# Biomimetic Inversion of C-3 in Monoterpenoid Indole Alkaloids 

By Richard T. Brown,* C. Lyn Chapple, and Ray Platt<br>(Department of Chemistry, The University, Manchester M13 9PL)<br>and Harry Spencer

(I.C.I. Pharmaceuticals Division, Mereside, Alderley Park, Macolesfield, Cheshire SK10 4TG)

Sommary An inversion of $3-\mathrm{H}$ analogous to that required in vivo has been found to occur spontaneously under mild conditions during a synthesis of Corynanthé-type alkaloids from a vincoside derivative.

Two intriguing points to emerge from biosynthetic studies on monoterpenoid indole alkaloids were (i) that even for the Corynanthé type with $3 \alpha(S)$ stereochemistry, the exclusive precursor was a $3 \beta(R)$ epimer, vincoside (1a), and (ii) that the consequent inversion of $\mathrm{C}-3$ occurred with retention of hydrogen. ${ }^{1}$ The former has been rationalised by invoking a hypothetical intermediate, mancunine (2), which was substantiated by synthesising a closely related structure readily converted into known alkaloids. ${ }^{2,3}$ We now report the synthesis from a vincoside derivative of two $3 \alpha$ Corynanthé alkaloids- $N^{1}$-methyltetrahydroalstonine (8a) and $N^{1}$-methyl-20 $\beta$-dihydrogeissoschizine (10a) in a sequence which provides a satisfactory analogy and mechanism for the latter process.

A recent synthesis ${ }^{4}$ of heteroyohimbine alkaloids from (1b) was forced to deviate from the presumed biogenetic route because (4b) could not be isolated owing to cyclisation of the indolic NH on to the $\mathrm{C}-21$ aldehyde. Hence it was repeated with a methyl blocking group on N-1. Removal of the sugar from $N^{4}$-benzyl- $N^{1}$-methylvincoside (1c), m.p. $219-221^{\circ},[\alpha]_{\mathrm{D}}^{25}-87^{\circ}(\mathrm{MeOH})$ gave two major products, isomeric with a simple aglycone but lacking the characteristic rapid shift in the u.v. spectrum on addition of alkali. However, both eventually gave a base shift when left and were assigned the gross structure of a rearranged aglycone ( $\mathbf{4 a}$ ), supported inter alia by n.m.r. signals for aldehydic protons at $\tau 0.50$ and 0.62 and $C$-methyl doublets at $\tau 8.44$ and 8.56 . Hydrogenation of the mixture with $\mathrm{Pd}-\mathrm{C}$ in $\mathrm{MeOH}-\mathrm{AcOH}$ afforded mainly two compounds together with a small amount of a third.
One major product, $\mathrm{C}_{22} \mathrm{H}_{26} \mathrm{~N}_{2} \mathrm{O}_{3},[\alpha]_{\mathrm{D}}^{25}-163^{\circ}(\mathrm{MeOH})$, [picrolonate, m.p. $176^{\circ}$ (decomp.)] did not exhibit a u.v. base shift, and its mass spectrum suggested a heteroyohimbine structure which was substantiated by appropriate n.m.r. signals. Furthermore, since $3-\mathrm{H}$ was above $\tau 6 \cdot 2$, and $19-\mathrm{H}$ appeared at $\tau 5.5$ with a trans-diaxial coupling ( 8 Hz ) to $20-\mathrm{H},{ }^{5}$ the stereochemistry was $3 \alpha, 15 \alpha, 19 \beta, 20 \alpha$, corresponding to that of tetrahydroalstonine ( $\mathbf{8 b}$ ). Methylation of the latter with $\mathrm{MeI}-\mathrm{NaOMe}-\mathrm{Me}_{2} \mathrm{SO}$ in a modification of Heaney's method ${ }^{6}$ afforded a good yield of $N^{1}$ methyltetrahydroalstonine (8a) which was identical with the previous compound.
On the other hand, the second major product, $\mathrm{C}_{22} \mathrm{H}_{28} \mathrm{~N}_{2} \mathrm{O}_{3}$, $[\alpha]_{\mathrm{D}}^{25}+92^{\circ}(\mathrm{MeOH})$ gave an immediate u.v. base shift typical of a $\beta$-hydroxyacrylate chromophore, and the mass and n.m.r. spectra were consistent only with a dihydrogeissoschizine structure. A negative Cotton effect in the c.d. spectrum between 260 and 300 nm , and an n.m.r. absorption at $\tau 5 \cdot 46$ showed that $3-\mathrm{H}$ was still $\beta$. After methylation of the enol with diazomethane, an oxidation-
reduction sequence with $\mathrm{Pb}(\mathrm{OAc})_{4}$ and $\mathrm{NaBH}_{4}$ afforded the $3 \alpha$ isomer. This proved identical with $N^{1}$-methyldihydrocorynantheine (10d), prepared from dihydrocorynantheine (10c) as above, and also with the methyl ether obtained by treatment of the minor product $\left(\mathrm{C}_{22} \mathrm{H}_{28} \mathrm{~N}_{2} \mathrm{O}_{3}\right)$ with diazomethane. Hence the second major compound must be $3 \beta, 20 \beta-N^{1}$-methyldihydrogeissoschizine ( $\mathbf{1 0 b}$ ) and the minor its $3 \alpha$ epimer (10a).

Inversion of $3-\mathrm{H}$ has thus occurred readily in two cases, and since nothing similar had been observed in previous

reactions, ${ }^{2-4}$ it could neither be caused by the $\mathrm{Pd}-\mathrm{C}$ catalyst nor take place after reduction of C-21. Deuteriation studies confirmed that $3-\mathrm{H}$ was retained throughout. We therefore postulate the following mechanism: cleavage of the C-3-$\mathrm{N}-4$ bond in the obligatory immonium intermediate (5a) is promoted by electron release from $\mathrm{N}-\mathrm{l}$ and gives the imine (6a); re-closure can then occur by attack of N-4 on C-3 from the opposite side to afford the more stable structure $(\mathbf{7 a})$, which is subsequently reduced to (8a). Formation of the dihydrogeissoschizine derivatives is presumably via retro-Michael cleavage of (5a) to (9b), and a similar conversion of (7a) into (9a) or epimerisation of (9b) prior to reduction.

Since C-H is not lost at any stage this sequence furnishes an apposite model for the in vivo process: opening of the
ether bridge in mancunine (2) generates the immonium ion (3b), convertible by inversion of $3-\mathrm{H}$ as above into demethyldehydrocorynantheine (3a). From this all the $3 \alpha$ Corynanthé alkaloids can be derived by various combinations of reduction, methylation, tautomerism, and conjugate addition. Obviously the epimerisation could occur at other stages, e.g. (9b) or (5b), but the essential mechanism would remain the same. The Ipecac alkaloids could presumably be epimerised in a similar manner but with electron release from oxygen rather than nitrogen.

We thank the S.R.C. and I.C.I. Pharmaceuticals Division for a C.A.S.E. award (R.P.).
(Received, 16th September 1974; Com. 1165.)
${ }^{1}$ J. Staunton, in 'The Alkaloids.' ed. J. E. Saxton, Specialist Periodical Reports, The Chemical Society, London, 1972, vol. 2, p. 1, and refs. therein.
${ }_{2}^{2}$ R. T. Brown and C. L. Chapple, J.C.S. Chem. Comm., 1973, 886.
${ }^{3}$ R. T. Brown, C. L. Chapple, and A. A. Charalambides, J.C.S. Chem. Comm., 1974, 756.
${ }^{4}$ R. T. Brown and C. L. Chapple, J.C.S. Chem. Comm., 1974, 740.
${ }^{5}$ R. T. Brown and A. A. Charalambides, Tetrahedron Letters, 1974, 1649.
${ }^{6}$ H. Heaney and S. V. Ley, J.C.S. Perkin I, 1973, 499.

