Biosynthesis of Phytosterols in Calendula officinalis Flowers from (2R)- and (2RS)-[2-¹⁴C, 2-³H]-Mevalonic Acid. The Incorporation of a 15α-Tritium Atom into Sitosterol

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Summary It is proven that only the 2-pro-R hydrogen of MVA is retained at the 15α -position of sitosterol biosynthesized by excised petals of Calendula officinalis flowers.

RECENTLY results indicating the possible retention of both the 2-pro-R and 2-pro-S hydrogen atoms of mevalonic acid at C-15 of sterols biosynthesized in marigold flowers (*Calendula officinalis*) were reported.¹ This could imply that the biosynthesis of phytosterols from cycloartenol, which is thought to be an intermediate in higher plants,^{2,3} proceeds by a different route from the biosynthesis of sterols from lanosterol in rat livers⁴ and yeast preparations.⁵

We have established that cycloartenol (or an intermediate with an anionic terminus at C-19) is a key precursor of phytosterols in the pea.³ In addition we showed that the end result of events around C-14 and C-15 in the biosynthesis of sitosterol in germinating peas⁶ was the same as in cholesterol⁴ and in sterols⁵ biosynthesized in rat livers and yeast homogenates, respectively. In view of the obvious inconsistency of the results for *Calendula* flowers we reinvestigated the biosynthesis of phytosterols in this plant.

Calendula officinalis plants (cv. Radio) of the same variety as previously used¹ were grown from seeds in an environmental chamber (8 weeks). Petals (120, ca. 1·18 g fresh wt.) isolated from several flowers, were placed vertically in an open weighing glass, with the ends previously attached to the flowers immersed in an aqueous solution of (3RS,2R)- $[2^{-14}C,2^{-3}H]MVA$ (11 μ Ci of ¹⁴C; ³H; ¹⁴C ratio 11·3). The assembly was then placed in the environmental chamber and when the MVA solution was absorbed (ca. 6 h) water was supplied as needed. After 6 days the petals were collected and saponified to yield a non-saponifiable residue (5·57 × 10⁶ d.p.m. of ¹⁴C). TABLE

Specific activities of ¹⁴C and ³H: ¹⁴C ratios of metabolites and their transformation products (see text)^a

					Expt. A: ¹⁴ C Specific	(3RS,2R)-[2-14C,2-3H]MVA 3H : 14C Ratio		Expt. B: 14C Specific	: (3RS,2RS)-[2-14C,2-3H ₂] ³ H : ¹⁴ C Ratio	
No.	Compound				activity	Isotopic	Atomic	activity	Isotopic	Atomic
1	MVA-amide				18·1Ő	11.09		12.4	$20 \cdot \hat{7}6$	
2	Squalene 6HCl .				8.50	9.14		30.1	13.33	
3	(1)				$22 \cdot 80$	8.90	5.00:5	7.26	14.30	8.00:5
3a	Ba (1) after bromination and									
	debromination				22.88	8.87		7.23	14.36	
4	(2)	• •			7.58	8.90	5.00:5	7.20	14.44	8.07:5
5	(3)				7.54	7.57	4.25:5	7.17	12.94	7.27:5
6	(4)				7.71	7.53	4.23:5	7.20	13.01	7.30:5
7	(5)				7.61	7.37	$4 \cdot 14 : 5$	3.36	12.75	7.16:5
8	(6)				7.54	8.92	5.01:5	7.25	14.48	8.10:5
9	Stigmasterol acetate							6.66	12.97	7-25:5

^a The results are the average of at least three crystallizations in which the ¹⁴C specific activity and ³H: ¹⁴C remained constant (±3%). Specific activity $\times 10^4$ in d.p.m. per mmol. The MVA and squalene were counted as benzhydrylamide and hexahydrochoride, respectively. The results are significant to ± 3 %. The atomic ratios are calculated on the basis of atomic ratio of sitosteryl acetate (assumed to be 5-³H:5-¹⁴C in expt. A and 8-³H:5-¹⁴C in expt. B).

iii Sitosterol acetate $\xrightarrow{i} 5\alpha$ -stigmastanol acetate $\xrightarrow{ii} 5\alpha$ -stigmast-14-en-3 β -ol acetate $\xrightarrow{iii} 5\alpha$ -stigmastane-3 β ,14 α ,15 β -triol 3-acetate (2) (1) $(\mathbf{3})$ (4)

 \rightarrow 15-oxo-5 α -stigmastane-3 β ,14 α -diol 3-acetate.

(5) SCHEME. i: EtOAc; HClO₄; PtO₂; H₂. ii: C₆H₆; C₆H₅ICl₂; $h\nu$. iii: a, CHCl₃; m-ClC₆H₄COOOH; b, Me₂CO; H₂O; HIO₄. ii: CrO₈-pyridine.

An analogous experiment (102 petals; 1.19 g fresh wt.) with $(3RS, 2RS) - [2^{-14}C, 2^{-3}H_2]MVA$ (19 µCi of ¹⁴C; ³H: ¹⁴C ratio 20.7) was carried out and a non-saponifiable residue $(9.87 \times 10^6 \text{ d.p.m. of } {}^{14}\text{C})$ was obtained. The 'R' and 'RS' squalenes and sitosteryl acetates were extensively purified by chromatography, derivatization, and crystallization to yield homogenous metabolites.1,5

The calculated and observed ³H:¹⁴C atomic ratios of MVA benzhydrylamide, squalene hexahydrochloride, and sitosteryl acetate (Table; expts. A and B, entries 1-3) show significant discrepancies. Such discrepancies were previously noted and are considered to be related to the metabolism of polyprenoids in plants.1,6

The obtained 'R' and 'RS' situates were then submitted to the sequence of transformations outlined in the Scheme. The transformation of the derived 'R' and 'RS' 5α -stigmastanyl acetate to 5α -stigmast-14-en-3 β -ol acetate and 5α -stigmast-9(11)-en-3 β -ol acetate was carried photochemically in the presence of $C_{6}H_{5}ICl_{2}$.⁷ We have proven that the C-14 dehydrogenation of 5α -cholestan-3 β -ol acetate involved the overall abstraction of the cis 14α and 15 α hydrogen atoms.⁵ It may be assumed with certainty that the C-14(15) dehydrogenation of 5α -stigmastan-3 β -ol acetate will also proceed via the removal of the 14α and 15α hydrogen atoms.

For the discussion of the results we will assume that the

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specimens of sitosteryl acetates have the indicated ³H:¹⁴C atomic ratios (Table; expts. A and B, entries 3). These ratios are based on results on the biosynthesis of sterols in several species (rat, pea, yeast, etc.).

The results show that the 'R' sitosteryl acetate (Table; expt. A) and 'RS' sitosteryl acetate (Table; expt B) contain tritium only in the 15α position (ca. 0.75 atom) and are devoid of tritium at the 15β position.

In view of these results it must be concluded that the previous observations were in error.¹ It is also apparent that the end result of the biosynthetic events at C-14 and C-15 in Calendula flowers, rat liver homogenates,4 yeast homogenates,⁵ and in germinating peas⁶ are analogous. Very likely the loss of a hydrogen derived from 2-pro-S of MVA from C-15 occurs in the course of the formation of the C-14(15) double bond. However, since 5α -stigmast-8(14),-15,24(28)-trien-3 β -ol was isolated from Vernonia anthelminitica plant,⁸ this hypothesis requires experimental verification.

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