

## Structures of Mycinamicins

By MITSUO HAYASHI\*, MASARU OHNO, and SHUZO SATO

(Research Laboratories, Toyo Jozo Co Ltd, Ohito-cho, Shizuoka 410-23, Japan)

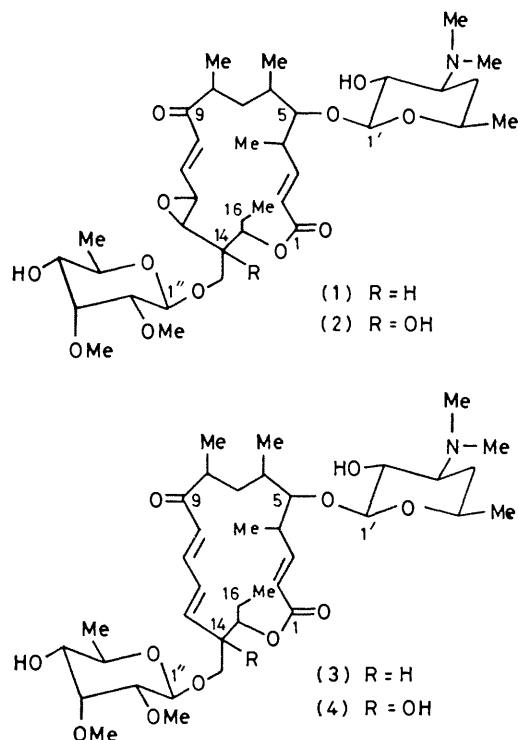
*Summary* A new family of basic 16-membered macrolide antibiotics with novel skeletons was isolated from *Micro-*  
*monospora sp* A-11725 and their structures were determined on the basis of physico-chemical properties.

MYCINAMICINS, a new group of macrolide antibiotics, have been obtained from *Micromonospora* sp. A-11725, and named as mycinamicin I (1), II (2), IV (3), and V (4),† respectively; they were characterized as in Table 1.

The mass spectral fragmentation pattern of mycinamicin II was very similar to that of mycinamicin I. The molecular ion ( $m/z$  727) and the fragment ions containing the aglycone unit ( $m/z$  553, 536, and 361) of II appeared at

TABLE 1. Physicochemical properties of mycinamicins.

Formula	I C <sub>37</sub> H <sub>61</sub> NO <sub>12</sub>	II C <sub>37</sub> H <sub>61</sub> NO <sub>13</sub>	IV C <sub>37</sub> H <sub>61</sub> NO <sub>11</sub>	V C <sub>37</sub> H <sub>61</sub> NO <sub>12</sub>
M.p./°C	103—107	102—106	174—176	148—150
[α] <sub>D</sub> <sup>25</sup> (c, 1.0, MeOH)	-40.0°	-31.0°	+2.7°	+18.7°
λ <sub>max</sub> /nm (MeOH)	218	218	215	215
(ε)	(23,700)	(23,200)	(20,700)	(20,800)
	240	240	281.5	280
	(sh., 12,900)	(sh., 12,200)	(21,500)	(21,400)
ν <sub>max</sub> /cm <sup>-1</sup> (KBr)	3470, 1715, 1685, 1645, 1620	3460, 1715, 1690, 1645, 1620	3460, 1715, 1675, 1645, 1625, 1590	3440, 1710, 1675, 1640, 1595



These data suggested that (1) and (2) resemble the neutral macrolide, chalomycin,<sup>1</sup> except in the aminosugar unit. Comparison of the <sup>13</sup>C n.m.r. spectra of mycinamicins with those of chalomycin and tylosin<sup>2</sup> afforded evidence for the presence of an aglycone unit with a novel skeleton in the mycinamicins. The data in Table 2 clearly demonstrate that the <sup>13</sup>C n.m.r. spectra of the aglycone part of (1) and of chalomycin were quite similar to each other except for the signals due to C-8, C-16, C-17, and 8-Me in (1). The <sup>1</sup>H n.m.r. spectra (in CD<sub>3</sub>COCD<sub>3</sub>) showed similar features for all the compounds [*i.e.*, resonances in the ranges δ 2.28—2.31 (s, NMe<sub>2</sub>), 4.29—4.32 (d, *J* 7.3 Hz), 3.49—3.55 (2 × OMe), and 4.57—4.60 (d, *J* 7.8 Hz)]. These <sup>1</sup>H and <sup>13</sup>C n.m.r. spectra suggested that desosamine and mycinose comprise the sugar units in mycinamicin and accordingly structure (1) was proposed for mycinamicin I.

16 a.m.u. (oxygen atom) higher than the corresponding ions ( $m/z$  711, 537, 520, and 345) of I, suggesting that II had an additional oxygen in the aglycone structure. Furthermore, in comparing the <sup>1</sup>H n.m.r. spectrum of II with that of I, it became apparent that the splitting pattern

TABLE 2. <sup>13</sup>C n.m.r. chemical shifts for mycinamicin I<sup>a</sup>, II<sup>a</sup>, IV<sup>a</sup>, V<sup>a</sup>, chalomycin (A), and tylosin (B)

	I	II	IV	V	(A)	(B)
C-1	165.7	165.9	166.1	166.3	164.7	173.9
C-2	120.1	120.0	120.9	120.7	120.3	39.4
C-3	151.5	151.9	151.6	151.8	151.0	71.7
C-4	41.9	42.0	41.3	41.3	41.5	45.1
C-5	87.5	87.5	87.9	87.7	87.5	81.6
C-6	34.2	34.4	34.1	34.1	36.7	32.3 <sup>b</sup>
C-7	32.1	32.0	32.6	32.7	33.9	32.9 <sup>b</sup>
C-8	44.7	44.6	44.9	44.8	78.4	40.3
C-9	200.8	200.8	203.4	203.8	199.4	202.8 <sup>c</sup>
C-10	125.6	126.2	123.2	123.8	124.5	118.8
C-11	143.7	143.0	141.7	141.4	145.8	148.0
C-12	59.6 <sup>b</sup>	60.4	133.0	130.5	59.4 <sup>b</sup>	134.9
C-13	59.0 <sup>b</sup>	54.0	141.3	143.5	58.8 <sup>b</sup>	142.2
C-14	47.5	73.1	49.2	77.4	49.4	44.7
C-15	72.4	74.2	72.7	75.8	68.5	75.3
C-16	24.7	21.2	25.3	21.4	18.3 <sup>c</sup>	25.5
C-17	8.9	10.1	9.6	10.4	—	9.0 <sup>d</sup>
4-Me	18.9 <sup>c</sup>	19.0 <sup>b</sup>	19.4 <sup>b</sup>	19.5 <sup>b</sup>	19.1 <sup>c</sup>	9.6 <sup>d</sup>
6-Me	17.1 <sup>c</sup>	17.1 <sup>b</sup>	17.4 <sup>b</sup>	17.4 <sup>b</sup>	18.5 <sup>c</sup>	—
6-CH <sub>2</sub>	—	—	—	—	—	43.9
8-Me	17.5 <sup>c</sup>	17.4 <sup>b</sup>	17.8 <sup>b</sup>	17.6 <sup>b</sup>	27.7	17.4 <sup>e</sup>
12-Me	—	—	—	—	—	13.0
14-CH <sub>2</sub>	67.1	72.6	68.6	75.2	66.7	68.2
CHO	—	—	—	—	—	203.0 <sup>c</sup>
C-1'	105.5	105.0	104.9	104.8	—	—
C-2'	70.3	70.3	70.4	70.4	—	—
C-3'	65.8	65.8	65.8	65.8	—	—
C-4'	28.5	28.5	28.3	28.4	—	—
C-5'	69.4	69.4	69.5	69.4	—	—
C-6'	21.1	21.2	21.2	21.2	—	—
NMe <sub>2</sub>	40.1	40.2	40.2	40.2	—	—
C-1''	100.9	101.2	101.0	101.6	100.5	101.1
C-2''	81.9	81.9	81.9	81.7	81.6	82.0
C-3''	79.3	79.3	79.9	79.2	79.4	79.9
C-4''	72.7	72.6	72.7	72.5	72.5	72.9
C-5''	70.6	70.7	70.5	70.7	70.4	70.6
C-6''	17.8 <sup>c</sup>	17.7 <sup>b</sup>	17.8 <sup>b</sup>	17.6 <sup>b</sup>	17.7 <sup>b</sup>	17.8 <sup>e</sup>
2''-OMe	59.0	59.3	59.7	59.1	58.5	59.6
3''-OMe	61.6	61.6	61.7	61.7	61.4	61.7

<sup>a</sup> <sup>13</sup>C n.m.r. spectra were recorded in CDCl<sub>3</sub> solution on a JEOL FX-100 Fourier transform spectrometer. Chemical shifts are given in p.p.m. with respect to Me<sub>4</sub>Si as internal standard. <sup>b-e</sup> Assignments within any vertical column may be reversed.

† Formerly named A 11725 I, II, Ia and IIa, respectively.

(dd) at  $\delta$  5.42 (H-15) in II was different from the splitting pattern (ddd) at  $\delta$  5.37 (H-15) in I. These facts indicated that II had an additional hydroxy group in the aglycone unit. In the  $^{13}\text{C}$  n.m.r. spectrum of II, a downfield shift of C-14 (47.5  $\rightarrow$  73.1 p.p.m.) was observed. Therefore, this additional hydroxy group was finally determined to be located at position 14 [structure (2)].

In order to investigate the structural relationships of mycinamicin I and II to IV and V, we studied their de-

epoxidation ( $\text{CrCl}_2$ -acetic acid)<sup>3</sup>. Reaction products from I (1) and II (2) are completely identical with natural mycinamicin IV (3) and V (4) in all respects. Details of these isolation<sup>4</sup> and structural<sup>5</sup> studies will be published elsewhere.

We thank Drs. M. Suzuki and K. I. Harada of Meijo University for their helpful advice.

(Received, 22nd October 1979, Com 1122)

<sup>1</sup> P. W. K. Woo, H. W. Dion, and Q. R. Bartz, *J. Amer. Chem. Soc.* 1964, **86**, 2724.

<sup>2</sup> S. Omura, A. Nakagawa, A. Neszmelyi, S. D. Gero, A. M. Sepulchre, F. Piriou, and G. Lukacs, *J. Amer. Chem. Soc.*, 1975, **97**, 4001.

<sup>3</sup> M. L. Kuehne and B. W. Benson, *J. Amer. Chem. Soc.* 1965, **87**, 4660.

<sup>4</sup> S. Sato, N. Mutō, M. Hayashi, M. Otani, and T. Fujii, *J. Antibiotics*, 1979, **32**, in the press.

<sup>5</sup> M. Hayashi, M. Ohno, S. Sato, K. I. Harada, and M. Suzuki, *J. Antibiotics*, in the press.