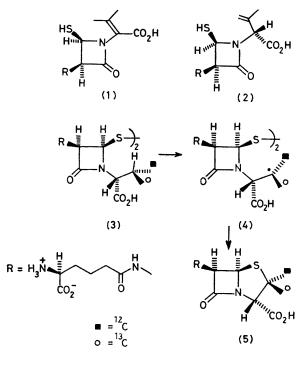
Penicillin Biosynthesis. A Model for Carbon-Sulphur Bond Formation

By JACK E. BALDWIN* and TERENCE S. WAN

(Dyson Perrins Laboratory, South Parks Road, Oxford OX1 3QY)

Summary Oxidative decarboxylation of bis(4-carboxy-4,4dimethylbutyl) disulphide yields 2,2-dimethyltetrahydrothiophen; this experiment is presented as a chemical model for the biosynthesis of the carbon-sulphur [C(2)-S(1)] bond of penicillins.

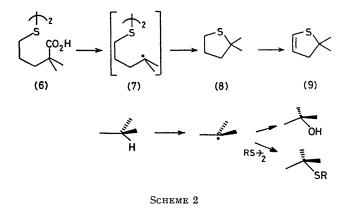
In spite of much effort, details of the nuclear biosynthesis of penicillins-cephalosporins remain obscure.¹ However, the incorporation of L-a-aminoadipyl-L-cysteinyl-D-[2-3H]valine into penicillin N, by an extract of Cephalosporiumacremonium,² suggests that dehydropeptides, such as (1) or (2), are not involved contrary to earlier speculations.³ Furthermore, the incorporation of [2Hg]valine into penicillin N with retention of all six deuterons,⁴ and the retention of valine C(3) stereochemistry,⁵ strongly limits the type of biomechanism for formation of the carbon-sulphur bond. We suggest a pathway which is in accord with these restrictions and is supported by in vitro experiments.



SCHEME 1

It seemed reasonable that dehydrogenation of the peptide (3) to the carbon radical (4) would result in intramolecular trapping by the disulphide linkage to yield the cyclised peptide (5) (Scheme 1). Provided that intramolecular trapping were faster than rotation in the radical (4) then this mechanism agrees with all known facts.

To evaluate the chemical feasibility of this mechanism, a model system was examined. Thus the disulphide (6), m.p. 54-55 °C, was prepared from isobutyric acid by sequential allylation (lithium di-isopropylamide, allyl bromide), anti-Markownikoff hydrobromination, thiourea displacement, followed by hydrolysis (NaOH) and oxidative coupling (I_2) . Oxidative decarboxylation of (6) [Pb(OAc)₄, tetrahydrofuran, 25 °C, 2 h] gave 2,2-dimethyltetrahydrothiophen (8) [27%, m.p. (HgCl₂ complex)⁶ 131-132 °C] and 2,2-dimethyl-2,3-dihydrothiophen (9) [15%, oil; δ (CDCl₃) 1.50 (s, 6H), 2.5 (m, 2H), 5.4 (m, 1H), and 6.1 (br d, 1 6 Hz, 1H)], separated by preparative g.l.c. (10% SE-30/Chrom. P.). The most reasonable interpretation of these observations is that the carbon radical (7), produced by oxidative decarboxylation of (6), is intercepted by the disulphide to yield (8), which is subsequently partially converted into (9). a reaction with ample precedence.7 A recent preparative observation by Maki and Sako,⁸ within a penicillin system. adds further support to the chemical basis of our hypothesis.



It might be reasonably asked what source of the carbon radical (4) might exist in the cell. One intriguing possibility derives from observations which suggest that the mechanism of hydroxylation enzymes has characteristics in accordance with carbon radical intermediates,⁹ which are normally converted into alcohols. However the close location of a disulphide might yield the corresponding sulphide, as a diversion of the normal pathway (Scheme 2).

(Received, 6th November 1978; Com. 1200.)

¹ For recent reviews, see: E. P. Abraham, J. Antibiotics, XXX Suppl., 1977, 1; D. J. Aberhart, Tetrahedron, 1977, 33, 1545. ² P. A. Fawcett, J. J. Usher, and E. P. Abraham, in The Proceedings of the Second International Symposium on the Genetics of Industrial Microorganisms, August 1974, Sheffield, England, ed. K. D. Macdonald, Academic Press, New York, 1976. ³ E.g., cf. J. E. Baldwin, S. B. Haber, and J. Kitchin, J.C.S. Chem. Comm., 1973, 790.

⁴ H. Kluender, F. C. Huang, A. Fritzberg, H. Schnoes, C. J. Sih, P. Fawcett, and E. P. Abraham, J. Amer. Chem. Soc., 1974, 96, 4054; D. J. Aberhart, J. Y. R. Chu, N. Neuss, C. H. Nash, J. Occolowitz, L. L. Huckstep, and N. De La Higuera, J.C.S. Chem. Comm., 1974, 564.

⁵ N. Neuss, C. H. Nash, J. E. Baldwin, P. A. Lemke, and J. B. Grutzner, *J. Amer. Chem. Soc.*, 1973, **95**, 3797. ⁶ Yu. K. Yurév, G. Ya. Kondratéva, and P. A. Akishin, *J. Gen. Chem.* (U.S.S.R.), 1953, **23**, 2059.

- ⁷ G. Sosnovsky, *Tetrahedron*, 1962, 18, 903.
 ⁸ Y. Maki and M. Sako, *J.C.S. Chem. Comm.*, 1978, 836.
 ⁹ J. T. Groves, G. A. McClusky, R. E. White, and M. J. Coon, *Biochem. Biophys. Res. Comm.*, 1978, 81, 154.