O-Aryl S-Methyl Phosphorochloridothioates: Terminal Phosphorylating Agents in the Phosphotriester Approach to Oligonucleotide Synthesis

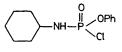
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Summary O-Aryl S-methyl phosphorothioates (e.g., 6), prepared by the action of O-aryl S-methyl phosphorochloridothioates (e.g., 4c) on alcohols may be converted into the corresponding phosphate esters (e.g., 7b) by treatment first with 4-nitrobenzaldoximate ion and then with iodine in aqueous pyridine.

THE procedures which have been devised so far for the synthesis of oligonucleotides by the phosphotriester approach¹ lead usually to products containing one more base-residue than phosphate ester group. A method which is suitable for the introduction of phosphate groups at the

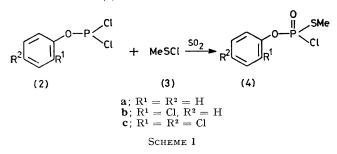
3'- and 5'-ends of oligonucleotide chains is therefore required. The use of aryl phosphoramidochloridates, such as phenyl phosphorocyclohexylamidochloridate (1), has



(1)

been recommended² for this purpose. However, the unblocking conditions then required² include an acidic hydrolysis step which makes the method unsuitable for

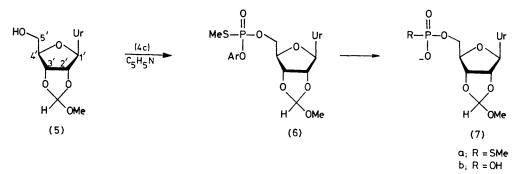
use in oligodeoxyribonucleotide synthesis. Although the acidic hydrolysis could possibly be replaced by a nitrosation step,³ we believe that a much better solution of this problem lies in the use of O-aryl S-methyl phosphorochloridothioates (4).



The latter reagents (4) may be readily prepared⁴ (Scheme 1) by adding methanesulphenyl chloride (3) to a solution of the appropriate aryl phosphorodichloridite (2) in sulphur dioxide at -67 °C. As methanesulphenyl chloride (3) may be prepared⁵ in situ by the action of sulphuryl chloride on commercially available dimethyl disulphide, *O*-aryl *S*-methyl phosphorochloridothioates (4) are easily accessible. Thus the 2,4-dichlorophenyl derivative (4c), which we consider to be a particularly suitable reagent for the present purpose, may be prepared from 2,4-dichlorophenyl phosphorodichloridite⁶ (2c) in 64% isolated yield.[†] The corresponding phenyl and 2-chlorophenyl derivatives (4a and 4b) have been prepared in 32 and 33% yields, respectively, by the same procedure.

The utility of (4c) as a terminal phosphorylating agent was first demonstrated with 2', 3'-OO-methoxymethyleneuridine⁷ (5) as the substrate. The reaction between (5) and 1.5 mol equiv. of (4c) in pyridine solution (Scheme 2) was complete within 30 min at 20 °C. The desired product (6) [³¹P n.m.r. ([²H₆]acetone-pyridine): δ 25.93 and 26.07 p.p.m.] was isolated pure in 74% yield. Treatment of (6) with 5 mol equiv. each of syn-4-nitrobenzaldoxime⁸ and triethylamine in dioxan-water (8:5 v/v) at 20 °C for 1 h gave (7a) [³¹P n.m.r. ([²H₆]acetone-pyridine): δ 16.75 p.p.m.] as the sole phosphorus-containing product. The purity of this material was confirmed by t.l.c. $(R_{\rm F} 0.77^{+})$, chromatography on DEAE-Sephadex A25, and paper electrophoresis (0.05 м phosphate buffer, pH 7.6). Treatment of (7a) with ca. 15 mol equiv. of iodine in pyridinewater (1:1 v/v) at 20 °C for 15 min gave 2',3'-OO-methoxymethyleneuridine 5'-phosphate (7b), $R_{\rm F}$ 0.44⁺ as virtually the sole nucleotide product. The latter compound (7b) was converted7 quantitatively into uridine 5'-phosphate $(R_F 0.23^{\dagger})$ by treatment first with dilute hydrochloric acid and then with dilute aqueous ammonia. The conversion of (7a) into (7b) follows the procedure developed by Cook et al.9 for the similar transformation of nucleoside S-ethyl phosphorothioates.

In the same way (Scheme 3), the partially-protected dinucleoside phosphate (8) was converted into the dinucleotide, pUpU (10). Phosphorylation of (8) with (4c) gave the fully-protected dinucleotide [³¹P n.m.r. ([²H₆]acetone-pyridine): $\delta - 7.24$ and 26.87 p.p.m.] corresponding to (6) in 76% isolated yield.§ Treatment of the latter product with 12.5 mol equiv. each of syn-4-nitrobenzaldoxime and $N^1N^1N^3N^3$ -tetramethylguanidine in dioxan-water (4:3 v/v) at 20 °C for 4 h gave (9a) [³¹P n.m.r. ([²H₆]acetone-pyridine): $\delta - 1.07$ and 17.69 p.p.m.; $R_F 0.67$;] which was isolated pure following chromatography on DEAE-Sephadex A25.¶ Treatment of (9a) with a twentyfold excess of iodine in aqueous pyridine gave (9b) as virtually the sole



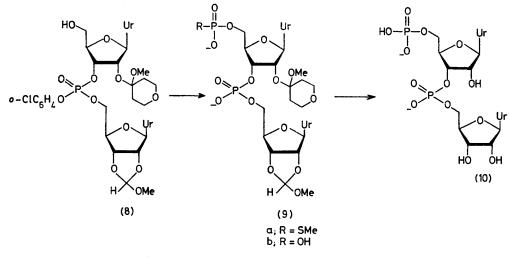
Scheme 2. Ur = Uracil-1-yl, $Ar = 2,4,-Cl_2C_6H_3$.

† It is particularly convenient to prepare (3) in one arm of an H-shaped apparatus and then pour it directly into (2) and SO_2 , contained in the other arm. Sulphuryl chloride (50 mmol) is added dropwise to dimethyl disulphide (50 mmol) at -67 °C (acetone-dry ice bath). The resulting crude orange (3) is added to a solution of (2c; 100 mmol) in sulphur dioxide (ca. 20 ml) also at -67 °C. The orange colour is immediately discharged and a pale green solution is obtained. The products are allowed to stand at -67 °C for 30 min and then allowed to warm up to room temperature. Distillation gives (4c; 18.8 g, 64%), b.p. 140–145 °C at 1.5 mmHg; ³¹P n.m.r. (CDCl₃): $\delta 34.71$ p.p.m.; ¹H n.m.r. (CDCl₃): $\delta 2.61$ (3H, d, J 20 Hz) and 7.1–7.5 (3H, m).

 \ddagger T.l.c. data are given for Merck DC-Alufolien Cellulose chromatograms developed (ascending) in the solvent system: EtOH-M-NH₄OAc (6:4 v/v).

\$ Yields of (6) and the phosphorylation product of (8) have not yet been optimized. If the phosphorylation reactions were carried out in acetonitrile solution in the presence of a 1-alkylimidazole catalyst, yields would very probably be much higher.

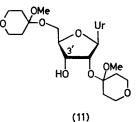
 \P Compound (9a) accounted for only 77% of the total nucleotide products eluted from the column. The DEAE-Sephadex chromatogram obtained suggested that a longer reaction time (say, 8–10 h) with 4-nitrobenzaldoximate ion is required for the complete unblocking of the 2-chlorophenyl-protected internucleotide linkage.



SCHEME 3. Ur = Uracil-1-yl.

nucleotide product. The desired dinucleotide, pUpU (10) $(R_F \ 0.16^{+})$ was obtained in almost quantitative yield by treating (9b) first with dilute hydrochloric acid (pH 2, 20 °C, 6 h) and then with dilute aqueous ammonia; this material was completely digested to uridine 5'-phosphate $(R_{\rm F} \ 0.25 \ddagger)$ in the presence of Crotalus adamanteus snake venom phosphodiesterase.

Preliminary experiments suggest that O-aryl S-methyl phosphorochloridothioates (4) are equally suitable for the phosphorylation of relatively hindered terminal 3'-hydroxy groups in oligoribonucleotide synthesis. Thus (4b) reacted readily with (11)¹⁰ and the product was unblocked by the reaction sequence described above to give uridine 3'phosphate.



Ur = Uracil-1-yl.

We thank the S.R.C. for generous financial support.

(Received, 21st August 1978; Com. 915.)

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