Biosynthesis of Vitamin B₁₂: Evidence from Double-labelling Studies (¹³CD₃) for Intact Incorporation of Seven Methyl Groups

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Summary The deuterium noise-decoupled ¹³C n.m.r. spectrum of dicyanocobalamin (5) biosynthesised from [methyl-¹³CD₃]methionine shows intact incorporation of seven methyl groups into the macrocycle of vitamin B_{12} without exchange of D with H of the medium.

EXPERIMENTS with labelled uro'gen-III (1) have shown¹⁻⁴ its enzymic conversion into cobyrinic acid (2), the corrin precursor⁵ of vitamin B_{12} (3). Current interest centres on the intermediates between uro'gen-III (1) and the corrin



(2) during which the following operations are needed: (a) introduction of seven methyl groups, one each at carbons 1, 2, 5, 7, 12 (*pro-R*), 15, and 17, all derived from methionine⁴ and those at C-7 and C-12 (*pro-R*) are transferred intact;⁶ (b) loss of C-20 from (1); (c) decarboxylation of the ring-c acetic acid side chain to give the *pro-S* methyl group at C-12 of (2); (d) redox changes; (e) insertion of cobalt.

The number of possible sequences between uro'gen-III (1) and cobyrinic acid (2) is thus enormous and it would be helpful to distinguish between processes for C-1 to C-19 bond formation in which the C-1 methyl group has temporarily been a methylidene residue having ring-A as *e.g.* (6) and those where it remains unaffected, ring-A *e.g.* as (7); for an elegant reductive corrin ring-closure *in vitro* involving a methylidene pyrrolenine (as 6) see ref. 7. It was not possible to examine the history of the C-1 methyl group by an earlier method⁶ because of the impracticability of isolating ring-A by degradation of cobester (4). However, the synthesis from ¹³CD₃I and homocysteine of [*methyl*-¹³CD₃]-methionine (90 atom % ¹³C; 98 atom % D) and its use for the biosynthesis of vitamin B₁₂ by *Propionibacterium shermanii* gave the desired information.

The vitamin B_{12} (3) isolated from this experiment was examined by ¹³C-n.m.r. spectroscopy as dicyanocobalamin

(5) using deuterium noise-decoupling and ¹H-lock. Under these conditions, ¹³CD₃ groups will give rise to singlets, ¹³CD₂H to doublets (*J ca.* 130 Hz), ¹³CDH₂ to triplets, and ¹³CH₃ to quartets. The observed spectrum (Figure) shows no splitting of any of the signals from ¹³C-enriched methyl groups by protium directly bonded to ¹³C. Long-range



 ^{1}H — ^{13}C couplings broaden the ^{13}C resonances, however, so two signals merge together at *ca*. 18.7 p.p.m.[†] It should be noted that the two signals at high field from the methyl groups at C-5 and C-15 are very sharp, the nearest ^{1}H -atom being four bonds away.

The ¹³C n.m.r. spectrum of cobester⁶ (4) derived from the ¹³C-enriched vitamin B_{12} was recorded in benzene under

[†] The deuterium substitution causes a small upfield shift of the ¹³C-signals.



FIGURE. Deuterium noise-decoupled ¹³C-Fourier transform n.m.r. spectrum (methyl region) of 13C-enriched dicyanocobalamin (5) biosynthesised from $[methyl-13CD_3]$ methionine. (0.20м in 0.1m KCN-H₂O, 84, 785 transients); chemical shifts downfield from Me₄Si.

the same decoupling conditions; it was in agreement with that above, though for computing reasons the quality was somewhat less.

The combined results show that (a) the seven methyl groups of vitamin B_{12} (3) derived from methionine are transferred intact and (b) none of them undergoes significant exchange of hydrogen with the medium during the biosynthesis of vitamin B_{12} (3). The intermediacy of a system with the C-1 methyl group temporarily as a methylidene residue (e.g. as 6) is thus unlikely since to satisfy the spectroscopic findings, deuterium removed from the C-1 methyl group would need to be returned to the same carbon atom without exchange with the medium.

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