

A Hydroperoxy-epidioxide from the Autoxidation of a Hydroperoxide of Methyl Linolenate

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Summary Unlike autoxidation of linolenic acid or methyl linolenate which yields mixtures of hydroperoxy-epidioxides, sequential oxidation employing an enzymic first

step followed by an autoxidation step can give rise to a single hydroperoxy-epidioxide

OXIDATION of unsaturated lipids containing more than two double bonds, *e.g.* methyl linolenate, can proceed readily with the incorporation of two molecules of oxygen to yield hydroperoxy-epidioxides. Previous investigations^{1,2} using the fatty acid or its ester as the starting material resulted in complex mixtures of hydroperoxy-epidioxides. Whereas methods are available for the analysis of the mixtures of hydroperoxides formed³ and for the preparation of a single hydroperoxide,⁴ procedures for the preparation and analysis of individual hydroperoxy-epidioxides have not hitherto been devised. A method has been developed in this study whereby a single hydroperoxide isomer derived from the enzymic oxidation of linolenic acid was autoxidised to yield a hydroperoxy-epidioxide which was purified and analysed by h.p.l.c.

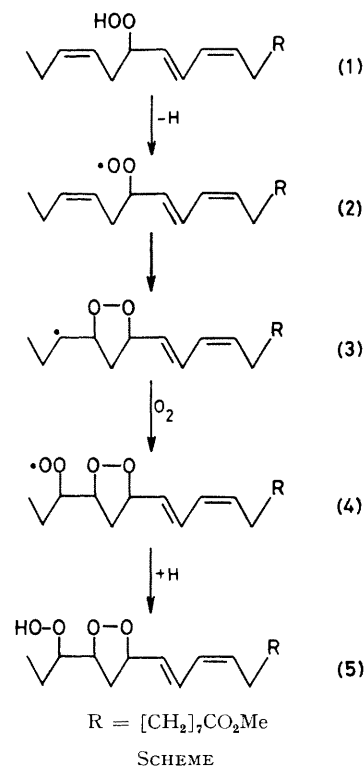
The hydroperoxide (1) (isomeric purity 97%; 5.7 $\mu\text{mol ml}^{-1}$ in hexane) prepared by methylation (CH_2N_2) of the hydroperoxyacid obtained from the lipoxygenase-catalysed oxidation⁴ of linolenic acid was heated (40 °C) in air for 96 h to yield a mixture containing (1) (25.4%), (5) (31.5%), and a mixture of polar products (43.0%). Preparative h.p.l.c.³ yielded (5) (λ_{max} 233 nm; ν_{max} 952, 988, and 1745 cm^{-1} ; *cis-trans* conjugated diene and COOR; peroxide to diene ratio³ 1.71). Reduction (NaBH_4 followed by Pt/H_2) of (5) yielded methyl 13,15,16-trihydroxystearate {mass spectrum of tri- Me_3Si derivative after g.c. contained $m/e = 131$ (EtCHOSiMe_3) and 315 ($\text{Me}_3\text{SiOCH}[\text{CH}_2]_{11}\text{CO}_2\text{Me}$)}. Spin decoupling experiments permitted the assignment of nine ^1H multiplets (Table) in the 360 MHz spectrum of (5) (CDCl_3) and confirmed the *cis*-9 ($J_{9,10}$ 10 Hz) and the *trans*-11 ($J_{11,12}$ 15 Hz) double bonds and the presence of the 5-membered cyclic peroxide ring.

TABLE. ^1H N.m.r. spectrum (360 MHz) of (5).

δ	Multiplicity	J/Hz	Assignment
2.47	octet ^a	12, 8, 5	14- H_A
2.84	octet ^a	12, 8, 9	14- H_B
4.15	octet ^a	9, 7, 3	16-H
4.49	octet ^a	9, 7, 5	15-H
4.80	q ^b	8, 8, 8	13-H
5.55	m ^c	10	9-H
5.62	q	15, 8	12-H
6.01	br t ^b	11, 10	10-H
6.67	q	15, 11	11-H

^a Partially resolved. ^b Apparent. ^c Unresolved.

Formation of (5) from (1) is consistent with the mechanism outlined in the Scheme in which H-abstraction from (1) by (4) constitutes a propagation step of a chain. The steps involved resemble those of autoxidation but with the incorporation of a cyclisation step (2) \rightarrow (3). Cyclisation of peroxy radicals has been used to produce prostaglandin-like products⁵ and the formation of peroxy radicals from unsaturated fatty acid hydroperoxides has been implicated



by the exchange of the peroxidic oxygen atoms with molecular oxygen in the rearrangement of the hydroperoxides.⁶ In the formation of (5) from (1), the intermediacy of the peroxy radical (2) was indicated by the enrichment in ^{18}O (26.1 atom %) in the oxygen atom at C-13 in (5) when the reaction was carried out in $^{18}\text{O}_2$ (70.4 atom %).

The formation of the mixture of hydroperoxy-epidioxides by the autoxidation of methyl linolenate has been assumed to proceed *via* the cyclisation of the peroxy radical (2) formed during autoxidation.¹ The isolation of a hydroperoxy-epidioxide from a hydroperoxide represents a new approach and is based on the previous observation⁶ that hydroperoxides give rise to peroxy radicals which undergo further radical chain reactions. By carrying out the two oxygenation reactions as distinct steps, each with a single starting material, a hydroperoxy-epidioxide has been obtained to enable detailed examination of its properties.

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¹ P. H. Begemann, W. J. Woestenburger, and S. Leer, *J. Agric. Food Chem.*, 1968, **16**, 679.

² M. Roza and A. Francke, *Biochim. Biophys. Acta*, 1978, **528**, 119.

³ H. W.-S. Chan and G. Levett, *Lipids*, 1977, **12**, 99, and 837.

⁴ J. A. Matthew, H. W.-S. Chan, and T. Galliard, *Lipids*, 1977, **12**, 324; T. Galliard and J. A. Matthew, *Biochim. Biophys. Acta*, 1975, **398**, 1.

⁵ N. A. Porter and M. O. Funk, *J. Org. Chem.*, 1975, **24**, 3614.

⁶ H. W.-S. Chan, G. Levett, and J. A. Matthew, *J.C.S. Chem. Comm.*, 1978, 756; *Chem. and Phys. Lipids*, 1979, **24**, 245.