Stereochemistry of Cyclization in Carotenoid Biosynthesis; Use of ¹³C-Labelling to Elucidate the Stereochemical Behaviour of the C-1 Methyl Substituents during Zeaxanthin Biosynthesis in a *Flavobacterium*

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Summary [2-1³C]Mevalonate has been incorporated into the carotenoid zeaxanthin and its acyclic precursor, lycopene, in a *Flavobacterium* species; n.m.r. analysis showed ¹³C enrichment in both compounds at C-4, C-8, C-12, C-16, C-4', C-8', C-12', and C-16' so that in lycopene the C-1 methyl substituent *trans* to the main carbon chain was enriched and in zeaxanthin the 1α (axial) methyl substituent was labelled, enabling us to define the stereochemistry of the behaviour of the C-1 methyl groups during cyclization.

In order to define the stereochemistry of the cyclization reaction which gives the β -ring in carotenoid biosynthesis, two features have to be elucidated.¹ The first of these, the stereochemistry of hydrogen attack at C-2 of the acyclic precursor, was recently determined for zeaxanthin (3R,3'R- β,β -carotene-3,3'-diol) in a *Flavobacterium* species by use of deuterium labelling.² We now report the use of ¹³Clabelling, for the first time in studies of carotenoid biosynthesis, to determine the behaviour of the C-1 methyl substituents during the formation of zeaxanthin in *Flavobacterium*.

In the first of two experiments a Flavobacterium preparation was incubated with $[2-1^{3}C]$ mevalonate in the presence of nicotine (10 mm). Under these conditions cyclization is inhibited and the acyclic precursor lycopene (ψ , ψ -carotene) accumulates as the main carotenoid in place of the normal zeaxanthin. Examination of the lycopene (1) sample by ¹³C n.m.r. spectroscopy (67.89 MHz) showed that the ¹³C label (1-2% enrichment at each position) was localized at four positions, C-4 (δ 40·30 p.p.m.), C-8 (135·50), C-12 (137.43), and C-16 (25.67).³ These are exactly the positions predicted from a consideration of the general biosynthetic pathway. Of particular importance is the fact that the C-16 methyl group, *i.e.* that methyl substituent at C-1 which is trans to the main carbon chain of the lycopene molecule, was enriched with 13C, whereas the C-17 methyl group (δ 17.70 p.p.m.), which is *cis* to the main carbon



chain was not enriched. This result is significant in that (a) it confirms the isoprenoid labelling pattern in lycopene, (b) it shows that the two C-1 methyl substituents retain their individuality up to the lycopene stage of biosynthesis, and (c) it defines the positions of ¹³C-labelling in the acyclic precursor which normally undergoes cyclization to give β -carotene and thence zeaxanthin.

In the second experiment a *Flavobacterium* preparation was incubated with [2-1³C]mevalonate with no nicotine present. A ¹³C enrichment of 3—4% was achieved in the appropriate positions of the zeaxanthin (2) isolated. The ¹³C n.m.r. spectrum showed the ¹³C label again to be located in the C-4 (δ 42.76 p.p.m.), C-8 (138.56), and C-12 (137.66) positions, and in one of the methyl substituents at C-1 (δ 28.84 p.p.m.). ¹³C N.m.r.⁴ and ¹H n.m.r.² studies have shown that in (3*R*,3'*R*)-zeaxanthin the 3 β -hydroxy-group occupies the equatorial position so that the 3 α -proton is axial, the 2 β -proton is quasi-axial, and the 2 α -proton quasiequatorial. The β -methyl substituent at C-1 will therefore occupy the equatorial position and the α -methyl substituent will be axial. In the ¹³C n.m.r. spectrum of zeaxanthin separate signals are given by the two C-1 methyl substituents, at δ 28.84 and 30.35 p.p.m. In cyclohexane derivatives axial substituents resonate at higher field than the corresponding equatorial substituent.⁵ The signal at $\delta 30.35$ p.p.m. in the zeaxanthin spectrum can therefore be assigned to the equatorial (1β) substituent and that at δ 28.84 p.p.m. to the axial (1 α) methyl group. Studies with lanthanide shift reagents are in full agreement with this conclusion.

The ¹³C label in the *Flavobacterium* sample is thus located in the axial $l\alpha$ -methyl substituent. The behaviour of the C-1 methyl substituents during cyclization is therefore as illustrated in the Scheme. The stereochemistry of hydro-



SCHEME. Stereochemical course of the cyclization of lycopene to give zeaxanthin in Flavobacterium. $\dot{\bullet} = {}^{13}C.$

gen introduction at C-2, previously deduced from deuteriumlabelling experiments,² is also included. These two results together define the stereochemistry of the reaction at the

- ¹ G. Britton, Pure Appl. Chem., 1976, 47, 223.
 ² G. Britton, W. J. S. Lockley, N. J. Patel, T. W. Goodwin, and G. Englert, J.C.S. Chem. Comm., 1977, 655.
 ³ G. Englert, Helv. Chim. Acta, 1975, 58, 2367.

⁴ G. P. Moss, *Pure Appl. Chem.*, 1976, 47, 97.
⁵ W. McFarlane, in 'Elucidation of Organic Structures by Physical and Chemical Methods,' Pt. 1, 2nd Ed., eds. K. W. Bentley and G. W. Kirby, Wiley-Interscience, New York, 1972.

⁶ J. D. Bu'Lock, D. J. Austin, G. Snatzke, and L. Hruban, Chem. Comm., 1970, 255.

C-1-C-2 double bond (equivalent to an overall trans addition). It is still not possible to distinguish between the alternative chair (3) and boat (4) foldings, both of which would lead to the same labelling pattern.



It is interesting that the labelling pattern in the C-1 methyl groups of zeaxanthin in Flavobacterium is opposite to that deduced for trisporic acid and β -carotene in *Blakeslea* trispora.6

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