

Mechanism of Alkylation during Sitosterol Biosynthesis in *Larix decidua*

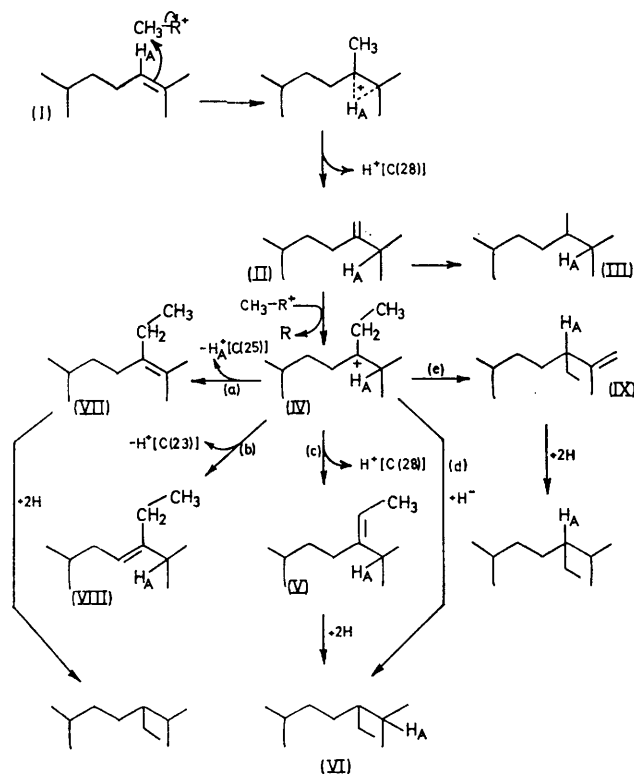
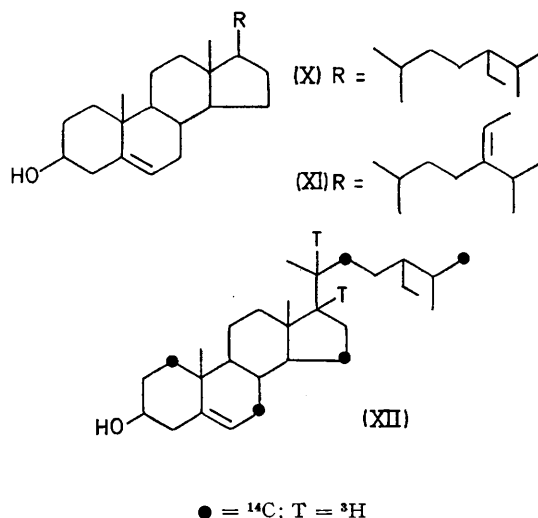
By PATRICIA J. RANDALL, H. H. REES, and T. W. GOODWIN*

(Department of Biochemistry, The University, P.O. Box 147, Liverpool L69 3BX)

Summary Incorporation of $[2-^{14}\text{C}(4R)-4-^3\text{H}_1]$ mevalonic acid into sitosterol in *Larix decidua* demonstrates that during alkylation, the C-24 hydrogen of the Δ^{24} precursor is eliminated.

THE alkyl side chain at C-24 in phytosterols arises by transmethylation from *S*-adenosylmethionine.¹ A suggested² mechanism for this process involves hydrogen migration from C-24 to C-25 and formation of 24-methylene (II) and 24-ethylidene (V) compounds as precursors of C-24 methyl (III) and C-24 ethyl (VI) sterols, respectively (Scheme). In support of this Scheme, certain C-24 methyl and ethyl sterols biosynthesised by lower organisms in the presence of $(\text{CD}_3)_3$ -methionine contain only two and four deuterium atoms, respectively.³ In contrast to this, C_{28} and C_{29} alkyl-saturated sterols biosynthesised in other lower organisms retained three and five deuterium atoms, respectively^{4,5} thus excluding the involvement of 24-methylene (II) and 24-ethylidene (V) intermediates in their biosynthesis. This has been explained⁵ by invoking a $\Delta^{24(25)}$ intermediate, which is in agreement with the observation that the hydrogen at C-24 in the Δ^{24} sterol precursor is eliminated during stigmasterol biosynthesis in *Nicotina tabacum* and *Dioscorea tokoro*.⁶

in biosynthesis of any C-24 methyl- and ethyl- sterols in higher plants. Similarly, the hydrogen at C-24 in the precursor Δ^{24} sterol has only been conclusively shown to be retained at C-25 during ergosterol biosynthesis in yeast,⁷ but never during biosynthesis of any C-24 saturated alkylsterol in higher plant tissue. In order to establish the fate of the C-24 hydrogen of the Δ^{24} -sterol precursor in phytosterol biosynthesis in higher plants we report the incorporation of $[2-^{14}\text{C}(4R)-4-^3\text{H}_1]$ mevalonic acid into sitosterol (X) and 28-isofucoesterol (XI) in *Larix decidua* leaves.



SCHEME. H_A is derived from the 4-pro-R hydrogen of mevalonic acid.

However, 24-methylene (II) and 24-ethylidene (V) sterols have not been conclusively shown to be intermediates

Young leaves of *Larix decidua* were chosen since sitosterol is the principal sterol⁸ and 28-isofucoesterol has also been radiochemically detected⁹ in this tissue. We have now further characterized 28-isofucoesterol as a minor component of *Larix decidua*, from a partly purified sterol fraction by g.l.c. and g.l.c.-m.s. (3% OV-17 column).

Chopped young leaves (2 g) of *Larix decidua* were incubated with $[2-^{14}\text{C}(4R)-4-^3\text{H}_1]$ mevalonate ($10 \mu\text{Ci}^{14}\text{C}$) overnight at 25° and the non-saponifiable lipids isolated. The sterols and squalene were obtained by t.l.c., and the squalene further purified after addition of carrier material, *via* the thiourea adduct (twice) and hexahydrochloride. Compounds (X) and (XI) were separated from the sterol fraction, after addition of carrier (XI) by t.l.c. on AgNO_3 impregnated silica gel. G.l.c. (3% OV 17 column) of the respective zones with trapping of samples at 1 min intervals for radioassay, indicated that all the radioactivity within each zone was associated with sitosterol and 28-isofucoesterol, respectively. After dilution with carrier material, sitosterol and 28-isofucoesterol were recrystallised to constant specific radioactivity. Since the $^3\text{H}:^{14}\text{C}$ atomic ratio (Table) of the recrystallised sitosterol (X) was between 2:5 and 3:5, portions of (X) were further processed by (i) formation of the dibromide derivative and regeneration of the free sterol in ether with zinc-acetic acid and recrystallization¹⁰ and (ii) treatment with Jones reagent yielding 24*R*-ethylcholest-4-en-3,6-

dione.¹¹ The latter was further transformed after dilution with carrier material by treatment with zinc and acetic acid into 24*R*-ethylcholestane-3,6-dione,¹² m.p. 186—188°; *m*/*e* 428.

The ³H:¹⁴C ratios for the sitosterol purified *via* dibromide, the 24*R*-ethyl-cholest-4-ene-3,6-dione and the 24*R*-ethyl-cholestane-3,6-dione are in good agreement (2:5), and indicate the presence of only two tritium atoms in the biosynthesised sitosterol. By analogy with cholesterol biosynthesis in animals,¹³ the compound (X) should be labelled

formation in *Nicotiana* and *Dioscorea* tissue cultures, and support the operation of route (a) (Scheme) involving a Δ²⁴⁽²⁵⁾ intermediate (VII) but not routes (b), (c), (d), or (e), during sitosterol biosynthesis. A ³H:¹⁴C ratio of 3:5 for 28-isofucosterol (XI)≡(V) is in agreement with the established¹⁴ operation of route (c) in its biosynthesis. The present results indicate that 28-isofucosterol is not on the major pathway to (X) in *Larix decidua*, at least not without isomerization to a Δ²⁴ structure. In view of this, other earlier reports¹⁵ of the incorporation of [2-¹⁴C-(4*R*)-

TABLE. Incorporation of [2-¹⁴C-(4*R*)-4-³H₁]mevalonate into sitosterol and 28-isofucosterol in *Larix decidua*

Compound	Specific radioactivity (in d.p.m. ¹⁴ C mg ⁻¹)	³ H: ¹⁴ C radioactivity ratio ^a	³ H: ¹⁴ C atomic ratio (based on squalene)
Squalene	7.54	
Sitosterol (X) (recrystallized)	228	2.43:5
Sitosterol (X) (purified <i>via</i> dibromide)	350	2.23:5
24 <i>R</i> -Ethylcholest-4-ene-3,6-dione	340	2.21:5
24 <i>R</i> -Ethylcholestane-3,6-dione ^b	174	2.08:5
28-Isifucosterol (XI) (recrystallized)	253	3.00:5

^a The specific radioactivities and radioactivity ratios quoted are the mean of three sequential crystallisations in each case. ^b Diluted with carrier material before recrystallization.

as shown (XII). The higher ³H:¹⁴C ratio observed for the recrystallized sitosterol which had not been purified further indicates that a radioactive impurity, with a higher ³H:¹⁴C ratio (possibly the corresponding Δ⁷ sterol), had been carried through the recrystallization.

The present results for sitosterol (X) are in agreement with those of Tomita and his co-workers⁶ for stigmaterol

4-³H₁]mevalonic acid into higher plant sterols, which gave inconclusive results will require further careful reinvestigation.

We thank the S.R.C. for a Studentship to P.J.R. and Dr. J. G. Lloyd-Jones for helpful discussions.

(Received, 5th October 1972; Com. 1700.)

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