

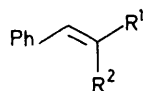
Synthesis of Homoserine Samples Stereospecifically Labelled with Isotopic Hydrogen in the β - and γ -Positions

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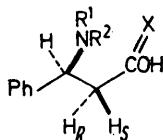
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Summary (βR)[β - 2H]-L-Homoserine (**17**) and the (βS)-isomer (**8**) are synthesised through a sequence involving as key step the formal *cis* addition of hydroxylamine onto cinnamic acid to form 3-amino-3-phenylpropionic acid; (γR)[γ - 2H]-DL-homoserine (**20**) and the (γS)-isomer (**19**) are obtained from the enantiomeric forms of stereospecifically labelled 3-phenyl[1- 2H]propanol.

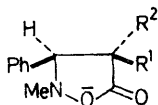
THE amino-acid L-homoserine is a key intermediate in the conversion of L-aspartic acid into L-threonine, L-homocysteine, and α -ketobutyrate in microbia and fungi.¹ This set of transformations catalysed by different pyridoxal phosphate-dependent enzymes is thought² to proceed through the intermediacy of an enzyme-bonded vinylglycine derivative arising by elimination of one of the stereo-heterotopic³ protons from the prochiral centre in the β -position together with the γ -substituent from the Schiff's base formed between a suitable *O*-derivative of homoserine



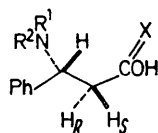
- (1) $R^1 = CO_2H$; $R^2 = D$
 (2) $R^1 = CO_2H$; $R^2 = H$
 (3) $R^1 = H$; $R^2 = CO_2Et$
 (4) $R^1 = CO_2Et$; $R^2 = H$
 (5) $R^1 = CO_2NHMe$; $R^2 = H$
 (6) $R^1 = CO_2Et$; $R^2 = D$



- (9) $R^1 = R^2 = H_5 = H$; $H_R = D$; $X = O$
 (10) $R^1 = R^2 = H_R = H$; $H_5 = D$; $X = O$
 (11) $R^1 = R^2 = Me$; $H_R = H$; $H_5 = D$; $X = O$
 (12) $R^1 = R^2 = H_5 = H$; $H_R = D$; $X = H_2$
 (13) $R^1 = R^2 = H_R = H$; $H_5 = D$; $X = H_2$



- (7) $R^1 = R^2 = H$
 (8) $R^1 = H$; $R^2 = D$

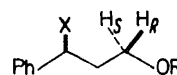
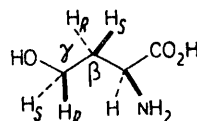


- (14) $R^1 = R^2 = H_R = H$; $H_5 = D$; $X = O$
 (15) $R^1 = R^2 = H_5 = H$; $H_R = D$; $X = O$
 (16) $R^1 = R^2 = Me$; $H_5 = H$; $H_R = D$; $X = O$

and pyridoxal phosphate. Addition of the sulphur nucleophile onto the γ methylene group gives rise, by formal reversal of the reaction pathway, to homocysteine, whereas proton addition gives rise to enzyme-bonded α -aminocrotonate, the precursor of L-threonine and α -ketobutyrate. In view of the present interest⁴ in the stereospecificity of enzyme reactions we undertook a stereochemical analysis of the above mentioned set of enzymic transformations.

We report now on the synthesis of homoserine samples asymmetrically labelled with isotopic hydrogen in the β - and γ -positions.

Addition⁵ of hydroxylamine in ethanol to (*E*)-[α - 2H]-cinnamic acid (**1**) (*ca.* 95% 2H_1) affords 3-amino-3-phenyl-[2- 2H]propionic acid [(**9**) and (**14**)], showing 1H n.m.r. signals (CF_3CO_2H ; 100 MHz) due to the side chain protons at δ 4.98 and 3.5 in an AX pattern, J_{AX} 9.5 Hz, whereas under conditions in which all the exchangeable hydrogen in the reactants and in the solvents had been substituted for deuterium the diastereoisomers [(**10**) and (**15**)] are obtained



- (17) β - $H_R = D$; β - $H_5 = \gamma$ - $H_R = \gamma$ - $H_5 = H$
 (18) β - $H_5 = D$; β - $H_R = \gamma$ - $H_R = \gamma$ - $H_5 = H$
 (19) γ - $H_5 = D$; β - $H_R = \beta$ - $H_5 = \gamma$ - $H_R = H$
 (20) γ - $H_R = D$; β - $H_R = \beta$ - $H_5 = \gamma$ - $H_5 = H$
 (21) $H_R = R = X = H$; $H_5 = D$
 (22) $H_5 = R = X = H$; $H_R = D$
 (23) $H_R = H$; $H_5 = D$; $R = COMe$; $X = Br$
 (24) $H_R = H$; $H_5 = D$; $R = COMe$; $X = N_3$
 (25) $H_R = H$; $H_5 = D$; $R = COMe$; $X = NH_2$

from (*E*)-cinnamic acid (**2**), showing a BX pattern at δ 4.98 and 3.25 with J_{BX} 4.0 Hz. The absolute steric course of the addition of the nitrogen nucleophile across the double bond of cinnamic acid was established to be *cis* since ozonolysis of (**9**) and (**14**) in formic acid, gave DL-[2- 2H]-aspartic acid, shown to be the *erythro*-isomer by 1H n.m.r. spectroscopy and comparison with an authentic sample.⁶

The above-mentioned steric course was also observed using ethyl cinnamate (**4**) as substrate. However, from (*Z*)-ethyl cinnamate (**3**) in deuteriated solvents a mixture of [(**9**) and (**14**)] and [(**10**) and (**15**)] in a ratio of *ca.* 8:2 was obtained as shown by the relative intensities of the 2-H signals, thus indicating that with the *cis*-isomer as substrate the reaction is only partially stereospecific.

Recent studies⁷ on the mechanism of addition of α -nucleophiles onto $\alpha\beta$ -unsaturated substrates have shown that the isoxazolidone (**7**) is obtained from ethyl cinnamate (**4**) and *N*-methylhydroxylamine through the possible intermediacy of the *O*-acyl derivative (**5**). In our experiments, *N*-methylhydroxylamine and both (*E*)-[α - 2H]cinnamic acid (**1**) and ethyl (*E*)-[α - 2H]cinnamate (**6**) gave the deuteriated isoxazolidone (**8**), showing n.m.r. signals due to the ring protons at δ 3.45 and 2.53 (d, J_{BX} 11.8 Hz). The latter compound was converted (H_2 , Raney-nickel, followed by methylation) into 3-*NN*-dimethyl-3-phenyl[2- 2H]propionic acid [(**11**) and (**16**)], whose 1H n.m.r. spectrum was identical to that of the compound obtained from [(**9**) and (**14**)] upon methylation. Compound (**8**) loses deuterium upon mild alkaline treatment.

Resolution⁸ of [(**9**) and (**14**)] and [(**10**) and (**15**)] gave (2*R*, 3*S*)-(**9**) and (2*S*, 3*S*)-(**10**), reduced in boiling dioxan with $LiAlH_4$ to the alcohols (**12**) and (**13**). Compounds (**12**)

and (13), after acetylation, upon ozonolysis, oxidative work up, and acid hydrolysis, gave L-(βR) [β-²H]homoserine (17) and the (βS)-isomer (18), respectively.

Homoserine stereospecifically labelled in the terminal methylene group was prepared from (1S)-3-phenyl[1-²H]-propanol⁹ (21), which, after acetylation was brominated to (23), leading, in turn, to the azide (24). Hydrogenation of (24) gave the amine (25) which, upon ozonolysis in formic

acid, afforded DL-(γS)[γ-²H]homoserine (19). DL-(γR)-[γ-²H]Homoserine (20) was similarly prepared from (1R)-3-phenyl[1-²H]propanol (22) prepared from (21) by known procedures.¹⁰

We thank Mrs. Rosanna Bernardi for the g.l.c. analyses.

(Received, 21st November 1975; Com. 1305.)

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