionality to cyclise directly to the unusual dihydro-benzenoid system of gliotoxin. Either intermediate could give the required *trans*-stereochemistry in the end product. We report a reinvestigation of the status of *m*-tyrosine as a gliotoxin precursor.

Radio-labelling of DL-*m*-tyrosine† was effected in tritiated 5N-HCl at 100° for 1 h. A control experiment in DCl showed (n.m.r.) that nuclear exchange had occurred, at least predominantly, *ortho* and *para* to the phenolic hydroxy-group. DL-[2,4,6-<sup>3</sup>H<sub>3</sub>]-*m*-Tyrosine was fed to *Trichoderma viride* (strain NRRL-1828) under prescribed<sup>1</sup> conditions. Only a small (0.0095%) incorporation of tritium into gliotoxin was observed (see Table) although no

tritium from the site of attack. DL-[3-<sup>3</sup>H]Phenylalanine (4), prepared from 3-bromotoluene *via* [3-<sup>3</sup>H]toluene, was



Incorporation of phenylalanine and *m*-tyrosine into gliotoxin in *Trichoderma viride*

Precursor; wt. (mg);	<sup>3</sup> H : <sup>14</sup> C	% Incorp.	Wt. of (1) (mg)	(1) <sup>3</sup> H : <sup>14</sup> C Ratios in	(6)	(7)
DL-[2,4,6- <sup>3</sup> H <sub>3</sub> ]- <i>m</i> -Tyrosine .. .. .	20	4.9	233	<i>ca.</i> 0.1		
DL-[1- <sup>14</sup> C]Phenylalanine .. .. .	0.07					
DL-[2,4,6- <sup>3</sup> H <sub>3</sub> ]- <i>m</i> -Tyrosine .. .. .	9.6					
DL-[1- <sup>14</sup> C]Phenylalanine .. .. .	9.3					
DL-[2,4,6- <sup>3</sup> H <sub>3</sub> ]- <i>m</i> -Tyrosine .. .. .	8.6	9.5 × 10 <sup>-3</sup>	215			
DL-[1- <sup>14</sup> C,3- <sup>3</sup> H]Phenylalanine .. .. .	13	3.9	189	7.51	7.51	3.91
DL-[1- <sup>14</sup> C,3- <sup>3</sup> H]Phenylalanine .. .. .	17	2.1	215	6.73	6.79	3.59

significant loss of tritium from the precursor occurred under the same conditions in the absence of the organism. In a more decisive experiment, a mixture of DL-[2,4,6-<sup>3</sup>H<sub>3</sub>]-*m*-tyrosine and DL-[1-<sup>14</sup>C]phenylalanine (<sup>3</sup>H : <sup>14</sup>C ratio, 11.5) was incubated with *T. viride*. Tritium was barely detectable in the derived gliotoxin (<sup>3</sup>H : <sup>14</sup>C ratio, *ca.* 0.1)‡ although good incorporation (4.9%) of <sup>14</sup>C was observed. A similar result was obtained when the relative weights of the two amino-acids were changed *ca.* 300 fold. It appeared therefore unlikely that, under the conditions of our experiments, *m*-tyrosine could be an obligatory intermediate in the conversion of phenylalanine into gliotoxin. However, a further test was made. Hydroxylation of phenylalanine to give *m*-tyrosine must involve loss or migration<sup>5</sup> of

mixed with DL-[1-<sup>14</sup>C]phenylalanine and the double-labelled specimen incubated with *T. viride*. No loss of tritium occurred during conversion into gliotoxin [labelling pattern (5)]. Dehydration and desulphurisation§ gave the derivative (6) again without loss of tritium. Finally, dehydrogenation<sup>6</sup> gave dehydrogliotoxin (7) with retention of 52% of the tritium. Repetition of the entire experiment gave essentially the same results which are fully consistent with a biosynthetic intermediate of the type (2) or (3). Circumstantial evidence for this possibility is provided by the structures of the aranotins which contain an oxepin ring plausibly attributed<sup>7</sup> to valence tautomerism of a phenylalanine epoxide.

Bu<sup>1</sup>Lock and Ryles<sup>8</sup> have recently published independent

† The purity and identity of commercially available (Koch-Light Ltd.) *m*-tyrosine was checked by chromatography, n.m.r. spectroscopy, and conversion, with dimethyl sulphate and alkali, into *m*-methoxycinnamic acid.

‡ Accurate measurement of small tritium activities in the presence of <sup>14</sup>C is not possible.

§ Effected with grade I, neutral alumina in dry benzene at room temperature (*cf.* ref. 3).

observations leading to the same conclusions. In particular they showed that *m*-tyrosine was poorly incorporated (highest value, 0.053%) into gliotoxin, and phenylalanine, fully deuteriated in the aromatic ring, gave [<sup>2</sup>H<sub>5</sub>]-gliotoxin as the major deuteriated metabolite.

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