

## New Pterocarpinoid Phytoalexins of Soybean

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**Summary** Structures have been assigned to two new antifungal pterocarpinoids identified in cupric chloride-treated and pathogen-infected soybean plants.

SEEDLINGS of soybean [*Glycine max* (L.) Merr.] synthesise antifungal compounds in response to infection by *Phytoph-*

*thora megasperma* Drechs. var. *sojae* or on treatment with cupric chloride.<sup>1</sup> One phytoalexin has recently been assigned the revised structure (1).<sup>2</sup> We report the absolute configuration of this compound and the structures of two isomeric pterocarpinoid phytoalexins, produced in similar quantities by soybeans (var. Amsoy 71).

Metabolites extracted from cupric chloride-treated cotyledons were purified by chromatography on Sephadex LH-20 ( $\text{CHCl}_3$ ) and by h.p.l.c. on 10  $\mu\text{m}$  Partisil (propan-2-ol-hexane, 3:97). Three major antifungal components were isolated and characterised. The first compound ( $M^+$  338.1157) was identified by  $^1\text{H}$  n.m.r. spectroscopy (Table) as (1). On the basis of the large negative trough in its o.r.d. spectrum  $\{[\Phi]_{242}(\text{EtOH}) -100,000\}$  we assign the (6aR, 11aR) configuration<sup>3</sup> to this compound.

TABLE.  $^1\text{H}$  n.m.r. data for phytoalexins (1), (2), and (5).<sup>a</sup>

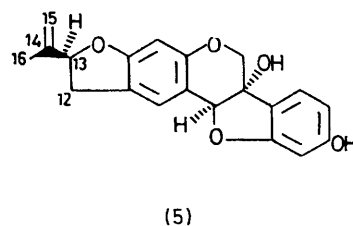
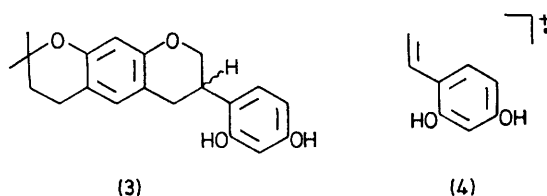
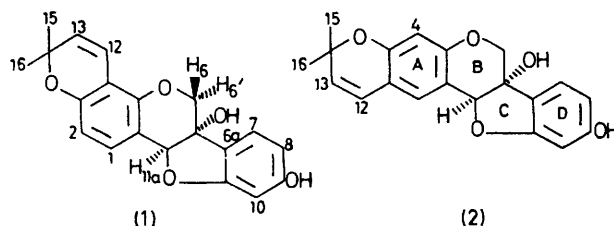
Proton	(1)		(2)		(5)	
	$\delta$	$J/\text{Hz}$	$\delta$	$J/\text{Hz}$	$\delta$	$J/\text{Hz}$
H-1	7.24d	8.5	7.15s	—	7.27s	—
H-2	6.47d	8.5	—	—	—	—
H-4	—	—	6.21s	—	6.27s	—
H-6	4.12d	12	4.04d	12	4.05d	11
H-6'	4.32d	12	4.13d	12	4.15d	11
H-7	7.22d	8	7.21d	8	7.23d	8
H-8	6.43q	8; 2	6.43q	8; 2	6.46q	8; 2
H-10	6.25d	2	6.25d	2	6.26d	2
H-11a	5.27s <sup>b</sup>	—	5.25s <sup>b</sup>	—	5.30s <sup>b</sup>	—
H-12	6.53d	10	6.41d	10	3.05 <sup>c</sup>	16; 8
					3.42 <sup>c</sup>	16; 9
H-13	5.65d	10	5.65d	10	5.26 <sup>c</sup>	8
H-15	1.36s <sup>d</sup>	—	1.37s <sup>d</sup>	—	4.91s <sup>b</sup>	—
					5.08s <sup>b</sup>	—
H-16	1.39s <sup>d</sup>	—	1.40s <sup>d</sup>	—	1.77s <sup>d</sup>	—

<sup>a</sup> Spectra measured in  $(\text{CD}_3)_2\text{CO}$  relative to internal reference  $\text{Me}_4\text{Si}$ , on a JEOL PFT-100 spectrometer at 99.54 MHz. <sup>b</sup> Broad singlet. <sup>c</sup> ABX system. <sup>d</sup> 3-Proton singlet.

The second compound crystallised from toluene {m.p. 89–93 °C;  $\text{C}_{20}\text{H}_{18}\text{O}_5$  ( $M^+$  338.1155);  $\lambda_{\text{max}}$  (EtOH) 227 ( $\epsilon$  38,500), 285 (8700), 307 (6200), and 318 (5800) nm;  $[\Phi]_{243}(\text{EtOH}) -78,000$ ;  $\nu_{\text{max}}$  (KBr) 3420, 3225, 1625, 1615, and 1585  $\text{cm}^{-1}$ } and was assigned structure (2). Comparison of spectra and the mass spectrometric fragmentation pattern with those of (1) suggested the presence of a 6a-hydroxypterocarpan ring system. This was confirmed by its ready conversion into an anhydro-derivative [ $M^+$  320;  $\lambda_{\text{max}}$  (EtOH) 232, 274, 348, and 366 nm] by formic acid treatment, and by the  $^1\text{H}$  n.m.r. spectrum of (2) (Table) which contains an AB quartet showing W-coupling of the low field doublet (H-6') to a benzylic proton (H-11a). Decoupling experiments defined the aromatic substitution pattern and the presence of a 2,2-dimethylpyran ring. The attachment of this heterocyclic system to ring A rather than to ring D was deduced from the isoflavan derivative (3), prepared by hydrogenation/hydrogenolysis of the anhydro-derivative of (2).<sup>2</sup> The  $^1\text{H}$  n.m.r. spectrum of (3) showed a shift of only the ABX aromatic system on formation of the phenoxide with KOH, while its mass spectrum contained the characteristic retro-Diels-Alder cleavage ion (4) ( $m/e$  136).<sup>4</sup>

The third phytoalexin crystallised from aqueous ethanol {m.p. 149–53 °C;  $\text{C}_{20}\text{H}_{18}\text{O}_5$  ( $M^+$  338.1146);  $\lambda_{\text{max}}$  (EtOH) 287 ( $\epsilon$  9400) and 292 (9600) nm;  $[\Phi]_{245}(\text{EtOH}) -79,000$ ;  $\nu_{\text{max}}$  (KBr) 3425, 3225, 1620, and 1610  $\text{cm}^{-1}$ }. Spectral

properties, dehydration, and conversion into the corresponding isoflavan established the molecule as a 2,3-substituted 6a,9-dihydroxypterocarpan. The structure of the side chain was deduced from the  $^1\text{H}$  n.m.r. spectrum (Table) which contained signals assigned to an ABX system containing *gem*-benzylic protons, a single methyl group attached to an  $sp^2$  carbon atom, and a 1,1-disubstituted olefin. The phytoalexin was thus assigned structure (5). The configuration at C-13 was determined by examination of the c.d. spectrum of the osmate esters at ca. 474 nm.<sup>5</sup> After treatment of (5) with  $\text{OsO}_4$ -pyridine, a positive Cotton effect was observed ( $[\theta]_{\text{max}} +3860^\circ$ ) whereas treatment of rotenone, which contains the same prenyl side-chain of defined *R* configuration,<sup>6</sup> resulted in a negative Cotton effect ( $[\theta]_{\text{max}} -5800^\circ$ ). Together with the negative trough in the o.r.d. spectrum this establishes the (6aR,11aR,13S) configuration of (5).



In a typical experiment the isomers (1), (2), and (5) were isolated in yields of 16, 7, and 17  $\mu\text{g/g}$  fresh weight of cotyledons. The three compounds are also produced in response to infection of hypocotyls by *P. megasperma* var. *sojae* and possess similar activity against this organism ( $\text{ED}_{50}$  ca. 60  $\mu\text{g ml}^{-1}$ ). It is interesting to note that no significant amount of the C-4 prenylated isomer of (5) has been detected.

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<sup>1</sup> N. T. Keen, J. J. Sims, D. C. Erwin, E. Rice, and J. E. Partridge, *Phytopathology*, 1971, **61**, 1084; W. L. Klarman and J. B. Sandford, *Life Sci.*, 1968, **7**, Part II, 1095; J. J. Sims, N. T. Keen, and V. K. Honwad, *Phytochemistry*, 1972, **11**, 827.

<sup>2</sup> R. S. Burden and J. A. Bailey, *Phytochemistry*, 1975, **14**, 1389.

<sup>3</sup> A. Pelter and P. I. Amenechi, *J. Chem. Soc. (C)*, 1969, 887; S. Ito, Y. Fujise, and A. Mori, *Chem. Comm.*, 1965, 595.

<sup>4</sup> A. Pelter, P. Stainton, and M. Barber, *J. Heterocyclic Chem.*, 1965, **2**, 262.

<sup>5</sup> L. J. Mulheim and G. Ryback, unpublished results.

<sup>6</sup> G. Büchi, L. Crombie, P. J. Godin, J. S. Kaltenbronn, K. S. Siddalingaiah, and D. A. Whiting, *J. Chem. Soc.*, 1961, 2843.