

4-Nitrophenyl Phenyl Phosphorochloridate: A New Phosphorylating Agent for Oligonucleotide Synthesis

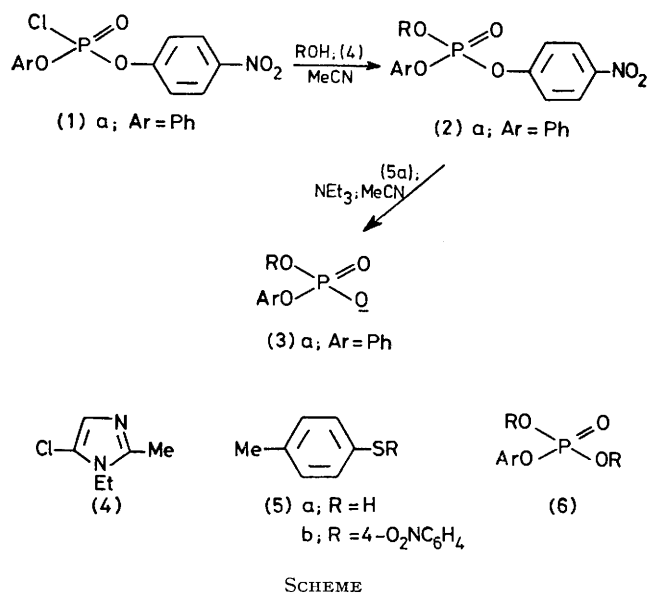
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Summary 2',5'-Protected ribonucleosides (7) and 5'-protected 2'-deoxyribonucleosides (8) readily react with 4-nitrophenyl phenyl phosphorochloridate (1a) in the presence of base to give good to high yields of phosphotriesters of general formula (2a) which, on treatment with *p*-thiocresol and triethylamine in acetonitrile are quantitatively converted into the corresponding phosphodiester (3a); the latter are intermediates in oligonucleotide synthesis by the phosphotriester approach.

ALTHOUGH the phosphotriester approach¹ is beginning to emerge as the method of choice for the chemical synthesis of oligo- and poly-nucleotides, several problems remain to be solved.² The most important of these is the development of a really satisfactory phosphorylation procedure. We now report that 4-nitrophenyl phenyl phosphorochloridate (1a) is an excellent phosphorylating agent for the first step of the phosphotriester approach.

The first step of the phosphotriester approach with an aryl protecting group involves the conversion of a protected nucleoside or oligonucleotide with a free 3'-hydroxy group (ROH) into the corresponding phosphodiester intermediate (3; Ar = aryl). In previous studies,^{1,2} like most other workers in this field, we have used a bifunctional phosphorylating agent such as an aryl phosphorodichloridate or a monoaryl phosphate in the presence of an arenosulphonyl chloride for this purpose. In the deoxy-series, this approach led to the formation of substantial quantities of symmetrical products² (6) in addition to the desired phosphodiester intermediates (3). A further disadvantage of the

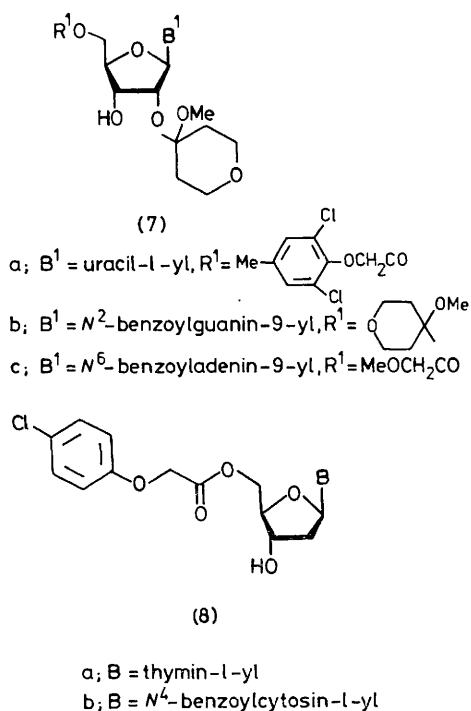


monoaryl phosphate procedure is that unacceptably low yields of products are obtained when the substrates contain guanine residues.

4-Nitrophenyl phenyl phosphorochloridate (1a), which may be obtained as a crystalline solid, in 65% isolated yield, from the products of the reaction between phenyl phosphoro-

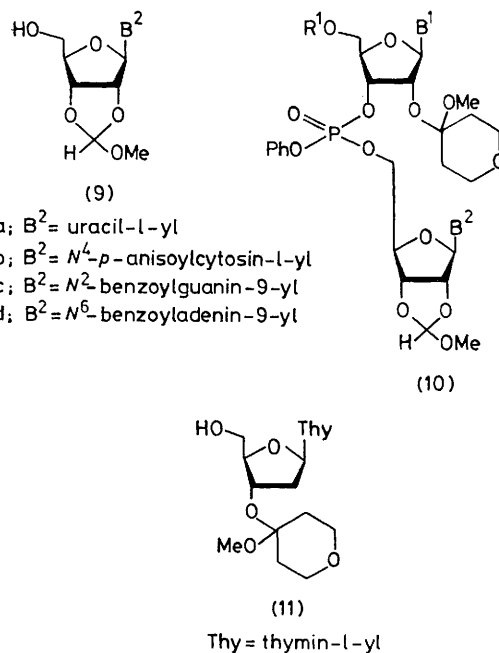
dichloridate and 4-nitrophenol† in the presence of a catalytic amount of 5-chloro-1-ethyl-2-methylimidazole³ (4), is a powerful phosphorylating agent. Thus the reactions (Scheme) between the relatively hindered ribonucleoside building blocks [ROH; (7a),⁴ (7b),⁵ and (7c)⁶] and 2-3-fold excesses of (1a) in the presence of (4) [3 mol. equiv. with respect to (1a)] in acetonitrile solution were complete within 3 h at 20 °C and the corresponding intermediate phosphotriesters (2a) were isolated as colourless solids, following short chromatography⁷ on silica gel, in 91, 92, and 82% yields, respectively. Under the same conditions, 5'-protected deoxyribonucleoside building blocks^{1,2} [ROH; (8a) and (8b)] underwent rapid phosphorylation and the corresponding intermediate phosphotriesters (2a) were isolated in 86 and 75% yields, respectively. The somewhat lower yields obtained in the deoxy-series are not significant as the preparations have not yet been optimized.

When the phosphotriesters (2a) were treated with 10 mol. equiv. of *p*-thiocresol (5a) and triethylamine in acetonitrile solution at 20 °C for *ca.* 1 h, they were quantitatively converted into (5b) and the triethylammonium salts of the corresponding phosphodiester intermediates (3a). The latter intermediates (3a) were isolated as pure colourless solids, free from (5a), (5b), and other impurities, by precipitation with cyclohexane from dichloromethane solution. Both the acid- and base-sensitive protecting groups are quite stable under these mild reaction conditions.



The phosphodiester intermediate (3a) derived from the uridine building block (7a) was treated with *ca.* 1.6 mol.

equiv. of 2,4,6-tri-isopropylbenzenesulphonyl chloride⁸ (TPS) in anhydrous pyridine solution at 20 °C followed, after 1 h, by *ca.* 1.2 mol. equiv. of each of the 2',3'-O-methoxymethylene derivatives⁹ (9a—d). After further periods of 16 h, the corresponding fully protected dinucleoside phosphates (10) [*i.e.* fully protected UpU, UpC, UpG, and UpA] were isolated, following short column chromatography,⁷ as pure solids in 85, 82, 70, and 81% yields, respectively. In the same way, the phosphodiester intermediate (3a) derived from the guanosine building block (7b) was converted into fully protected GpG (10; B¹ = B² = N²-benzoylguanin-9-yl, R¹ = 4-methoxytetrahydropyran-4-yl) in 72% yield and the intermediates (3a) derived from the deoxynucleoside building blocks [(8a) and (8b)] were treated first with TPS and then with (11)² to give fully protected TpT and dCpT in 96 and 92% isolated yields, respectively. In a preliminary experiment, the phosphodiester intermediate (3a) derived from the adenosine building block (7c) was condensed with



the partially protected dinucleoside phosphate (10; B¹ = uracil-1-yl, B² = N⁴-*p*-anisoylcytosin-1-yl, R¹ = H) to give fully protected ApUpC in 60% isolated yield. After complete removal of the protecting groups, all the products underwent total digestion in the presence of appropriate enzymes (ribonuclease A, ribonuclease T₁, or spleen phosphodiesterase) and were thereby shown to contain exclusively 3' → 5'-internucleotide linkages.

Other monofunctional phosphorylating agents have recently been suggested¹⁰ for the first step of the phospho-

† This is more convenient than the literature preparation (I. Dilaris and G. Eliopoulos, *J. Org. Chem.*, 1965, **30**, 686) of (1a) from 4-nitrophenyl phosphorodichloridate and phenol.

triester approach. Nevertheless, we believe that (**1a**) or a simple derivative of it, such as (**1**; Ar = 2-ClC₆H₄), will prove to be the most useful reagent for this purpose yet proposed.

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