

## Enzymic Formation of Siccanochromen-A, a Key Intermediate in the Biosynthesis of Siccanin

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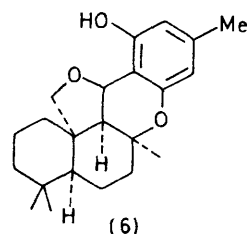
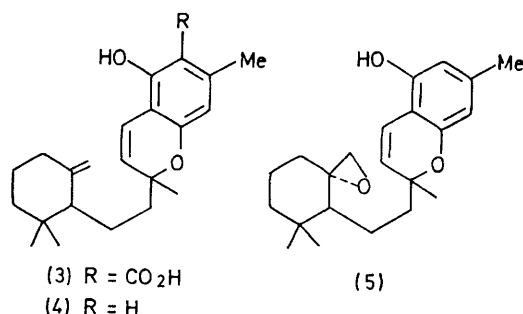
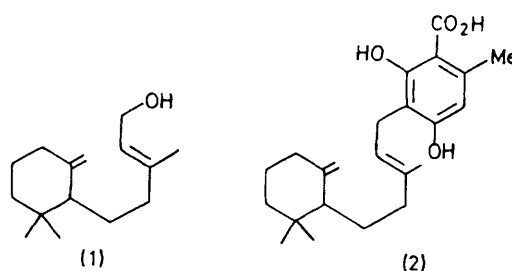
**Summary** Enzymic formations of siccanochromene-A (4) by incubation of cell-free systems of *Helminthosporium siccans* Drechsler with mevalonic acid lactone and orsellinic acid, *trans,trans*-farnesyl pyrophosphate and orsellinic acid, presiccanochromenic acid (2), and siccanochromenic acid (3) are described.

RECENTLY we demonstrated that *trans*- $\gamma$ -monocyclofarnesol (1),<sup>1</sup> a likely precursor of siccanin (6),<sup>2</sup> is derived in cell-free systems from mevalonic acid lactone. The importance of the intermediary chromen derivatives, siccanochromen-A (4)<sup>3</sup> and -B (5),<sup>3</sup> in siccanin biosynthesis has also been established by growing cell systems.<sup>4</sup> We now report the formation of siccanochromen-A (4) in cell-free systems of *H. siccans*.

[2-<sup>14</sup>C]Mevalonic acid lactone was incubated for 3 h at 36° in the presence of ATP and orsellinic acid† with the cell-free supernatant prepared by disrupting the cells from *H. siccans* with washed sea sand and centrifuging at 11,000 *g* in phosphate buffer (0.1 M) containing Mg<sup>2+</sup> (0.1 mM). The autoradiograph of the nonsaponifiable fraction of the extract contained a spot identical with (4) (conversion ratio: ~30%). The incubation of *trans,trans*-[4,8,12-<sup>14</sup>C]-farnesyl pyrophosphate‡ with the same cell-free supernatant and orsellinic acid, but without ATP, also afforded (4) in quantitative yield. The isolated siccanochromen-A (4) and its acetate were identical with authentic samples by co-chromatographs on silica gel, silica gel-silver nitrate, and reversed phase-silica gel plates, as well as by radio gas chromatography.

Other aromatic compounds such as 2-*n*-propyl-4,6-dihydroxybenzoic acid,§  $\alpha$ - and  $\beta$ -resorcylic acid, orcinol, and resorcinol were found not to be effective as co-substrates for the enzyme systems.

When unlabelled presiccanochromenic acid<sup>5</sup> (2) (1 mg/ml) was incubated aerobically with the same supernatant, (4) was obtained in 40% yield with recovered (2) (50%) and (3)



\* Hydrolysis product of gyrophoric acid from *Gyrophora esculenta*.

† Prepared by the incubation of [2-<sup>14</sup>C]mevalonic acid lactone with pig liver homogenate and purified by DEAE cellulose column. G. Popjak, J. Clifford, V. Williams, and J. Edmond, *J. Biol. Chem.*, 1969, **244**, 1897.

§ Hydrolysis product of divaricatic acid from *Ramalia subbreviscula*.

(4%). Anaerobic incubation however, afforded no transformation products, indicating that oxygen is required for the formation of the chromen ring.

Siccanochromenic acid<sup>5</sup> obtained from *H. siccans* was found to be a diastereomeric mixture of (3) and episiccanochromenic acid which differ in the configuration of the methyl group on the chromen ring.<sup>6</sup> Incubation of this mixture (1.1 mg/ml) with cell-free systems of *H. siccans* gave a mixture of siccanochromen-A (4) and episiccano-

chromen-A in quantitative yield. The ratio of products was identical to that of the precursor (3), suggesting that the decarboxylase is nonspecific for the chromen ring of the substrate. Since (2) yields isomerically pure (3) and (4), the mixture of (3) and its diastereoisomer isolated from *H. siccans* may be formed non-enzymically during the isolation process.

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<sup>1</sup> K. T. Suzuki, N. Suzuki, and S. Nozoe, *Chem. Comm.*, 1971, 527.

<sup>2</sup> K. Hirai, S. Nozoe, K. Tsuda, Y. Iitaka, K. Ishibashi, and M. Shirasaka, *Tetrahedron Letters*, 1967, 2177.

<sup>3</sup> S. Nozoe, K. T. Suzuki, and S. Okuda, *Tetrahedron Letters*, 1968, 3643.

<sup>4</sup> K. T. Suzuki and S. Nozoe, *Chem. Comm.*, 1971, 527.

<sup>5</sup> S. Nozoe and K. T. Suzuki, *Tetrahedron Letters*, 1969, 2457.

<sup>6</sup> S. Nozoe and K. T. Suzuki, *Tetrahedron*, 1971, 27, 6063.