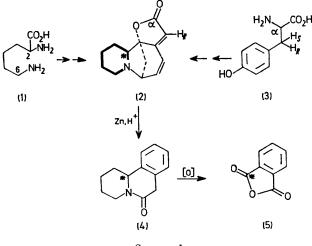
## Biosynthesis of Securinine: the Mode of Incorporation of Lysine

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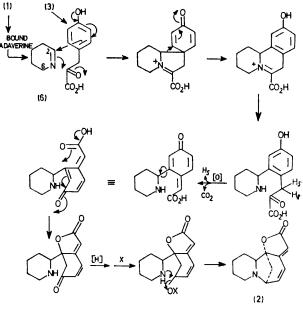
Summary The piperidine ring of securinine is derived from a  $C_{5}N$  unit,  $\Delta^{1}$ -piperideine, which originates from lysine in non-symmetrical fashion by loss of the carboxy carbon and the  $\alpha$ -amino group.

SECURININE (2), the major alkaloid of Securinega suffruticosa Rehd., originates from two amino acid fragments.<sup>1</sup> One of these, derived from tyrosine (3), yields the  $C_6-C_2$ unit of the alkaloid (light lines).<sup>1-3</sup> All the carbon atoms of tyrosine other than the carboxy group are maintained,<sup>2</sup> the  $\alpha$ -carbon supplies the lactone carbon atom of the alkaloid,<sup>1,2</sup> and the *pro-R* proton from the  $\beta$ -carbon of tyrosine is retained in the alkaloid whereas the *pro-S* proton is lost.<sup>3</sup>



SCHEME 1

piperidine alkaloids whose  $C_5N$  units are derived from lysine in non-symmetrical fashion<sup>4</sup> (*cf.*, anabasine, sedamine, *N*-methylpelletierine), or whether it belongs to the group of alkaloids whose  $C_5N$  units originate from lysine in a symmetrical manner (*e.g.*, the lupine, <sup>5</sup> decodon, <sup>6</sup> and lycopodium<sup>7</sup> alkaloids).



## SCHEME 2

The second fragment, a  $C_5N$  unit (heavy lines), is presumably derived from lysine,<sup>1,2</sup> via cadaverine. Specific incorporation of radioactivity from [1-<sup>14</sup>C]cadaverine into securinine (2), in the predicted symmetrical manner (50% of label at \*) has been demonstrated,<sup>2</sup> but the mode of incorporation of lysine has not yet been determined. In particular, it is not known whether securinine is one of the

In separate experiments  $[2^{-14}C]$ -DL-lysine  $(100 \,\mu\text{Ci}, 3 \,\text{mCi} \,\text{mmol}^{-1})$ , New England Nuclear) and  $[2^{-14}C]$ - $\Delta^{1-}$ piperideine  $(30 \,\mu\text{Ci}, 4.7 \,\text{mCi} \,\text{mmol}^{-1})$ , prepared<sup>8</sup> from  $[2^{-14}C]$ -DL-lysine) were administered to *Securinega* plants by the wick method. After 3 days in contact with tracer, the plants were harvested and securinine was extracted. The active sample was diluted with inactive carrier, crystallized to constant specific activity  $(1.5 \times 10^5 \,\text{dpm} \,\text{mmol}^{-1})$  and

 $3 \cdot 1 \times 10^4$  dpm mmol<sup>-1</sup>, respectively) and degraded<sup>2</sup> to the benzquinolizidine (4) and thence to phthalic anhydride (5) (Scheme 1). The molar specific activity of the samples of phthalic anhydride obtained  $(1.4 \times 10^5 \text{ and } 3.0 \times 10^4)$ dpm mmol<sup>-1</sup>, respectively) indicated that in each case more than 90% of the activity of securinine had been localized at the asterisked carbon atom (*i.e.*, at the only carbon atom of the non-tyrosine derived portion of securinine retained in phthalic anhydride), which is thus derived predominantly if not exclusively from C-2 of lysine and from C-2 of  $\Delta^{1}$ - piperideine.

Securinine is thus shown to be one of the piperidine alkaloids whose derivation from lysine avoids a symmetrical intermediate.<sup>4</sup> It is the first amongst this group of alkaloids whose nitrogen atom is common to two rings. All other lysine-derived alkaloids whose nitrogen lies at a ring junction (e.g., lupine,<sup>5</sup> decodon<sup>6</sup> and lycopodium<sup>7</sup> alkaloids) incorporate lysine by way of a symmetrical intermediate.

A further tracer experiment throws light on the origin of the nitrogen atom of securinine. This nitrogen might be supplied by the lysine-derived<sup>9</sup> or by the tyrosine-derived<sup>10</sup>

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<sup>3</sup> R. J. Parry, J.C.S. Chem. Comm., 1975, 144.

<sup>4</sup> E. Leistner and I. D. Spenser, *J. Amer. Chem. Soc.*, 1973, 95, 4715, and references therein.
<sup>5</sup> H. R. Schütte in 'Biosynthese der Alkaloide,' eds. K. Mothes and H. R. Schütte, VEB Deutscher Verlag der Wissenschaften, Berlin, 1969, p. 324. <sup>6</sup> S. H. Koo, R. N. Gupta, I. D. Spenser, and J. T. Wrobel, Chem. Comm., 1970, 396.

<sup>7</sup> M. Castillo, R. N. Gupta, D. B. MacLean, and I. D. Spenser, Canad. J. Chem., 1970, 48, 1893; Y. K. Ho, R. N. Gupta, D. B. MacLean, and I. D. Spenser, ibid., 1971, 49, 3352.

<sup>9</sup> M. Castillo, R. N. Gupta, Y. K. Ho, D. B. MacLean, and I. D. Spenser, *Canad. J. Chem.*, 1970, 48, 2911.
<sup>9</sup> R. B. Herbert in 'The Alkaloids,' ed. J. E. Saxton, Specialist Periodical Reports, The Chemical Society, London, 1975, Vol. 5, p. 10.
<sup>10</sup> E. Lecte in 'Biosynthesis,' ed. J. D. Bu'lock, Specialist Periodical Reports, The Chemical Society, London, 1976, Vol. 4, p. 97, and personal communication.

fragment. Administration to Securinega plants of a sample of  $[RS-6-^{3}H;6-^{14}C]$ -DL-lysine  $(^{3}H:^{14}C = 8\cdot 1 \pm 0\cdot 1)$ , prepared by mixing [RS-6-<sup>3</sup>H]-DL-lysine, 0.8 mCi, 21 Ci mmol<sup>-1</sup> and [6-14C]-DL-lysine, 0.1 mCi, 48 mCi mmol<sup>-1</sup>, both Commissariat á l'Ènergie Atomique, France) yielded securinine ( ${}^{3}H:{}^{14}C = 8 \cdot 0 \pm 0 \cdot 5$ ) which had retained all tritium, relative to 14C. This result indicates that the nitrogen atom of securinine is derived from the  $\epsilon$ -amino nitrogen of lysine. Derivation of the securinine nitrogen from any other source would require loss of the  $\epsilon$ -nitrogen of lysine along the route of biosynthesis of the alkaloid. Loss of the  $\epsilon$ -nitrogen of lysine, in turn, must be accompanied by labilization and loss of at least part of the tritium, relative to <sup>14</sup>C, from [RS-6-<sup>3</sup>H]lysine.

The present results, together with the data presented earlier<sup>1-3</sup> are consistent with the route to securinine shown in Scheme 2.

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